

Minireview Article

**New Chemical Dualities Illustrated by *Meso* and
 C_2 Symmetrical (*CTS*) Compounds**

UNDER PEER REVIEW

ABSTRACT

The most important relationship between chemistry and genetics is nonetheless the corpuscular nature of their objects, molecules (or atoms) and genes, respectively. On the other hand, one states, without a substantial proof, that philosophy should be one step ahead all sciences. Here is a proof that the reverse can also be true. Two internal enantiomeric halves of *meso* compounds or the two chiral halves of C_2 symmetrical isomers constitute pairs of entities suitable to work as duality phenomena in science. Four types of isomers have been identified: (A) *meso*, (B) C_2 symmetrical (*CTS*), (C) *irregular chiral (irrechi)* and (D) *constitutional (constit.)*. *Meso* and *CTS* are characteristic to plants and microorganisms. Almost all natural micromolecular compounds from vertebrate tissues are asymmetric, i.e. they are *constit.* isomers. An exception to this rule is *meso*-inositol, an isomer of hexoses, which are themselves, **as their congeners asymmetric**. By comparing the real (envisaged) *meso* isomers of these compounds with the asymmetric ones of vertebrate tissues, the reason for nature selected the latter became quite evident: it is **the omission of** a suite of structural restrictions. Delivering of *meso* isomers of natural compounds discloses a huge chemical philosophical potential of this issue. An intrinsic property of *meso* combinations is their character of dimerism, hence their molecule is formed of two entities that are contrary in a spatial, chemical and optical sense, i.e. good candidates for a duality concept. Moreover, a good deal of material is indicated, i.e. *CTS* isomers, whose sides are chiral and identical, for a new type of duality in philosophy, strongly expressed in nature by a chemical language.

Keywords: isomers, meso, C₂ symmetrical, irregular chiral (irrechi), constitutional, comparative chemistry, integrative, duality, mirror plane of symmetry

1. INTRODUCTION

Biochemical and genetic center running today is formed almost exclusively of genome [1-3]. All products, their molar ratios, phenomena of molecular and isomeric diversity, as well as entropy buffers are **produced by the genome**. However, genetic apparatus is tested especially by its products. Duality phenomena are known especially in physics, e.g. light duality [4-7]. The two opposed sides of a duality phenomenon should be well defined, equally (comparably) strong concerning their meaning, and relatively known (consecrated). In chemistry duality phenomena are of less amplitude [8-10] than in physics, and this can be correlated with the relative lower oldness of this science, as based on firm and general principles, in comparison with physics. One might suppose that the birth certificate of physics was signed by Pythagoras (abt 532 BC) when he found a mathematical relationship between sound quality, the length, thickness and tension of the producing string (or pipe) [11]. Chemistry began probably at the same time, but a series of occult influences slowed its development.

Structural analysis of numerous natural compounds as well as *in silico* integrative approach of isomers generated by the same molecular formula, applied to a large diversity of natural compounds and some synthetic ones, led to four groups (types) of compounds identified and defined within the same molecular formula: (A) *meso*, (B) C_2 symmetrical (*CTS*), (C) *irregular chiral (irrechi)* and (D) *constitutional (constit.)* [12-17].

(A) Meso, are either based on a mirror plane of symmetry (A1), or devoid of a mirror plane of symmetry but specified in this way as a result of Cahn-Ingold-Prelog rules [18,19] (A2). One can assert that molecules of the latter group are formed of two imaginary enantiomeric halves separated by an imaginary mirror plane of symmetry. Related to *meso* isomers are compounds characterized by a center of symmetry (A3) or an alternating axis of symmetry

(A4). The molecule of (A1)-(A3) is formed of two enantiomeric halves. *Meso* isomers are optically inactive (optinactive) due to an internal compensation.

At the time when Fischer invented xylitol [20], an optinactive polyol, at least one *meso* isomer was known, i.e. *meso*-tartaric acid, discovered by Pasteur [21,22]. *Meso*-tartaric acid has a homodimeric structure (an even number chain), and xylitol a **heterodimeric** (an odd number chain). Mirror plane of symmetry cuts a bond of *meso*-tartaric acid, and four atoms of xylitol (C, H, OH). One might theorize that mirror plane of symmetry hides (masks) the atoms cut by it from polarized light, and what remains, as evidenced by this physical instrument, is an entity containing an even number of atoms, i.e. a homodimer. Mirror plane of symmetry has to be regarded as an intrinsic **property of *meso* compounds, both a physical instrument and a natural phenomenon.**

Meso heterodimers constitute a chemical duality, the two opposed sides of duality are their heterodimeric character, on one hand, and their expression as homodimers, on the other. According to Kelvin and Prelog theory [23-25] "*meso* compounds are internally heterochiral. There is a fundamental difference between the mirror plane of symmetry in macrocosmos and at physical-chemical level in microcosmos. In the first case, the mirror plane of symmetry just indicates the limit of the two enantiomeric halves. At physical-chemical level, it can cut atoms and hide them, not of our seeing, but of polarized light. As will be evident of this paper, this spectacular property of mirror plane of symmetry plays an extremely important role in systematization of isomers emerging of the same molecular formula".

Imprecise breaking of identity of the two halves of *meso* isomers leads to two enantiomers [26] i.e. the internal enantiomerism is externalized. (In our days chemists try to overturn this feature of *meso* compounds and to predominantly **(or even exclusively)** prepare one product only [27-31]).

(B) " C_2 symmetrical (CTS) compounds have been defined in relation with an axis and a rotation of 180° . After this maneuver the same atoms should be regained as initially" [32-34].

All CTS compounds are chiral and optically active (optactive). Fischer demonstrated the existence of some chiral compounds with identical ends, that produced exclusively one derivative, by reactions randomly affecting their ends. E. g. D- and L-mannitol [35], D- and L-iditol [36], and their aldaric acids, as well as D- and L-threitol [37] and the enantiomers of tartaric acid [38]. Besides these compounds whose molecule is formed exclusively of two identical chiral halves, there are CTS combinations where the two chiral halves are linked on a matrix. E. g. 3-keto- and 3-deoxyxylitol, 3-keto- and 3-deoxyribitol, D- and L-diaminopimelic acid [15], etc. The chiral units of the third CTS group molecules can be recognized especially by Cahn-Ingold-Prelog rules [18,19]. According to Kelvin and Prelog theory [23-25], "*CTS* formed exclusively of two identical chiral halves are homochiral with each other" and internally homochiral [34,39]. Of this reason, they could be named also *twin* molecules [40]. "The exceptional properties of *twin* (CTS) compounds" were also noticed by Vickery [41]. "Homodimeric CTS compounds constitute a chemical duality, the two opposed sides of duality are optical activity, on one hand, and their symmetry, on the other. There is one universal rule concerning CTS compounds: every member of this group possesses a real or imaginary, but plausible, *meso* isomer. Some more clearings are requisite. Compounds based on 1,2-diamino-cyclohexane [29,33,42,43] are CTS as long as they are trans". "Their cis isomer should be *meso* only by adopting a planar cycle, as for allo-inositol. Of the six *meso* isomers of inositol [44,45], five are characterized by 1,4 mirror plane of symmetry, while allo-inositol is devoid of such a plane. Its *meso* nature can be explained only by a planar structure, hence the mirror plane of symmetry cuts two opposed bonds". (One can write a *meso* isomer of 1,2-diamino cyclohexane as 1,2-cyclobutane derivative).

