

Effect of different inoculum level of *Meloidogyne incognita* on chlorophyll content of *Cajanus cajan*

Abstract

The pot culture experiment was conducted during *Kharif* season in the year 2021- 2022 at Green polyhouse, Department of Nematology, College of Agriculture, Odisha University of Agriculture and Technology, Bhubaneswar, Odisha, in order to study effect of different inoculum level of *M. incognita* on chlorophyll content of pigeonpea. The experiment was laid out in Complete randomized design (CRD) with 5 treatments i.e T₁ (500 J₂/plant), T₂(1000J₂ / Plant), T₃(1500J₂/plant), T₄(2000J₂/plant), T₅(Control) and 4 varieties were UPAS-120(R), IPA-15-1 (MR), IPA 14-7(S), CO-6(HS). With increase in inoculum level there was decrease in chlorophyll content. Highest reduction in chlorophyll in treatment T₄ then T₃, T₂ and then T₁ over control T₅. The results have demonstrated that nematode infestation leads to highest decreased by 41.75% total chlorophyll content (a+b) in UPAS -120(R)) in the leaves of the Pigeonpea plants.

Introduction

Pigeon pea *Cajanus cajan* L. is considered as one of the most important pulse crop grown in Indian subcontinent. It is the second most important pulse crop after gram. It is a good source of protein (20-23%), dietary fiber, and various vitamins: thiamin, magnesium, phosphorus, potassium, copper, and manganese. India ranked first in the world with 79.65% and 67.28% of world's acreage and production respectively. Globally it is cultivated on 4.9 m ha of which India alone occupies 3.5 m ha i. e. 72% of the total area[13]. This crop is highly vulnerable to many plant parasitic nematodes, which cause an annual yield loss of over 13% worldwide [12]. Root knot nematode, *Meloidogyne incognita*, is the most important nematode species with worldwide distribution in tropical and subtropical climate. The pathogenic effect of root-knot nematodes on growth parameters, yield and nutrient uptake of leguminous crops have been reported by several workers and it is documented as potential threat to various leguminous plants. Reduction in chlorophyll content of infected plant has been reported. [9][10][11][15]

Keywords: Pigeon pea (*Cajanus cajan*), Root knot nematode (*M. incognita*), chlorophyll

Material and Methods

In order to understand the basics of resistance to nematode (*Meloidogyne incognita*) inoculated four varieties UPAS-120 (Resistant), IPA-15-1 (Moderately resistant), IPA-14-7 (Susceptible) and CO-6 (Highly susceptible) were grown in pots filled with aerated sterilized soil (autoclaved at 1.1kg/cm² pressure for one hour daily for two consecutive days) mixed with sand and FYM in the ratio of 2:1:1 following Complete Randomized Design (CRD) with five treatments. The water used for irrigation had a five hundred mesh screen before use. Two weeks after seedling emergence aged nematodes were counted under a stereoscopic microscope and released into the holes @ 500 J2, 1000 J2, 1500 J2, 2000 J2 per seedling and one control. For chemical analysis three sets of plants were maintained. Each set was arranged on separate platform in the green house in order to avoid cross infection. At 30 days after inoculation, inoculated plants were removed from the pot soil carefully and the chlorophyll compositions were estimated.

One hundred fifty mg leaf portion of each treatment were cut from the composite leaves and were immersed in 50 ml of 80 % acetone in a conical flask and kept in dark for 24 hours for extraction of chlorophyll from the leaf samples. Thereafter, the chlorophyll extracts were filtered through Whatman No.1 filter paper. Absorbance of the chlorophyll extract was measured at 645 nm and 663 nm using a colorimeter. The amount of chlorophyll-a, chlorophyll-b and total chlorophyll were calculated in mg/g fresh weight according to the following equations (Anon, 1949).

