

Isolation of wine yeast from sugar rich sources

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Abstract

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A total of 21 yeasts was isolated from different sugar-rich sources (date palm, pomegranate, roselle, orange, grapes, and dry date palm juice) collected from different locations in West Bengal, India. Total 21 yeasts were isolated from different sugar rich sources (date palm, pomegranate, roselle, orange, grapes and dry date palm juice) collected from different locations of West Bengal, India. All isolated yeasts were identified and differentiated based on their colony morphology and growth pattern on in Yeast Extract- Peptone -Dextrose (YEPD) Agar medium. The isolates could continue to survive on the slants for up to a maximum of 6 months up to maximum 6 months at 8-10°C temperature without any contamination. However, these were maintained by transferring to YEPD agar slants after every four months to maintain the purity and maximum viability of cultures. Fermentation was carried out at 26±1°C temperature at total soluble solid 24°Brix and pH 4.70 in molasses medium for all different isolated yeast. The total soluble solid at the end of fermentation in molasses solution were was found in the range of 9.00-20.78°Brix in all isolated yeast. The pH of the substrate was decreased (0.10-1.12) during fermentation. The cell count after complete fermentation was found in the range of 3.60×10⁸ CFU/ml to 8.79×10⁸ CFU/ml for all the isolated yeast. The concentration of alcohol in the molasses was observed at a range from 4.21 to 13.61 %. After screening or checking out the efficiency of yeasts on the basis of the highest alcohol production in molasses media, five yeasts (Y₄, Y₁₀₁, Y₃₃, Y₁₀₂, and Y₇) were selected for wine fermentation out of 24 (new isolates 21, previous isolations Y₄, Y₇, and Y₃₃) at the end of 18 days fermentation.

Key Words: Different fruit juice, Fermentation, Sugar, Alcohol, Yeasts

Introduction

Fermentation is one of the oldest forms of food preservation technologies in the world.

Indigenous fermented foods such as bread, cheese, and wine, have been prepared and consumed for thousands of years and are strongly linked to culture and tradition,

especially in rural households and village communities. The development of fermentation technologies is lost in the midst of history (Yokotsuka, 1985).

Anthropologists have postulated that it was the production of alcohol that motivated primitive people to settle down and become agriculturists. Some even think the consumption of fermented food is pre-human (Stanton, 1985). The process of

fermenting is basically feeding sugars and nutrients in solution to the fermenting yeast, which returns the flavour by producing carbon dioxide gas and alcohol (Garrison, 1993).

This process goes on until either all the sugar is used or the yeast can no longer tolerate the alcoholic percentage of the beverage. Different yeasts produce different results, and have different tolerance levels (Anon, 2005). Fermentation is a process of deriving

energy from the oxidation of organic compounds, such as carbohydrates, and using an endogenous electron acceptor, which is usually an organic compound (Klien *et al.*, 2005), as opposed to respiration where electrons are donated to an exogenous electron

acceptor, such as oxygen, via an electron transport chain. In alcoholic fermentation, the conversion of hexoses into ethanol and CO₂, forms the very basis of a successful wine fermentation.

The health benefits of (moderate) wine drinking are recognized by modern medicine. In Europe, a daily glass of red wine is often recommended to treat mild anemia; a class of chemicals called flavonoids (antioxidants in grape skins) appears to have many beneficial properties; another class of chemicals, called polyphenols, prevents

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(Angewandte, 2003) heart disease, arteriosclerosis and maybe some cancers (Yu *et al.*, 2021; Vejarano and Lujan-Corro, 2022; Yu *et al.*, 2024).

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Material and methods

The present investigation was made in respect of the isolation of wine yeast from sugar rich sources, after collected from different place of West Bengal. The experiment was conducted in the laboratory of Post Harvest Technology of Horticultural Crops, Directorate of Research complex, Bidhan Chandra Krishi Viswa Vidyalaya Kalyani, West Bengal. Molasses solution and different fruit juices were used for isolation of different yeast such as date palm, pomegranate, orange, black grape, dry date palm and roselle juice. The date palm juice was collected from different places of West Bengal during the year 2015-16. Completely randomized block design was adopted for statistical analysis of the data.