"The first CTS combinations, the two enantiomers of tartaric acid, have been separated by Pasteur (1848) by crystallization from a racemic mixture that had been prepared by Kestner (1822)" [21,22,46]. "Pasteur noticed two types of crystals, that were enantiomorphic with one another. He separated the two types of crystals and found out that their aqueous solutions were dextrorotary and levorotary, respectively. Dextro-tartaric acid had been discovered by

Scheele (1770) in the sediment deposited in the vats during the grape juice fermentation" [47,48]. "Stereochemical theory of tetrahedral and asymmetric (chiral) carbon atom [49,50] led van't Hoff to molecular models based on tetrahedrons which unequivocally represented every chiral carbon atom". "By constructing and using these models, van't Hoff expanded the idea of enantiomorphism from crystals to molecules, a process initiated by Pasteur. (Dots and wedges representations of today come from van't Hoff's models). However, at that time no scientist could rationally associate structural models with the two enantiomers" [51]. "In fact, the discovery of Pasteur increased the dilemma of representation, i. e., the relationship between a sample of an optactive compound and the unique, characteristic, structural model possibly assigned to it. This dilemma was solved by X-ray diffraction, i. e., of zirconium K α rays, by sodium rubidium tartrate of the dextrorotary species, and the obtained model was assigned to (+)-tartaric acid" [52]. "Configuration of chiral centers of (-)-tartaric acid became also known, by the virtue of the law of enantiomorphism. By an impressive coincidence, this configuration of (+)-tartaric acid had been hypothetically attributed by E. Fischer" [38]. "Configuration of the two enantiomers has been connected with other chiral compounds, beginning with (-)- and (+)-glyceraldehyde" [53,54]. "A chemical relationship has been found between E. Fischer and his son, H. O. L. Fischer" [55,56]," due to a derivative of D- and L-mannitol prepared by the latter, i.e. 1,2-5,6-di-O-isopropylidene mannitol (*CTS*). By integration of finding of H. O. L. Fischer in the strategy of E. Fischer, a remarkable shortcut to structure elucidation of linear aldohexoses is obtained" [57].

The theory of van't Hoff and Le Bel was confirmed by three independent arguments. (a) Synthesis by E. Fischer of the major part of monosaccharides indicated by this theory [58]. (b) In a model of diamond based on crystal structure determined by X rays diffraction, C atoms appeared on the apices of an endless lattice of regular tetrahedrons [59]. (c) Pauling [60,61] explained "this model in mathematical-physical terms by using a concept, hybridization, taken from biology. Pauling demonstrated that by mixing three p orbitals, each having two lobes and being perpendicular to one another, with a spherical s orbital, four hybrid sp³ orbitals are obtained, that are identical and oriented along the axes of a regular tetrahedron".

A remarkable and unique feature of *CTS* compounds is that chemical modification of one of the two component chiral halves produces the same result.

(C) *Irrechi*. "The third subgroup of isomers of *meso* compounds are also chiral and they are characterized by a molecular skeleton identical to *meso* and *CTS*, i.e. a phenomenon of isoskeletomeric relationship" [62]. Still, chiral carbons are irregularly distributed in their molecule [14,15] (*irregular chiral, irrechi*). *Meso* isomers are characterized by a 1:1 ratio of numbers of R and S carbons while in *CTS* ones this ratio is n:0, 0:n or 1:1. In *irrechi* combinations the ratio R/S has other values.

(D) "Constitutional" (positional) (*constit.*) isomers form the fourth group. They are isomer with the preceding ones but their skeleton is different. They are either optactive or optinactive" [14]. With relatively few exceptions, compounds currently met in living things, especially in vertebrate tissues, are *constit.* isomers. They are probably the most abundant in these living things.

The term diastereomer has been destined to include all isomers that are not enantiomers [63-68]. Hence, it included all types of isomers mentioned above. However, since they have developed as independent groups, we considered that the term diastereomer became obsolete and have replaced it with the four redefined terms.

A direct application of our systematization to monosaccharides discovered/invented by Fischer would clearly indicate our strategy. Galactitol, iditol, xylitol, ribitol, erythritol, are *meso*, mannitol and iditol are *CTS*, altritol, glucitol, gulitol, talitol, arabinitol, lyxitol are *irrechi*, hamamelitol and apitol are *constit.* What has been affirmed for polyols is also valid for their aldaric acids.

"Concerning limits and possibilities of reciprocal changing of types mentioned above, both *CTS* and *irrechi* can be transformed into *meso*. Some interesting facts should be mentioned:

the molecule of iditols and idaric acids possesses an equal number of R and S carbons, similarly with galactitol, allitol, galactaric and allaric acids. However they are not *meso* but optactive" [44]. The difference can be explained probably by the fact that the molecule of the former is formed of two identical chiral halves and the latter of two chiral enantiomeric halves.

The two hydrogen atoms of central methylene of a *meso* derivative, i.e. 3-deoxyxylitol, 3-deoxyribitol, *meso*-diaminopimelic acid, etc., are not equivalent. If they are alternatively replaced by a hydroxyl function, the products are different. The two central hydrogen atoms of *CTS* compounds, i.e. 3-deoxyarabinitol, 3-deoxyxylitol, L,L- and D,D-diaminopimelic acid, etc., are equivalent: if they are alternatively replaced by a hydroxyl function, exclusively one product is obtained.

"The isomeric diversity is connected with the following factors: (i) Structures as diamond [59], graphite and fullerenes [69,70] illustrate the best the ability of C atoms to bind with each other". However, "the three forms present a very limited structural variety. (ii) What really confer molecular diversity to C combinations is the association of this element with hydrogen and this is evidenced by the remarkable molecular variety of aliphatic hydrocarbons" [45,71,72]. "Isomeric diversity is a physical-chemical magnitude concerning the ability of a compound to present a large number of isomers. (iii) Chemical functional groups, in relative low proportion, also favor molecular diversity. (iv) Aromatic hydrocarbons present the lowest isomeric diversity of all organic combinations. They contain an exceeding number of chemical functions, and they are in a state of advanced oxidation. In fact, they fill an intermediate place between elementary carbon and aliphatic hydrocarbons. Another remarkable feature of aromatic hydrocarbons is the fact that they do not present *meso* form as atropisomers. (v) Isomeric diversity increases exponentially with molecular weight" [63,72,73]. (vi) "Carbon dioxide is a terminal facet of metabolism and combustion of organic compounds. It is characterized by a high chemical inertia. Carbon dioxide has to be attached to a preexisting structure, as a piece of metal in a lathe, and stepwise reduced, the energy of sun playing an essential role in this process called photosynthesis" [74].

About eight classes of natural compounds contain *meso* isomers. **Monosaccharides:** *meso*-tartaric acid [22,49,75,76], erythritol [77], butanediol [78], galactitol [26,79], allitol [80], xylitol [81], ribitol [82,83], allaric acid [84], xylaric acid [85], ribaric acid [86], galactaric acid [26], D-galactitol-3R,4S-cinnamicacetal [87], galaocitol [88-90].

Amino acids and their derivatives: *meso*-cystine [91,92], *meso*-diaminopimelic acid [74,92-96], *meso*-lanthionine [96-98], L,D-homolanthionine [99], *meso*-DKP of pipercolic acid [100], dragmacidin B [101], fellutanines A and C [102], dimethyl fellutanine A [103], fellutanine D [104,105], trans-dragmacidin C [101], chimonanthine [106,107], petrobactin [108,109], phenazostatin D [110,111].