i) Chlorophyll -a (mg/g fresh weight of leaf)

$$= 12.7 \times (D-663) - 2.69 \times (D-645) \times \frac{V}{1000 \times W}$$

ii) Chlorophyll-b (mg/g fresh weight of leaf)

$$= 22.9 \times (D-645) - 4.68 \times (D-663) \times \frac{V}{1000 \times W}$$

iii) Total chlorophyll (mg/g fresh wt. of leaf)

$$= 20.2 \times (D-645) + 8.02 \times (D-663) \times \frac{V}{1000 \times W}$$

Where, D -645 = optical density at 645 nm

D-663= optical density at 663 nm

V = final volume of 80 % acetone chlorophyll extract in ml

W = Fresh weight in gram of corresponding amount of fresh leaves used in the extraction of chlorophyll.

Result and Discussion

Chlorophyll content is the most important constituent of the plants as it manufactures the food, which is necessary for the growth and development of the plant. It is directly correlated with the yield of the crops. Root-knot nematodes are known to reduce the chlorophyll content of plants by disrupting its nutrient uptake and partitioning of the photosynthates.

It is clear from data presented in the Table -1 in variety UPAS-120(R) that chlorophyll-a was found 1.34, 1.17, 1.03, 0.86 mg/g against 1.47 mg/g; chlorophyll-b was found 1.35, 1.19, 1.06, 0.88 mg/g against 1.51 mg/g; total chlorophyll was found 2.65, 2.35, 2.06, 1.73 mg/g against 2.97 mg/g in the experiment plant inoculated with 500, 1000, 1500, 2000 J₂ /plant respectively. The total chlorophyll content was reduced by 10.89, 20.88, 30.64, 41.75% in 500, 1000, 1500, 2000 J₂ /plant respectively over control.

It is clear from data presented in the Table -2 in variety IPA -15-1(MR) that chlorophyll-a was found 1.41, 1.25, 1.12, 0.96 mg/g against 1.53 mg/g; chlorophyll-b was found 1.42, 1.31, 1.24, 1.05 mg/g against 1.58 mg/g; total chlorophyll was found 2.82, 2.63, 2.24, 1.77 mg/g against 3.10 mg/g in the experiment plant inoculated with 500, 1000, 1500, 2000 J₂ /plant respectively. The total chlorophyll content was reduced by 9.13, 15.15, 27.93, 43.07% in 500, 1000, 1500, 2000 J₂ /plant respectively over control.

It is clear from data presented in the Table -3 in variety IPA-14-7(S) that chlorophyll-a was found 1.50, 1.37, 1.24, 1.10 mg/g against 1.61 mg/g; chlorophyll-b was found 1.61, 1.49, 1.27, 1.15 mg/g against 1.72 mg/g; total chlorophyll was found 3.11, 2.86, 2.50, 2.24 mg/g against 3.31 mg/g in the experiment plant inoculated with 500, 1000, 1500, 2000 J₂ /plant respectively. The total chlorophyll content was reduced by 6.04, 13.70, 24.37, 32.23% in 500, 1000, 1500, 2000 J₂ /plant respectively over control.

It is clear from data presented in the Table -4 in variety CO-6(HS) that chlorophyll-a was found 1.59, 1.46, 1.34, 1.21 mg/g against 1.69 mg/g; chlorophyll-b was found 1.62, 1.59, 1.37, 1.26 mg/g against 1.79 mg/g; total chlorophyll was found 3.20, 3.01, 2.70, 2.47 mg/g against 3.47 mg/g in the experiment plant inoculated with 500, 1000, 1500, 2000 J₂ /plant respectively. The total chlorophyll content was reduced by -7.77, 13.34, 22.17, 28.79% in 500, 1000, 1500, 2000 J₂ /plant respectively over control.