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The isolation of yeasts, preservation and maintenance were estimated by composition of medium (Deak, 1991). Total soluble solids (T.S.S.) were estimated using a pocket refractometer. The titratable acidity was determined by titrating against 0.1N NaOH and expressed as anhydrous citric acid, the ascorbic acid (mg/100g) content of fruit was determined by using 2, 6 dichlorophenol-indophenol dye by visual titration method. Reducing sugars and total sugars content of the fruits were estimated following the standard method described by Ranganna, (2000). Cell count was estimated using a digital microscope. Alcohol content was estimated by pycnometric method using distillation assembly following the procedure expressed by Sadasivum and Munickam, (2008).

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RESULT AND DISCUSSION

Over all 21 yeasts were isolated from different sugar rich sources and differentiated based on their colony morphology and growth pattern on Yeast Extract- Peptone - Dextrose (YEPD) Agar medium. The yeasts were isolated from date palm, pomegranate, roselle, orange, grapes and dry date palm juice under different agro-climatic zones of West Bengal regions in India and enlisted in Table 1.

Table 1: Different yeast isolates from different sources with colony morphology

Sl. No.	Yeast isolate	Source	Location (West Bengal)	Colony Morphology
1.	Y ₁₀₁	Date palm Juice	Jaguli	Whitish, smooth and shiny
2.	Y ₁₀₂	Date palm Juice	Mohanpur	Whitish, rough and dry
3.	Y ₁₀₃	Date palm Juice	Hoogly	Whitish, smooth and shiny
4.	Y ₁₀₄	Date palm Juice	Bardhaman	Whitish, smooth and shiny
5.	Y ₁₀₅	Date palm Juice	D.R.Building	Off white, rough and dry
6.	Y ₁₀₆	Date palm Juice	C. Block	Off white, rough and dry
7.	Y ₁₀₇	Date palm	Fatehnpur	Whitish, smooth and shiny

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		Juice			
8	Y ₁₀₈	Date palm	Sadapare	Off white, smooth and shiny	
		Juice			
9	Y ₁₀₉	Date palm	Pukarpare	Whitish, smooth and shiny	
		Juice			
10	Y ₁₁₀	Date palm	Raghunathpur	Whitish, smooth and shiny	
		Juice			
11	Y ₁₁₁	Date palm	Fatehhpur	Creamy white, smooth and shiny	
		Juice			
12	Y ₁₁₂	Date palm	Jaguli	Whitish, smooth and shiny	
		Juice			
13	Y ₁₁₃	Date palm	Jaguli Digha	Creamy white, smooth and shiny	
		Juice			
14	Y ₁₁₄	Date palm	Mubarakpur	Dull white, smooth and shiny	
		Juice			
15	Y ₁₁₅	Date palm	Tehatta	Off white, smooth and shiny	
		Juice			
16	Y ₁₁₆	Date palm	Gayeshpur	Creamy white, smooth and shiny	
		Juice			
17	Y ₁₁₇	Pomegranate	Mohanpur	White, rough and dry	
		Juice			

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18	Y ₁₁₈	Roselle Juice	C. Block	White, rough and dry
19	Y ₁₁₉	Orange Juice	Anukulmode	Creamy, smooth and shiny
20	Y ₁₂₀	Grape Juice	Anukulmode	Creamy, smooth and shiny
21	Y ₁₂₁	Dry date palm Juice	Anukulmode	White, rough and dry

Table 2: Growth of yeast at 26±1°C temperature on YEPD agar medium

Sl. No.	Yeast	26±1°C		
		16 hours	24 hours	48 hours
1	Y ₁₀₁	+++	++++	++++++
2	Y ₁₀₂	+++	++++	+++++
3	Y ₁₀₃	+++	++++	+++++
4	Y ₁₀₄	+	++	+++
5	Y ₁₀₅	+	++	+++
6	Y ₁₀₆	+	++	++
7	Y ₁₀₇	+	++	+++
8	Y ₁₀₈	+	++	+++
9	Y ₁₀₉	++	+++	+++++

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10	Y ₁₁₀	+++	++++	++++++
11	Y ₁₁₁	+++	++++	+++++
12	Y ₁₁₂	+++	++++	++++++
13	Y ₁₁₃	+++	++++	+++++
14	Y ₁₁₄	++	+++	+++++
15	Y ₁₁₅	+++	++++	+++++
16	Y ₁₁₆	++	+++	++++
17	Y ₁₁₇	++	+++	++++
18	Y ₁₁₈	++	+++	++++
19	Y ₁₁₉	Nil	+	++
20	Y ₁₂₀	++	+++	++++
21	Y ₁₂₁	++	+++	++++