Carotenoids and carotenes: zeaxanthin [(3R,3'S)- β,β -carotene-3,3'-diol] [112-115], (2R,2'S)-2,2'-dihydroxy- β -carotene [(2R,2'S)- β,β -carotene-2,2'-diol] [116], tunaxanthin D [(3R,6S,3'S,6'R)- ϵ,ϵ -carotene-3,3'-diol] [117-122], tunaxanthin E [(3R,6R,3'S,6'S)- ϵ,ϵ -carotene-3,3'-diol] [117,120], *meso*-astaxanthin [(3R,3'S)-3,3'-dihydroxy- β,β -carotene-4,4'-dione] [123,124], (6R,6'S)-3,3'-diketo- ϵ -carotene [(6R,6'S)- ϵ,ϵ -carotene-3,3'-dione] [119,122], ϵ -carotene [(6R,6'S)- ϵ,ϵ -carotene] [125], γ,γ -carotene [(6R,6'S)- γ,γ -carotene] [119], glabrescol [126], squalane [127], lycopane [128], carotane [128-130], isorenieratane [131], renierapurpurane [131], 1,10-bis[2',2',6'-trimethylcyclohexyl]-3,8-dimethyldodecane [132,133].

Lignans: nordihydroguaiaretic acid [134], *meso*-dihydroguaiaretic acid [134], machilin A [135], dimethyl *meso*-dihydroguaiaretic acid [136], saururin A [137], pre-gomisin [138,139], 7,7'-dioxodihydroguaiaretic acid [140], 3,3'-didemethoxynelectandrin B [134], nelectandrin B [135,141,142], galgravin [143,144], zuonin B [145], 4-O-Me-saurucinol J [146], isonelectandrin B (tetrahydrofuroguaiacin B) [142], di-O-Me tetrahydrofuroguaiacin B [146], *meso*-secoisolariciresinol [147,148].

Neolignans: asarolignan A [149].

Cyclobutane derivatives: endiandrin B [150,151], cinbalansan [152], heterotropan [152], α -duplicatin B [153], piplartine dimer A [154], α -truxillic acid, γ -truxillic acid, epi-truxillic acid, ϵ -truxillic acid, peri-truxillic acid, β -truxillic acid, ω -truxillic acid [155], caracasandiamide [156].

Phenols: (3R,5S)-hannokinol [157], (3S,5R)-octahydrocurcumin [158], (2S,3R)-diolmycin B1 [159], (2S,3R,4S,5R)-hybocarpone [160-162], (2S,3S,4R,5R)-hybocarpone [160-162], eurorubrin [163], isochamaejasmine [164].

Terpenoides: daibudilactone C and daibudilactone D [165]. All this suite of *meso* compounds, the majority of them natural combinations, is meant to indicate that this type of symmetry, i.e. antinomy, is a natural phenomenon, although relatively limited. They are found almost exclusively in microorganisms, plants and/or lower animals [12-14].

2. THE LIMIT OF SYMMETRY OF THE MAJOR METABOLITES

Real and envisaged *meso* isomers. There are few, if any, symmetric compounds produced by vertebrates. Of the common metabolites, only monosaccharides (aldoses and ketoses) possess *meso* isomers as natural compounds. However, the potential of natural metabolites to produce imaginary but plausible *meso* isomers is really huge. The major metabolites containing a significant alkane moiety possess at least one real or envisaged *meso* isomer. A guiding line of this paper is to find out at least one *meso* isomer for every molecular formula. A serious obstructor to this is an advanced degree of unsaturation. It is impossible to find out a *meso* isomer for e.g. $C_4H_4O_4$ (fumaric/maleic acids). However, $C_6H_{10}O_4$ (succinic acid, etc.) or $C_6H_8O_4$ (2,3-dimethyl derivative, etc) have a *meso* form (Fig. 1). Similarly, every tentative to construct a *meso*

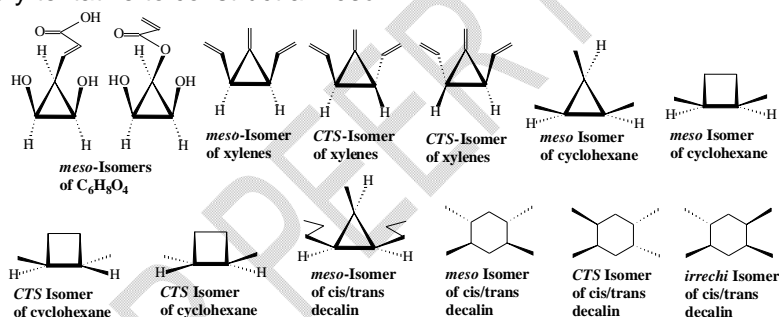


Figure 1. *Meso* isomers of unsaturated (fumaric/maleic acid), aromatic, and the latter's saturated derivatives.

isomer of benzene, fails. However, the thing is possible for xylenes, ethylbenzene, propylbenzene, etc. Also, reduction product of benzene, cyclohexane, presents *meso* and CTS isomers. Naphthalene, similarly to benzene, fails to give *meso* isomers, decalines instead presents all four types of isomers (Fig. 1). Compounds unable to produce symmetric isomers have been called by us *archaic*. Chemical transformations only intermediate between the two groups. For numerous *archaic* compounds a molecule of H suffices to convert them to symmetric entities.

At least two dozens of isomers with molecular formula $C_3H_7NO_2$ can be written, just by using the consecrated valence of every component element. However, of the envisaged isomers only some present elements of symmetry: two are *meso* (cis-1,2-dihydroxy-3-amino cyclopropane and cis-2,4-dihydroxy-azetidine), and two are CTS (trans-2,4-dihydroxy-azetidine, two enantiomers), and all the others, including (R)- and (S)-alanine, are *constit*.

Polygonal representations of *meso* isomers. *Meso* representations in this paper, of compounds with at least a minimum degree of unsaturation, are polygonal – triangular, square, pentangular, hexangular. An exception to this are saturated compounds (alkanes,

alcohols, etc). For a triangular representation (Fig. 2), a mathematical equation (1) have been imagined to illustrate *meso* isomers.

$$n-3=2x+2y+z+w \quad (1)$$

In equation (1), n is the number of atoms in molecular skeleton, x, y, z, w , are suitably selected numbers. In triangular representation there is a connection between x, y, z, w , and R_1, R_2, R_3, R_4 , respectively.

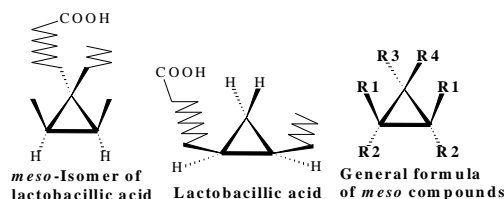


Figure 2. Lactobacillic acid as model for *meso* isomer. The sense of mathematical equation.