Leaf chlorophyll content provides a measure of photosynthetic capacity and is related to the nitrogen concentration in the plant [4] which *M. incognita* can influence by interfering with water and nutrient transport [3][6][8]. Therefore, because chlorophyll content is affected by nitrogen concentration, it can be an indicator of the damage caused to the plant by *M. incognita*. Previous studies have shown that infection of plants by *M. incognita* can result in reduced chlorophyll content and photosynthesis [5][7]. Decrease in chlorophyll content with increase in inoculum level [1][2].

Table-1 Effect of different inoculum level of *M. incognita* on chlorophyll content (var. UPAS-120)

Treatments	Chlorophyll- a	% change over control	Chlorophyll-b	% change over control	Total Chlorophyll	% change over control
T ₁ 500J ₂	1.34	-8.78	1.35	-10.60	2.65	-10.89
T ₂ 1000J ₂	1.17	-20.15	1.19	-21.19	2.35	-20.88
T ₃ 1500J ₂	1.03	-29.71	1.06	-29.80	2.06	-30.64
T ₄ 2000J ₂	0.86	-41.31	0.88	-41.94	1.73	-41.75
T ₅ (Control)	1.47		1.51		2.97	
SE(m)±	0.17		0.09		0.12	
CD(0.05)	0.05		0.03		0.04	

Table-2 Effect of different inoculum level of *M. incognita* on chlorophyll content (var. IPA-15-1)

Treatments	Chlorophyll-a	% change over control	Chlorophyll-b	% change over control	Total Chlorophyll	% change over control
T ₁ 500J ₂	1.41	-8.06	1.42	-10.34	2.82	-9.13
T ₂ 1000J ₂	1.25	-18.30	1.31	-17.09	2.63	-15.15
T ₃ 1500J ₂	1.12	-27.02	1.24	-21.73	2.24	-27.93
T ₄ 2000J ₂	0.96	-37.25	1.05	-33.54	1.77	-43.07
T ₅ (Control)	1.53		1.58		3.10	
SE(m)±	0.16		0.06		0.18	
CD(0.05)	0.05		0.02		0.06	

Table-3 Effect of different inoculum level of *M. incognita* on chlorophyll content (var. IPA-14-7)

Treatments	Chlorophyll- a	% change over control	Chlorophyll-b	% change over control	Total Chlorophyll	% change over control
T ₁ 500J ₂	1.50	-7.02	1.61	-6.58	3.11	-6.04
T ₂ 1000J ₂	1.37	-15.29	1.49	-13.35	2.86	-13.70
T ₃ 1500J ₂	1.24	-23.14	1.27	-26.11	2.50	-24.37
T ₄ 2000J ₂	1.10	-31.61	1.15	-33.08	2.24	-32.23
T ₅ (Control)	1.61		1.72		3.31	
SE(m)±	0.16		0.07		0.16	
CD(0.05)	0.05		0.02		0.05	

Table-4 Effect of different inoculum level of *M. incognita* on chlorophyll content (var. CO-6)

Treatments	Chlorophyll- a	% change over control	Chlorophyll-b	% change over control	Total Chlorophyll	% change over control
T ₁ 500J ₂	1.59	-5.53	1.62	-9.14	3.20	-7.77
T ₂ 1000J ₂	1.46	-13.24	1.59	-10.82	3.01	-13.34
T ₃ 1500J ₂	1.34	-20.55	1.37	-23.32	2.70	-22.17
T ₄ 2000J ₂	1.21	-28.06	1.26	-29.29	2.47	-28.79
T ₅ (Control)	1.69		1.79		3.47	
SE(m)±	0.15		0.09		0.12	
CD(0.05)	0.05		0.03		0.04	

Conclusion

In conclusion, the present study suggested that clearly indicated that *Meloidogyne incognita* played key role in altering the normal physiology of the tested host plant. Further, Basic studies relating to physiology mechanism of resistance in pigeonpea to the Root knot nematode made elucidate the physiology basis of resistance to host to the nematode observation were made in the changes in physiology parameters. Chlorophyll a, b and total chlorophyll content decreases in leaves of infected plants as compared to control.