+ = Very poor growth ++ = Poor growth +++ = Slightly moderate growth

++++ = Moderate growth +++++ = Good growth ++++++ = Very good growth

Growth response of different yeasts at 26±1°C temperature in incubation

Data presented in Table 2 on growth of yeasts at 26±1°C temperature indicated that majority of yeast isolates started to grow within 16 hours of incubation at 26±1°C except Y₁₁₉. Ten isolates viz. Y₁₀₁, Y₁₀₂, Y₁₀₃, Y₁₀₉, Y₁₁₀, Y₁₁₁, Y₁₁₂, Y₁₁₃, Y₁₁₄, and Y₁₁₅ showed good to very good growth at 26±1°C. Yeast Y₁₀₄, Y₁₀₅, Y₁₀₆, Y₁₀₇, Y₁₀₈, Y₁₁₆, Y₁₁₇, Y₁₁₈, Y₁₂₀ and Y₁₂₁ showed poor to moderate growth at 26±1°C after 48 hours of

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incubation. The isolate Y₁₁₉ failed to grow after 16 hours and after 24 hours growth was started at 26±1°C in incubation. The Y₁₁₉ yeast also showed poor growth with respect to colony thickness development on YEPD agar medium at 26±1°C.

While growing the yeast isolates on the YEPD agar medium it was observed that, the isolates could continue to survive on the slants up to 6 months at 8-10°C temperature without any contamination on all slants except Y₁₀₁, Y₁₀₂, Y₁₀₃, Y₁₁₁, Y₁₁₂ and Y₁₁₃. The survival was confirmed through transferring the same to the fresh YEPD agar slant and incubated at 26±1°C. The growth was observed by 24-hour, except the slants of Y₁₁₉. However, all the yeast isolated were maintained throughout the period of this investigation by transferring it to YEPD agar slants after every four months and preserving those at 8-10°C temperature to maintain the purity and maximum viability of cultures beyond any doubt.

It was also observed that in order to have significant growth on the YEPD agar slants, irrespective of extent of growth it had to be incubated for 24 - 48 hours at 26±1°C except above 48 hours for Y₁₁₉.

After observing the growth on the medium all the yeasts with prolong viabilities were tried to test their fermentation capability to produce ethyl alcohol. It was apparently confirmed through production of bubbles in the medium and the odour of alcohol, till the end of fermentation. In this study the yeasts Y₁₀₁, Y₁₀₂, Y₁₀₃, Y₁₁₁, Y₁₁₂ and Y₁₁₃ had shown very distinct bubbling and rest of the yeasts did not give much indication of alcoholic fermentation during the period of this experiment. However, at the end of the period of fermentation the analysis for the production of ethyl alcohol was done for all the yeast, under study, to confirm their alcohol production ability for those even which did not show any physical change of medium. Fermentation was carried out at 26±1°C

temperature at pH 4.70 without nutrient supplements in molasses medium. The yeasts had shown sufficient growth at 24°C and might very easily be tried to test their alcohol production ability in this investigation as this range of temperature, was within the range mostly used to produce wine (Joshi *et al.*, 1999b) and this is the range which prevails

during most of the time of the year, in the eastern part of the country (Baidya, 2010).

Yeasts are active in a very broad temperature range from 0 to 50°C, with an optimum temperature range of 20°C to 30°C (Mountney and Gould, 1988).

Alcohol production efficiency using molasses solution at 26±2°C

The initial data of broth solution have been showed in Table 3.

Table 3: Initial data of broth solution

Sl. No.	Parameters	Source
1.	Total soluble soiled (°Brix)	24.00
2.	Reducing sugar (%)	11.76
3.	Total sugar (%)	16.66
4.	pH	4.70

Changes in total soluble solids (°Brix)

The results shown (Table 4) in this study the residual degree Brix was measured after every 2 days of fermentation to study the sugar utilization by all the yeasts. All yeast isolates could utilize sugar after 24-hour of fermentation. These indicated that these 4 yeasts (Y₁₀₇, Y₁₀₉, Y₁₁₁ and Y₁₁₉) either could not survive on this molasses substrate at 26±1°C or these were not the fermenting agents of molasses.