The rings of three or four atoms, as cycles or heterocycles, synthetic [30,166-168] or found in natural materials, are well known. “Cis- and trans-1,2-dimethyl cyclopropane are indistinguishable of thermodynamic point of view” [169]. “1,2,3-Trihydroxycyclopropane is known as an unstable combination” [170,171], however “no attempt was made to stabilize it. 1,2-Dihydroxycyclopropane has been prepared by a reduction reaction of a diketone derivative” [172]. “Cis-1,2-dihydroxycyclopropane has been discovered in natural material as a glycoside of α -D-galactopyranose [173] as well as in the constitution of mycolic acids” [174] and lactobacillic acid [74] (Fig. 2). Oxirane ring has been identified as (3S)-2,3-oxidosqualene in sterols biosynthesis. Two syntheses of cis-1,2,3,4-tetrahydroxy cyclobutane have been reported [175]. Numerous real or envisaged *meso* isomers have been presented in the following (Figs. 3-10).

3. NATURAL COMPOUNDS WITH BIOCHEMICAL IMPLICATIONS

3.1. The fundamental amino acids. “Compounds with a ubiquitous distribution in living matter, the twenty fundamental amino acids are characterized by an unequalled structural variety. These amino acids are met especially integrated in proteins and in this state they manifest themselves by their tails” [74]. An interesting picture presents the real and envisaged symmetric isomers of the twenty fundamental amino acids: without any exception, they present *meso* isomers (Fig. 3), hence no one is *archaic*. Besides *constit.* isomers, Gly, Ala, Val, Pro, Thr, Asp, Arg, present *meso* and *CTS* isomers. Leu, Ile, Glu, Asn, Lys, present all four types. Trp, Phe, Tyr, His, Ser, Gln, Cys, Met, possess, beside *constit.*, *meso* and *irrechi* isomers. It is evident that all these compounds present symmetric isomers and implicitly dimeric character. The twenty fundamental amino acids, with different molecular formula, are related with each other by their common

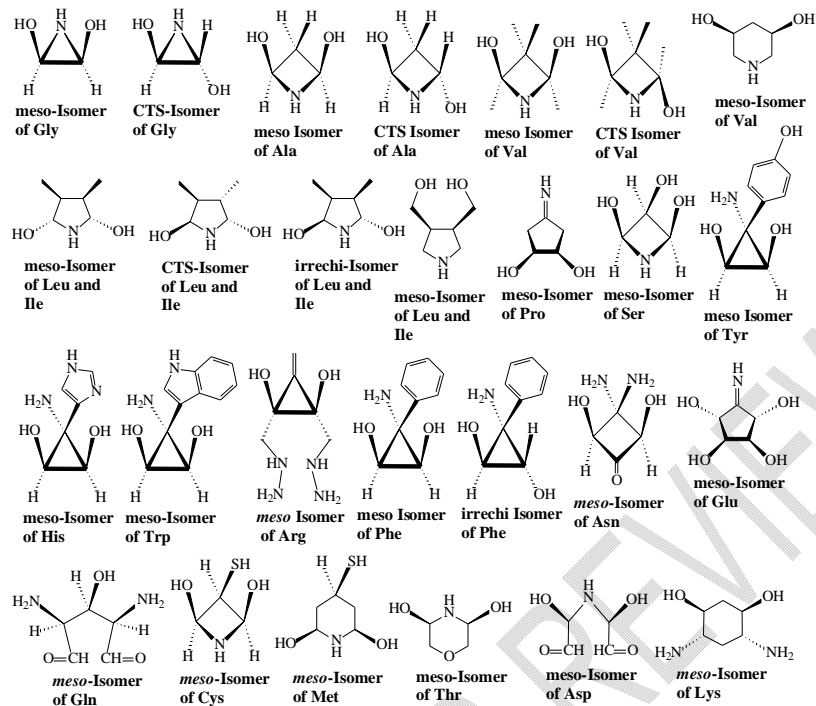


Figure 3. *Meso* isomers of the twenty fundamental amino acids. (see also text).

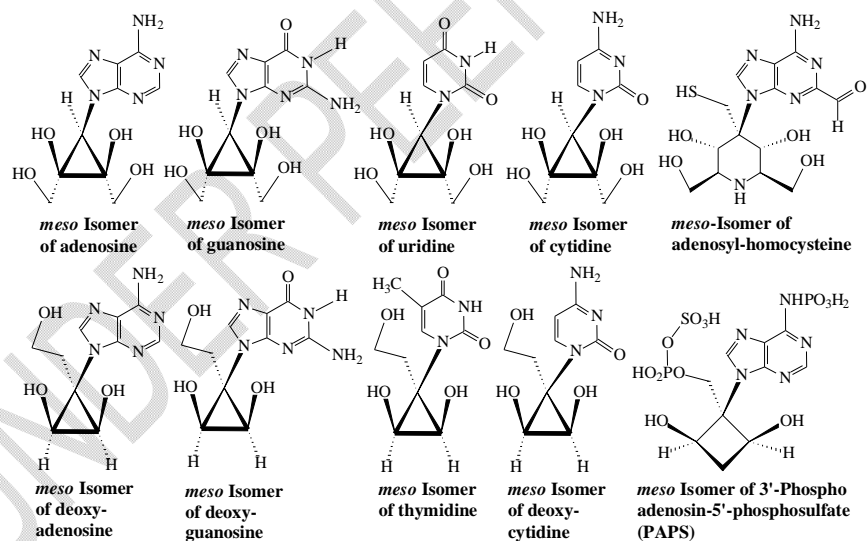


Figure 4. *Meso* isomers of nucleotides, deoxynucleotides, adenosyl-homocysteine and PAPS.

functional groups and by their biochemical role. The association **by this criteria of** amino acids is consecrated and supported by numerous arguments. On the other hand, their relationship within the same molecular formula, although quite obvious, is more discrete. L-Ala is associated with e. g. *cis*-2,4-dihydroxy azetidene due to the same molecular formula,

i.e. a chemical and a philosophical relationship. Although discrete, the relationship *via* molecular formula is undeniable.

3.2. Nucleotides and deoxy-nucleotides. Nucleosides and nucleotides, as the constituents of all types of RNA, and their deoxy counterparts, as constituents of DNA, are represented by their *meso* isomers (Fig. 4); adenosin is cis-3,4-dihydroxy-cis-2,5-dihydroxy-1-adenin cyclopentane. We have also added *meso* isomers of adenosyl homocysteine, a compound involved in methylation reactions, and 3'-phosphoadenosyl-5'-phosphosulfate (PAPS), the major sulfate donor.

3.3. Hydrosoluble vitamins and their coenzymes. Hydrosoluble vitamins represented by thiamine (vitamin B1), isoaloxazine, pyridoxol (vitamin B6), biopterin, pantoic acid, vitamin biotin, nicotinamide have a variety of *meso* isomers. The planar structure of benzenoid compounds has been successfully used in *meso* isomers of hydrosoluble vitamins (Fig. 5): cis-2,4-dihydroxy-3-methyl-3-adenin oxetane (biopterin), cis-2,4-dihydroxy-3-propyl-3-(3,4-diamino-thiophene-2)oxetane (biotin), 2,4-diphosphonate-3-hydroxy-amino-pyrimidine-3-thiazol (thiamine; vitamin B1), 1-hydroxy cis-2,6-diphosphate-cis-3,5-ethyleneglycol-3-dihydroxyisoaloxazine-3-adenin-cyclohexane (vitamin B2) (as FADH₂ and FMNH₂), and even pyridoxol (vitamin B6), coenzyme A and NADH. In order to write *meso* isomer of FMNH₂ we extracted an O atom from a keto bond, however leaving redox system intact. An excellent alternative to this is to link the isoaloxazine system and a phosphonic residue on C-3 of ribitol. A component of coenzyme A, pantoic acid, has tetrahydroxy cyclohexane as a *meso* pair.