References

1. Abbasi and Hisamuddin, 2014. Effect of Different Inoculum Levels of *Meloidogyne incognita* on Growth and Biochemical Parameters of *Vigna radiata*. *Asian Journal of Nematology*, 3: 15-20.
2. Amal, TC., Karthika, P., Balamurugan, V., Shanmugam, G., Selvakumar, S., Sundararaj, P. and Vasanth, K. (2020). Effects of Root-Knot Nematode Inoculums Densities on Morphological and Phytochemical Analysis of Selected Horse Gram Germplasm. *Indian Journal of Agricultural Research*. 54(6): 708-715.

3. Carneiro RG, Mazzafera P, Ferra LCCB, Muraoka T, Trivelin PCO. Uptake and translocation of nitrogen, phosphorus and calcium in soybean infected with *Meloidogyne incognita* and *M. javanica*. *Fitopatologia Brasileira*. 2002; 27:141–150.
4. Evans JR. Photosynthesis and nitrogen relationships in leaves of C3 plants. *Oecologia*. 1989; 78:9–19.
5. Haseeb A, Srivastava NK, Pandey R. The influence of *Meloidogyne incognita* on growth, physiology, nutrient concentration and alkaloid yield of *Hyoscyamus niger*. *Nematologia Mediterranea*. 1990; 18:127–129.
6. Kirkpatrick TL, Oosterhuis DM, Wullschlegel SD. Interaction of *Meloidogyne incognita* and water stress in two cotton cultivars. *Journal of Nematology*. 1991; 23:462–467.
7. Loveys BR, Bird AF. The influence of nematodes on photosynthesis in tomato plants. *Physiological Plant Pathology*. 1973; 3:525–529.
8. Melakeberhan H, Webster JM, Brooke RC, D'Auria JM, Cackette M. Effect of *Meloidogyne incognita* on plant nutrient concentration and its influence on the physiology of beans. *Journal of Nematology*. 1987; 19:324–330.
9. Poornima, K. and S. Vadivelu, 1998. Parthenogenicity of *Meloidogyne incognita* to turmeric (*Curcuma longa* L.). Proceedings of the 3rd international symposium of Afro-Asian Society of Nematologists, April 16-19, 1998, Coimbatore, pp; 29-31.
10. Parveen, K., A. Haseeb and P.K.I. Shukla, 2006. Pathogenic potential of *Meloidogyne incognita* on *Mentha arvensis* cv. Gomti. *Ind. J. Nematol.*, 36: 177-180.
11. Ramakrishnan, S. and G. Rajendran, 1998. Effect of individual and concomitant initial inoculum of *Meloidogyne incognita* and *Rotylenchulus reniformis* on growth, physiology and nutrient content of papaya (*Carica papaya* L.). Proceedings of the 3rd International Symposium of Afro-Asian Society of Nematologists, April 16-19, 1998, Coimbatore, pp: 17-28.
12. Sasser JN & Freckman DW, A World Perspective on Nematology: The Role of the Society. In: *Vistas on Nematology* (Eds. JA Veech & DW Dickson Hyattsville, Maryland) 1987, 7.
13. Saxena K B 2009. Evaluation of Hybrid Breeding Technology in pigeonpea. In: *Milestones in Food Legume Research* (eds. Ali, M. and Kumar, S.). *Indian Journal of Pulses Research*, Kanpur. Pp 82-11.
14. Swain, B. and J.S. Prasad, 1988. Chlorophyll content in rice as influenced by the root-knot nematode, *Meloidogyne graminicola* infection. *Curr. Sci.*, 57: 895-896.
15. Vashishth, K., M. Fazal, M. Imran, M.M.A. Raza and Z.A. Siddiqui, 1994. Morphological and biochemical response of blackgram cultivars of *Meloidogyne incognita*. *Ann. Plant Prot. Sci.*, 2: 13-18

UNDER PEER REVIEW