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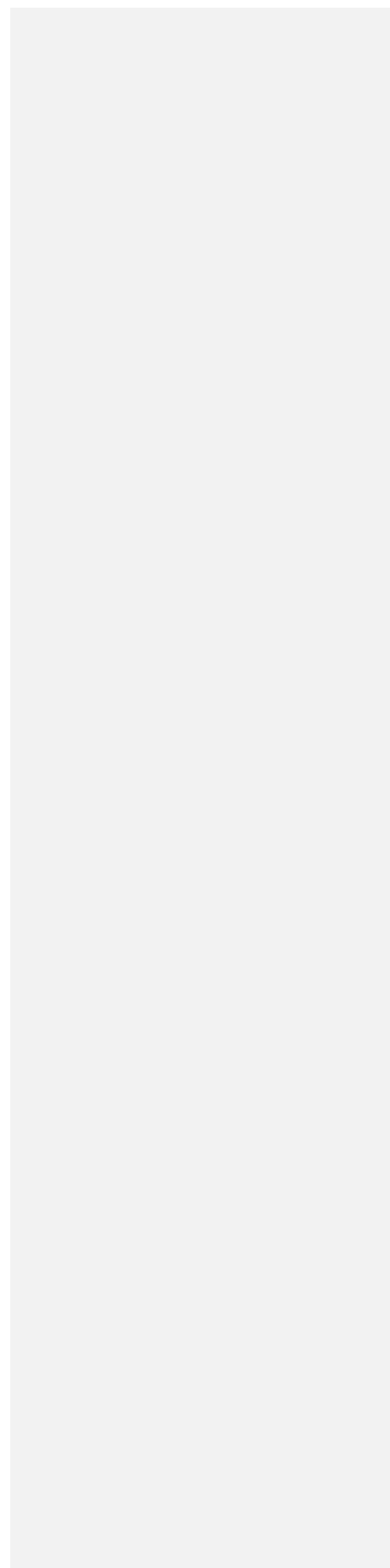
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**Table 4: Changes in total soluble solids (°Brix) during fermentation in molasses
broth at $26 \pm 1^\circ\text{C}$**

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Sl. No.	Yeast	Total soluble solid (°Brix)									
		0 days	2 days	4 days	6 days	8 days	10 days	12 days	14 days	16 days	18 days
1	Y ₄	24	16.50	15.97	12.83	12.27	11.10	11.00	10.90	10.77	10.67
2	Y ₇	24	11.50	10.33	9.87	9.80	9.70	9.63	9.63	9.63	9.50
3	Y ₃₃	24	15.43	15.17	10.43	9.53	9.13	9.13	9.10	9.10	9.00
4	Y ₁₀₁	24	15.43	15.37	14.57	13.47	12.33	12.00	11.33	10.87	10.40
5	Y ₁₀₂	24	13.93	10.73	10.43	10.37	10.27	10.27	10.20	9.90	9.87
6	Y ₁₀₃	24	12.40	11.93	9.73	9.63	9.50	9.47	9.47	9.43	9.40
7	Y ₁₀₄	24	19.00	15.37	15.37	15.30	15.20	15.20	15.17	15.17	15.13
8	Y ₁₀₅	24	19.50	16.70	15.40	15.37	15.37	15.30	15.27	15.27	15.23
9	Y ₁₀₆	24	18.57	17.97	16.17	16.16	16.13	15.83	15.70	15.67	15.13
10	Y ₁₀₇	24	23.96	23.57	22.73	22.53	22.13	16.60	16.60	16.60	16.57
11	Y ₁₀₈	24	19.07	17.63	15.47	15.40	15.30	15.20	15.16	14.37	14.37
12	Y ₁₀₉	24	23.67	23.73	23.43	23.13	21.07	18.00	17.00	16.83	16.17
13	Y ₁₁₀	24	20.87	18.00	11.73	11.00	11.00	10.97	10.37	10.23	9.50
14	Y ₁₁₁	24	24.00	23.70	23.03	23.00	22.60	22.40	22.27	21.65	20.78