3.4. Sterols are represented by a diversity of structures, however all of them present *meso* isomers (Fig. 6). "Sterols have been exemplified by cholesterol, stigmasterol, sitosterol, campesterol, ergosterol and digitoxigenin. Digitoxigenin also presents the four types of isomers. A similar solution has been found for estrone, C₁₉ (5 α -androstanolone), C₂₁ (prednisolone, 11 β -hydroxy-progesterone, pregnenolone, progesterone, corticosterone, cortisol, aldosterone), C₂₄ (biliary acids: cholic, chenodeoxycholic, deoxycholic, lithocholic). Squalene presents at least one *meso* compound" [17].

3.5. Lipophilic vitamins. All lipophilic vitamins – A, D, E, K – present *meso* isomers (Fig. 7). Vitamin E is represented by α -tocopherol and α -tocotrienol, but all members of this vitamin have *meso* isomers, and the same are vitamins K1 and K2. Both *meso* isomers of vitamin K1 and K2 are indicated.

3.6. Fatty acids, sphingosines, prostaglandins. "Saturated, mono- and polyenoic fatty acids are represented by the isomers of stearic acid, oleic and eicosapentaenoic acid (the famous omega-3) (Fig. 8). As is obvious, an isomer of C₁₈H₃₆O₂ (cis-1,3-dihydroxy-cis-4,6-diheptyl-cyclohexane) present all four type of isomers: *meso* (cis-1,3-dihydroxy-cis-4,6-diheptyl-cyclohexane), *CTS* (as pairs of enantiomers) (trans-1,3-dihydroxy-trans-4,6-diheptyl- cyclohexane, etc.), *irrechi* (cis-1,3-dihydroxy-trans-4,6-diheptyl-cyclohexane, etc.) *constit.*, (stearic acid, etc.). A general formula has been elaborated for mono- and polyunsaturated fatty acids" [17]. For long chain bases (LCB) (sphingosines), LCB d18:1 and LCB d18:0 have been selected. *Meso* isomers have been also found for LCB t16:0, LCB d16:0, LCB d16:1, LCB t18:0, LCB t18:1, LCB t20:0, LCB t20:1. *Meso* isomers of saturated LCB should use *meso* isomer of nonanol as a model. All prostaglandins have matching *meso* isomers, as indicated by PGE1, PGF2 α , PGE2, PGF3 α (Fig. 8).

3.7. Aliphatic hydrocarbons: alkanes, alkenes (cycloalkanes), alkynes (alkadienes). A tentative to evaluate molecular diversity of C₈H₁₈ indicated 18 [73] or 19 [72] isomers. If one takes into account optical activity [67], the total number of isomers for C₈H₁₈ is 24. Of these 24, one is *meso*, two are *CTS* [176] (Fig. 9) and the others are *constit.* An unequivocal conclusion can be drawn: all alkanes beginning with C₈H₁₈ present at least one *meso* isomer, and all alkanes below this limit are *archaic*. The first term of C_nH_{2n}, alkenes or cycloalkanes, according to our reasoning, is the *meso* isomer cis-1,2-dimethyl cyclopropane (C₅H₁₀) [177]. For C_nH_{2n-2} (alkynes and alkadienes) the first term is C₇, cis-1,2-dimethyl-3-

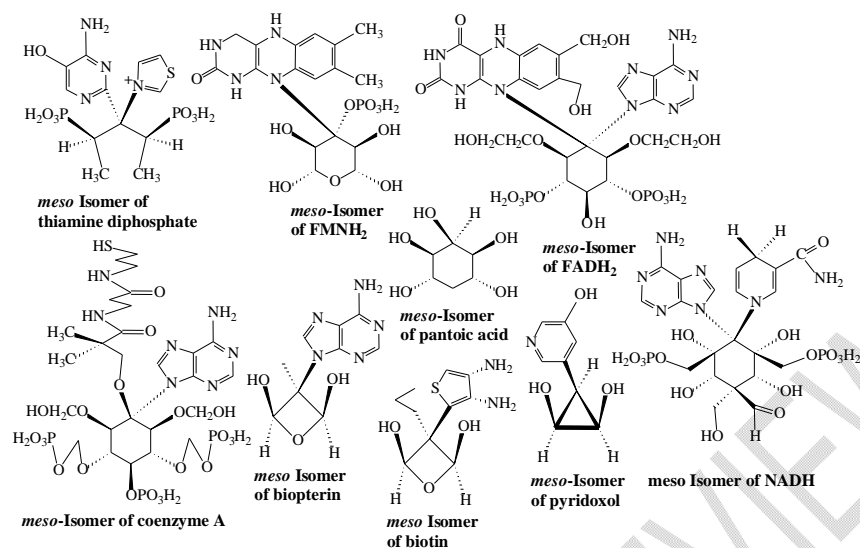


Figure 5. Hydrosoluble vitamins and their natural reagents (FADH₂, FMN, NADH, coenzyme A).

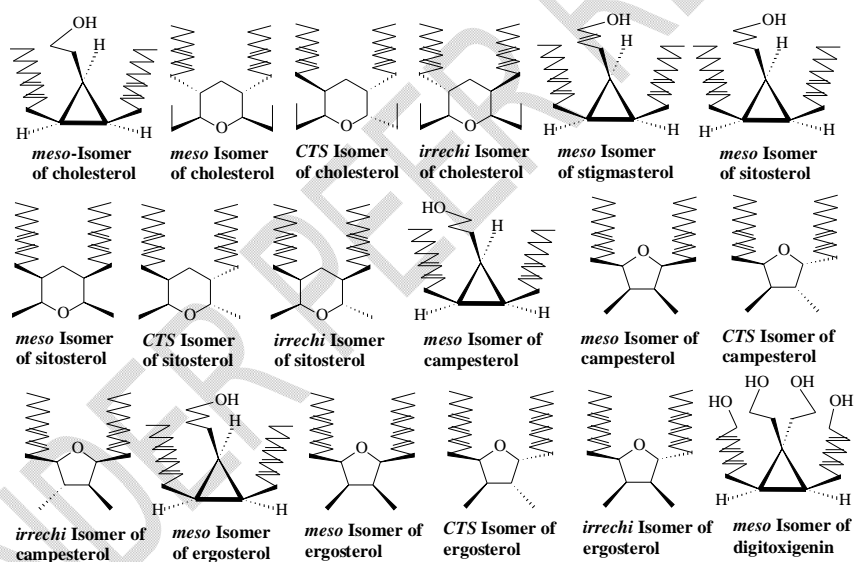


Figure 6. Meso isomers of some natural sterols.

vinyl cyclopropane or cis-1,3-dimethyl-2-methylene cyclobutane; compounds below C₇ are *archaic*.

3.8. Serial compounds with functional groups. For monohydroxylic alcohols the first term is C₉ (3,5-dimethyl-4-hydroxy heptane) (Fig. 10). For aldehydes and ketones the first term is C₅ (cis-1,2-dimethyl-3-hydroxy-cyclopropane), and similar combinations below C₅ are *archaic*. The first term of organic acid is C₃ (cis-1,2-dihydroxy-cyclopropane). C₃ **yet**, as well as C₄ and C₅ have three types of isomers only (*meso*, *CTS*, *constit.*), while C₆ and higher terms possess four; saturated organic acids below C₃ are *archaic*. The first term of monoenoic acids is C₅ (cis-1,2-dihydroxy-3-allyl cyclopropane), and the first term of dienoic acids is C₇ (cis-1,2-dihydroxy-3-(1-butadienyl) cyclopropane).

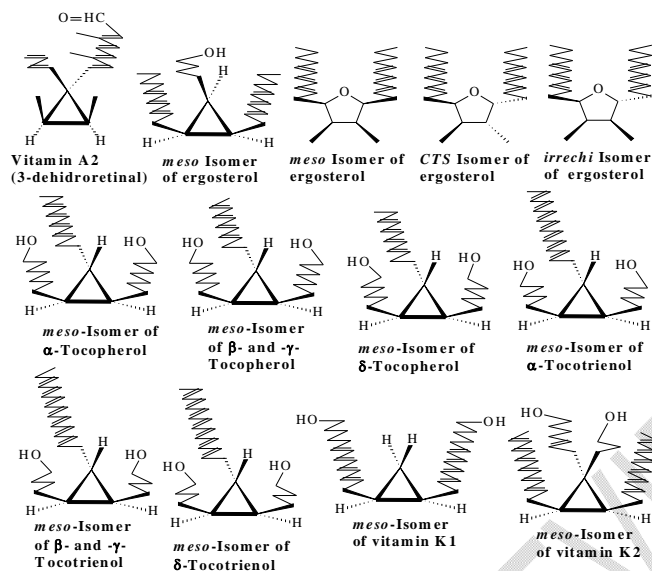


Figure 7. *Meso* isomers of lipophilic vitamins.

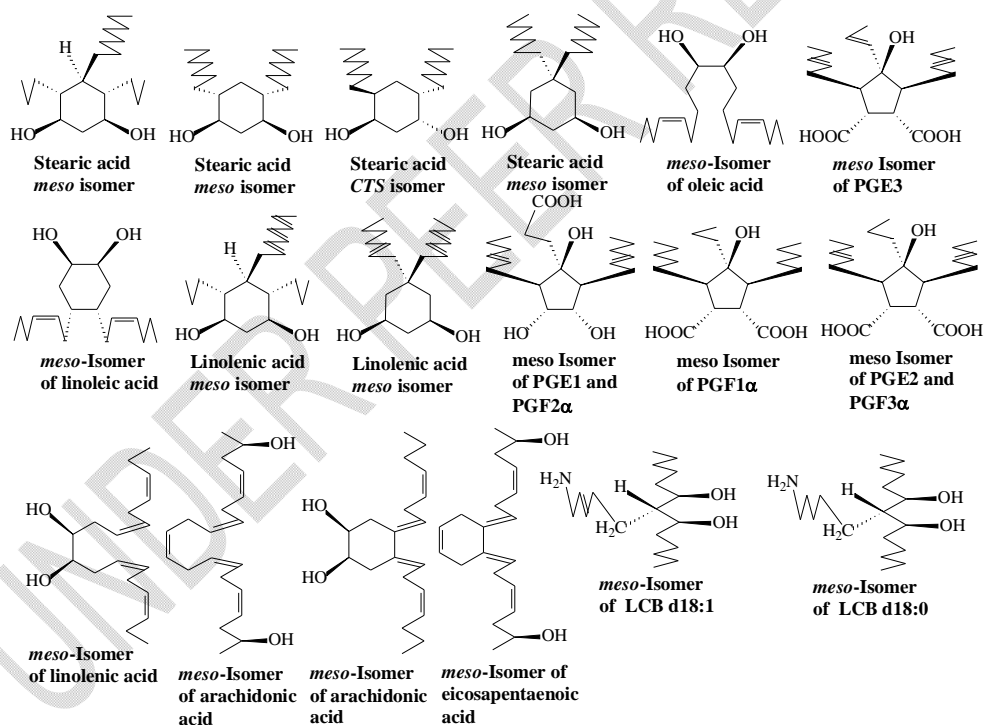


Figure 8. *Meso* isomers of fatty acids, prostaglandins and sphingosines (long chain bases, LCB).

"The following isomers are considered *constit.* isomers of valproic acid (2-propyl pentanoic acid; $C_8H_{16}O_2$): 2-ethyl-3-methyl pentanoic acid, di-isopropyl acetic acid, (R)-2-isopropyl pentanoic acid, (S)-2-isopropyl pentanoic acid, octanoic acid" [178]. According to our systematics, we have to begin with the finding of a $C_8H_{16}O_2$ *meso* isomer. This can be cis-

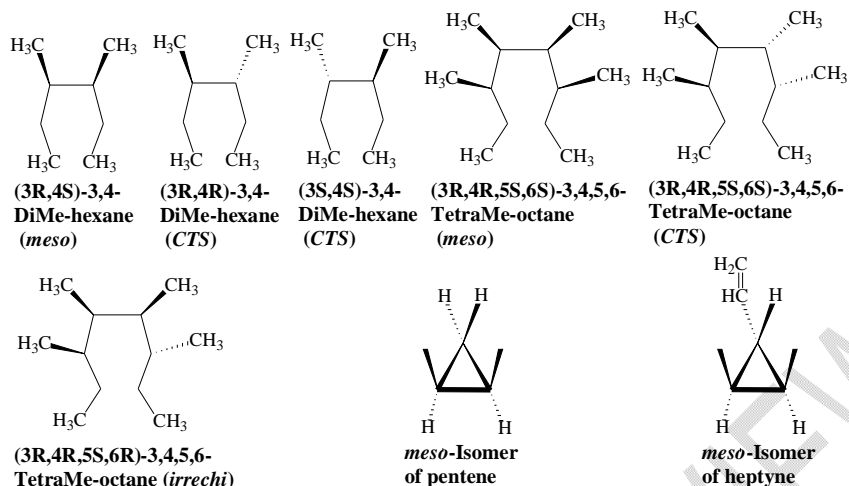


Figure 9. *Meso* isomers of saturated and unsaturated hydrocarbons.

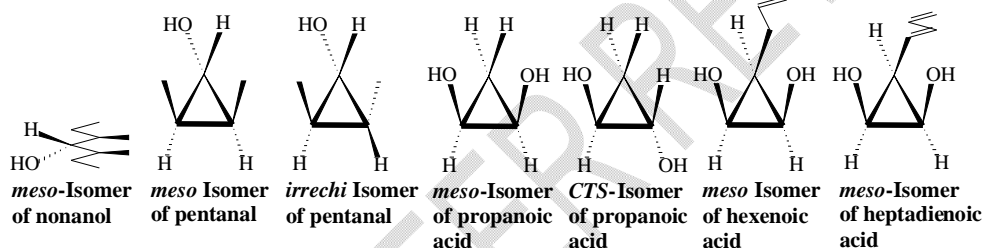


Figure 10. *Meso* isomers of some serial compounds with functional groups.

1,2-dihydroxy-1,2-diethyl-3-methyl cyclopropane, cis-1,3-dihydroxy-2,2-diethyl-cyclobutane, 1 β ,2 β ,3 α ,4 α -1,2-diethyl-3,4-dihydroxy cyclobutane, or 1 β ,3 β ,4 α ,6 α -1,3-dihydroxy-4,6-dimethyl-cyclohexane, or others. As can be seen from their structure, the latter three isomers present also *CTS* and *irrechi* forms. And the C₈H₁₆O₂ isomers mentioned earlier, valproic acid inclusively, are all *constit.*

4. AN EXERCISE OF COMPARATIVE CHEMISTRY GIVES AN ANSWER TO AN UNANSWERED QUESTION – WHY WERE CONSTITUTIONAL ISOMERS FAVORED ?