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15	Y ₁₁₂	24	19.17	15.80	15.30	13.77	9.23	9.17	9.13	9.08	9.00
16	Y ₁₁₃	24	22.87	16.03	15.60	12.83	12.20	10.33	10.23	10.12	9.87
17	Y ₁₁₄	24	23.80	23.10	22.43	22.10	21.00	17.10	13.87	13.52	12.88
18	Y ₁₁₅	24	22.23	18.90	18.87	18.30	17.27	15.17	15.20	14.93	13.65
19	Y ₁₁₆	24	20.87	16.70	15.87	15.73	14.50	13.30	13.23	13.10	12.56
20	Y ₁₁₇	24	23.53	20.17	17.17	16.33	16.23	15.80	14.30	14.27	12.60
21	Y ₁₁₈	24	23.47	21.03	17.47	15.90	14.87	14.17	12.37	12.37	11.17
22	Y ₁₁₉	24	24.00	23.83	23.33	22.00	21.43	20.27	18.97	18.77	17.80
23	Y ₁₂₀	24	24.00	22.80	20.67	19.63	17.93	15.07	13.00	12.60	11.33
24	Y ₁₂₁	24	23.23	19.90	16.47	15.07	14.87	13.57	11.70	11.57	9.93
SE(m)		NS	NS	NS	NS	NS	NS	NS	NS	NS	0.194
CD at 5%		NS	NS	NS	NS	NS	NS	NS	NS	NS	0.553

The yeast isolates viz. Y₇, Y₃₃, Y₁₀₂, Y₁₀₃, Y₁₁₀, Y₁₁₂, Y₁₁₃ and Y₁₂₁ had utilized significant quantity of sugar after 18 days of study which was reflected from the residual °Brix of 9.00-9.93 respectively. At the end of fermentation period under study (after 18 days) it was revealed that the residual degree brix was in the range of 10.40-15.23 when molasses was inoculated with Y₄, Y₁₀₁, Y₁₀₄, Y₁₀₅, Y₁₀₆, Y₁₀₈, Y₁₁₄, Y₁₁₅, Y₁₁₆, Y₁₁₇, Y₁₁₈ and Y₁₂₀ for the purpose of studying alcohol production efficiency by these yeasts. It means that sugar utilized approximately by these yeasts was in the range of 8.77-13.60 % which was very negligible. This quantity of sugar might

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have been utilized for their own survival. A gradual decrease in TSS which is a typical nature of fermentation was observed in this experiment by many workers as it was observed by Ezeronye (2004) in their experiment with fruit wine. Molasses solution having 24°brix and pH 4-5 was selected as the most effective and economic substrate for this purpose as it is normally utilized for the industrial alcohol production by many distilleries and by the researchers (Bardiya *et al.*, 1976).

Changes in reducing sugar, total sugar and pH

Reducing sugar and total sugar were decreased during the fermentation of molasses. After 18 days of fermentation reducing sugar and total sugar were found in the range of 3.06-7.67 per cent and 3.73-13.34 per cent in all isolated yeasts respectively. The maximum reducing and total sugar were found in Y₁₀₉ (7.67%) and (13.23%) and minimum in Y₁₁₀ (3.06%) and (3.73%) after 18 days of fermentation respectively.

During the period of study, in most of the cases pH was changing and later it became stable. The pH of substrate generally 0.10-1.12 was decreased by over a period of 18 days during fermentation by various yeasts but pH was 4.81-5.02 i.e., 0.11-0.32 increase from initial pH of 4.70 and it was found in case of Y₇, Y₃₃ and Y₁₀₃ pH was slightly increased as that of initial pH of 4.70 during fermentation periods (Table 5).

It is due to the production of organic acids by the yeasts and the stability of the pH was probably due to efficient buffering of protons in the fermenting medium by the weak acids produced (Abiose and Adedji, 1994; Elijah *et al.*, 2007).