A question should be raised concerning the hierarchy [62] of the four types of isomers, in other words which of them fills the top place. An intrinsic property of *meso* combinations is their character of dimerism, hence their molecule is formed of two entities that are contrary in a spatial, chemical and optical sense. Of this reason, nine philosophers of ten, probably, would declare *meso* group as being on the top. We ourselves have selected them as structural reference since we thought they have a higher rank than *CTS* and *irrechi*. Nonetheless, that some people could be fascinated by *CTS* molecules, since they are produced by doubling of the same entity. If we compare the four types, it's quite obvious that *meso*, *CTS* and even *irrechi* are characterized by some structural restrictions. *Constit.*, molecules are characterized by the least such structural restrictions. Of this reason, probably, natural chemistry opted for them. The structure of all natural compounds is written in a program, i. e. a genome. Hence this hierarchy is registered in fact in genome, and the

evolution process from plants and microorganisms to vertebrates has been accomplished at this level.

When physical chemistry appeared and developed, biologists and other scholars connected with biomolecules, hoped that physical chemists would discover a marker for natural compounds, as density is for gold. Till now such hope never met, according to our knowledge. Nonetheless, natural combinations possess some unique characteristics, and one of them, in our opinion, is the fact that they are less restricted, in structural sense, than *meso*, *CTS* and *irrechi*. A proof for this assertion is the fact that as soon as a living thing dies, nature sends a thousand messengers to recover its component materials. We reckon that at least one of these characteristics is that *constit.* compounds have a higher number of **structural** freedom degrees, in comparison with the other types. Somehow, this phenomenon is a chemical expression of freedom, inscribed in genome.

In different classes of compounds which constitute series, a limit has been noticed, and above this limit at least *meso* isomers are possible, or even all four types. Compounds under this limit have to be considered as *archaic*. They can reach to the group of combinations able of producing *meso* isomers only by chemical transformations. E. g. propane belongs to *archaic* group, however, by oxidation it becomes propanoic acid, an advanced form able to present *meso* form; however, propanoic acid is again *archaic*. Fischer [58,179,180] illustrated this by preparing a variety of C_6 monosaccharides from formaldehyde or C_3 derivatives.

Natural micromolecular organic combinations can be also classified in a different manner, partially superposing with the afore mentioned classification: (i) symmetric (*meso* and *CTS*); (ii) potential symmetry generators (*irrechi*, *constit.*); (iii) *archaic*. There is yet a vast group of natural compounds, i.e. products of desymmetrization reactions. Nonetheless, they can be integrated in one of the preceding groups (types).

5. C_2 SYMMETRICAL COMPOUNDS OR TWIN DIMERIC CHIRALITY – A NEW TYPE OF CHEMICAL DUALITY

Compounds as trans-3,4-divinyl-1-cyclobutene, trans-1,2-dimethyl-cyclobutane, 1 α ,2 α ,4 β ,5 β -1,2,4,5-tetramethyl cyclohexane (Fig. 1), trans-2,3-dihydroxy aziridine, trans-2,4-dihydroxy azetidene, trans-2,4-dihydroxy-3,3-dimethyl azetidene, 2 β ,3 β ,4 α ,5 α -2,5-dihydroxy-3,4-dimethyl pyrrolidine (Fig. 3), and many other types of compounds – steroids (Fig. 6), lipophilic vitamins (Fig. 7), saturated and polyunsaturated fatty acids, etc., are possibly C_2 symmetrical. Beside, an impressive number of natural C_2 symmetrical compounds is known [39, 181-184]. It seems that the number of C_2 symmetrical compounds is about ten times higher than their *meso* isomers. Hence, there are much unexploited material for chemical philosophy, and some improvement is needed for the present concepts of the so called the science of sciences.

6. CONCLUSIONS

1. At most four types (groups) of isomers have been found, in natural things or as envisaged structures: *meso*, C_2 symmetrical, *irrechi*, *constitutional*.
2. Practically all fundamental natural combinations, found as *constitutional* isomers in vertebrates, are able to form symmetric isomers. Hence, they keep symmetry as a potentiality and not as a reality.
3. An exercise of comparative chemistry is presented between the real *constitutional* isomers and the envisaged *meso* ones.
4. At chemical level symmetry phenomenon is much better represented in plants and microorganisms than in vertebrates.

5. The mirror plane of symmetry has been defined as an area capable to hide (mask) atoms or planar structures of polarized light, and to transform a heterodimer in a homodimer.
6. Two duality phenomena have been identified in chemistry of natural compounds. For one of them the two component sides are opposed chemically, spatially and optically, and they lead to two different (enantiomeric) compounds when distinctively affected.
7. The duality formed of chiral dimers (CTS), uniformly linked with each other or on a more or less complex matrix, constitutes a novelty for chemical philosophy.

REFERENCES

1. Devlin ThM. (ed), Textbook of Biochemistry with Clinical Correlations. New York: Wiley and Sons; 1997.
2. Lieberman MA, Ricer R. Biochemistry, Molecular Biology and Genetics. London: Wolters Kluwer; 2014.
3. King MW. Integrative Medical Biochemistry, London: Mc Graw Hill; 2014.
4. Heisenberg W. Physics and Philosophy, The Revolution in Modern Science. London: George Allen & Unwin; 1958.
5. Feynman RP, Leighton RB, Sands M. The Feynman Lectures on Physics. Mainly Mechanics, Radiation and Heat. London: Addison-Wesley Reading; 1963.
6. Born M. Atomic Physics, London: Blackie & Son Lim; 1969.
7. Qian X, Konthasinghe FK, Manikandan SK, Spiecker D, Vamivakas AN, Eberly JH. Turning off quantum duality. Phys Rev Res. 2020;2: 012016(R).
8. Trapp R. The duality of chemistry: Chemistry for peaceful purposes versus chemical weapons. Pure Appl Chem. 2008;80(8):1763-72.
9. Kaya E, Erduran S. Integrating Epistemological Perspectives on Chemistry in Chemical Education: The Cases of Concept Duality, Chemical Language, and Structural Explanations. Sci Educ. 2013;22:1741-55.
10. Volkova YA, Budynina EM, Kaplun AE, Ivanova OA, Chagarovskiy AO, Skvortsov DA, et al. Duality of Donor–Acceptor Cyclopropane Reactivity as a Three-Carbon Component in Five-Membered Ring Construction: [3+2] Annulation Versus [3+2] Cycloaddition. Chem Eur J. 2013;19:6586-90.
11. Hegel GWF. *Lectures on the History of Philosophy [Geschichte der Philosophie, 1833]*, in three volumes, translated by E.S. Haldane and Frances H. Simson, New Jersey: Humanities Press; 1974.
12. Iga DP. Chitwin Compounds: A New Revelation of Chemistry and Biology. Chem. Res. J. 2018;3:63-79.
13. Iga DP. A New Kind of Symmetry in Chemistry and Biology and a Virtual Mirror Intrinsic to Vegetable Tissues Evidenced by Comparative Structural Analysis of Dochi Compounds. Chem. Res. J. 2020;5:71-91.
14. Iga D P. Carotenoid Structures an Illustration of a New Kind of Symmetry in Chemistry. Chem. Res. J. 2021;6:20-48.
15. Iga DP, Popescu D, Niculescu VIR. On the impact of meso compounds and their isomers: towards a new type of oscillation?. Chem. Res. J. 2022;7:39-48.
16. Iga DP, Popescu D, Niculescu VIR. Dimerization of indole derivatives with hypervalent iodines(III): a new entry for the concise total synthesis of *rac*- and *meso*-chimonanthines. Asian J Chem. Sci. 2022;12(2):14-30.
17. Iga DP. An integrative action based on molecular formula and an exercise of comparative chemistry indicate a relationship of hierarchy and a phenomenon of duality in chemistry. Chem. Res. J. 2022;7(4):64-76.
18. Cahn RS, Ingold C, Prelog V. Specification of Molecular Chirality. Angew Chem Int Ed Eng. 1966;5:385-415.