Table 5: Changes in reducing sugar, total sugar and pH during fermentation using molasses at 26± 1°C after 18 days of fermentation

Sl. No.	Yeast	Reducing sugar (%)	Total sugar (%)	pH
1	Y ₄	5.11	8.58	4.33
2	Y ₇	3.56	7.89	4.81
3	Y ₃₃	3.28	7.65	5.02
4	Y ₁₀₁	3.23	9.40	4.48
5	Y ₁₀₂	3.15	8.23	4.03
6	Y ₁₀₃	3.18	7.46	4.94
7	Y ₁₀₄	6.27	12.03	3.92
8	Y ₁₀₅	5.75	12.12	3.87
9	Y ₁₀₆	5.38	12.45	4.04
10	Y ₁₀₇	6.52	13.24	4.17
11	Y ₁₀₈	6.33	12.00	4.09
12	Y ₁₀₉	7.67	13.34	3.95
13	Y ₁₁₀	3.06	3.73	4.60

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14	Y ₁₁₁	7.03	8.60	3.84
15	Y ₁₁₂	6.53	6.80	4.08
16	Y ₁₁₃	5.90	7.27	3.81
17	Y ₁₁₄	6.37	8.80	3.58
18	Y ₁₁₅	6.23	7.22	3.94
19	Y ₁₁₆	6.67	7.13	4.10
20	Y ₁₁₇	5.84	6.80	4.21
21	Y ₁₁₈	6.08	6.54	4.24
22	Y ₁₁₉	6.10	13.31	4.11
23	Y ₁₂₀	5.59	6.37	4.30
24	Y ₁₂₁	4.48	7.01	4.24
SEm(±)		0.082	0.136	0.062
CD at 5%		0.235	0.387	0.176

Production of alcohol and cell count from substrate

As shown in Table 6, a steady increase in alcohol content was observed in the molasses throughout the period of fermentation with the test yeast strains. At the end of the 18 days fermentation, the concentration of alcohol in the molasses was observed at range from 4.21 to

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2	Y ₇	10.35	6.93
3	Y ₃₃	11.88	8.79
4	Y ₁₀₁	12.01	5.14
5	Y ₁₀₂	11.06	6.21
6	Y ₁₀₃	8.20	5.84
7	Y ₁₀₄	5.44	3.60
8	Y ₁₀₅	5.78	5.78
9	Y ₁₀₆	7.02	5.68
10	Y ₁₀₇	4.28	6.21
11	Y ₁₀₈	9.41	6.10
12	Y ₁₀₉	4.21	5.45
13	Y ₁₁₀	8.55	7.60
14	Y ₁₁₁	5.44	6.57
15	Y ₁₁₂	10.01	7.13
16	Y ₁₁₃	6.09	8.17
17	Y ₁₁₄	8.96	8.78
18	Y ₁₁₅	7.23	8.14

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19	Y ₁₁₆	4.65	7.69
20	Y ₁₁₇	8.55	7.26
21	Y ₁₁₈	7.55	6.84
22	Y ₁₁₉	5.91	4.74
23	Y ₁₂₀	7.90	6.30
24	Y ₁₂₁	8.06	6.52
SEm(±)		0.121	0.094
CD at 5%		0.344	0.268

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Conclusion

Molasses solution and different fruit juices were used for isolation of different yeast such as date palm, pomegranate, black grape and roselle, juice under different agro-climatic zones of West Bengal regions in India. Yeast isolate numbers Y₁₀₄, Y₁₀₅, Y₁₀₆, Y₁₀₇, Y₁₀₈, Y₁₁₆, Y₁₁₇, Y₁₁₈, Y₁₂₀ and Y₁₂₁ showed poor to moderate growth at 26±1°C after 48 hours of incubation. Isolate numbers Y₁₀₁, Y₁₀₂, Y₁₀₃, Y₁₀₉, Y₁₁₀, Y₁₁₁, Y₁₁₂, Y₁₁₃, Y₁₁₄, and Y₁₁₅ showed good to very good growth at both 26±1°C. The isolate Y₁₁₉ failed to grow after 16 hours and after 24 hours growth was started at 26±1°C in incubation. The total soluble solids after end of fermentation in molasses solution were found in the range of 9.00-20.78^oBrix in all isolated yeast. After 18 days of fermentation reducing sugar, total sugar and pH were found in the range of 3.06-7.67 per-cent, 3.73-13.34 per-cent and 0.10-1.12 in all isolated yeasts respectively. The cell count after complete fermentation was found in the range of 3.60×10⁸ CFU/ml to 8.79×10⁸ CFU/ml for all

the isolated yeast. In this study the highest alcohol production was observed in the isolates Y₄, Y₁₀₁, Y₃₃, Y₁₀₂, and Y₇ at the end of the 18 days of fermentation. After screening or checking the efficiency of yeast in molasses media, five yeasts were selected for wine fermentation.

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