19. Prelog V, Helmchen G. Basic principles of the CIP-system and proposal for a revision. *Angew. Chem. Int. Ed. Eng.* 1982;21:567-83.
20. Fischer E, Stahel E. Zur Kenntniss der Xylose. *Ber. Deut. Chem. Ges.* 1891;24:528-39.
21. Hilditch TP. *A Concise History of Chemistry*. New York: D Van Nostr Company; 1911.
22. Kendall J. *Great discoveries by young chemists*. New York: Th Y Growell Company; 1953.
23. Kelvin WT Lord. *The molecular tactics of a crystal*. Oxford UK: Clarendon Press; 1894.
24. Prelog V. *Chirality in Chemistry*. *Croat. Chem. Acta.* 2006;79:XLIX-LVII. © The Nobel Foundation 1975 Nobel Lecture December 12; 1975.
25. Cronin J, Reisse J. 3. Chirality and the Origin of Homochirality. In *Lectures in Astrobiology*. Gargaud M, Barbier B, Martin H, Reisse J, eds. London: Springer-Verlag Vol 1 pp 73-114; 2005.
26. Fischer E, Hertz J. Reduction der Schleimsäure. *Ber deut chem Ges.* 1892;25:1247-61.
27. Woo S, Keay BA. "SN2' and "SN2' Like" Ring Openings of Oxa-n-Cyclo Systems. *Synthesis.* 1996;7:669-86.
28. Hoffmann RW. *meso* Compounds: Stepchildren or Favored Children of Stereoselective Synthesis? *Angew Chem. Int. Ed. Eng.* 2003;42:1096-109.
29. Trost BM, Crawley ML. Asymmetric Transition-Metal-Catalyzed Allylic Alkylations: Applications in Total Synthesis. *Chem. Rev.* 2003;103:2921-43.
30. Trost BM, Van Vranken DL. Asymmetric Transition Metal-Catalyzed Allylic Alkylations. *Chem. Rev.* 1996;96:395-422.
31. Wang M, Feng M, Tang B, Jiang X. Recent advances of desymmetrization protocol applied in natural product total synthesis. *Tetrahedr. Lett.* 2014;55:7147-55.
32. Kagan HB, Dang TP. Asymmetric Catalytic Reduction with Transition Metal Complexes. I. A Catalytic System of Rhodium (I) with (-)-2,3-(9-Isopropylidene-2,3-dihydroxy-1,4-bis(diphenylphosphino) butane, a New Chiral Diphosphine. *J. Am. Chem. Soc.* 1972;94:6429-33.
33. Whitesell JK. C2 symmetry and asymmetric induction. *Chem. Rev.* 1989;89:1581-90.
34. Reusch W. *Virtual textbook of organic chemistry*. Department of Chemistry, Michigan State University, Michigan: East Lansing; 2011.
35. Fischer E, Hirschberger J. Ueber Mannose. IV. *Ber. deut. chem. Ges.* 1889;22:3218-24.
36. Fischer E, Fay IW. Ueber Idonsäure Idose Idit und Idozuckersäure. *Ber deut chem Ges.* 1895;28:1975-83.
37. Fischer E, Tafel J. Oxydation des Glycerins. *Ber. deut. chem. Ges.* 1888;21(2): 2634-37.
38. Fischer E. Configuration der Weinsäure. *Ber deut chem Ges.* 1896;29:1377-83.
39. Kang EJ, Lee E. Total Synthesis of Oxacyclic Macrodiolide Natural Products. *Chem Rev.* 2005;105:4348-78.
40. Jaeger FM. *Lectures on the Principle of Symmetry and its Applications in All Natural Sciences*. Amsterdam: Elsevier Publishing Co; 1917.
41. Vickery HB. Assignment of D L prefixes to the tartaric acids. *J. Chem. Ed.* 1957;34:339-41.
42. Trost BM, Shi Z. From furan to nucleosides. *J. Am. Chem. Soc.* 1996;118:3037-38.
43. Pfaltz A, Drury III WJ. Design of chiral ligands for asymmetric catalysis: From C2-symmetric P,P- and N,N-ligands to sterically and electronically nonsymmetrical P,N-ligands. *Proc. Natl. Acad. Sci. USA.* 2004;101:5723-26.
44. Pigman W. *The Carbohydrates: Chemistry, Biochemistry, Physiology*. New York: Academic Press; 1957.
45. Finar IL. *Organic Chemistry*. Vol 2, London: Longmans Green and Co Ltd; 1964.
46. Derewenda ZS. On wine chirality and crystallography. *Acta Cryst A.* 2008;64:246-58.
47. Wisniak J. Carl Wilhelm Scheele Rev. *CENIC Cienc. Quím.* 2009;403:165-73.
48. Svedberg G. A Tribute to the Memory of Carl Wilhelm Scheele (1742-1786) Presented at the 2012 Annual Meeting of the Royal Swedish Academy of Engineering Sciences

Royal Swedish Academy of Engineering Sciences (IVA) Editor: Anna Lindberg. Stockholm Sweden: IVA Kaigan AB; 2012.

49. van 't Hoff JH. A suggestion looking to the extension into space of the structural formulas at present used in chemistry And a note upon the relation between the optical activity the chemical constitution of organic compounds. Arch. Neerland. Sci. Nat. 1874;9:445-54.
50. Le Bel JA. Sur les relations qui existent entre les formules atomiques des corps organiques et le pouvoir rotatoire de leurs dissolutions. Bull Soc Chim France. 1874;22:337-47.
51. Hoffmann R, Laszlo P. Representation in Chemistry. Angew Chem. 1991;30:1-16.
52. Bijvoet JM, Peerdemann AF, van Bommel AJ. Determination of the absolute configuration of optically active compounds by means of X-rays. Nature. 1951;168:271-72.
53. Wohl A, Momber Fr. Die sterische Beziehung zwischen Glycerinaldehyd und Weinsäure. Ber. Deut. Chem. Ges. 1917;50:455-62.
54. Klyne W, Buckingham J. Atlas of Stereochemistry Absolute Configurations of Organic Molecules. Vol 1, London: Chapman and Hall; 1978.
55. Baer E, Fischer HOL. Studies on acetone-glyceraldehyde. IV. Preparation of D-(+)-acetone glycerol. J Biol Chem. 1939;128:463-73.
56. Baer E, Fischer HOL. Studies on acetone-glyceraldehyde. VII. Preparation of L-glyceraldehyde and L-(-)-acetone glycerol. J Am Chem Soc. 1939;61:761-65.
57. Iga DP. Basic Principles of the Strategy Concerning the Elucidation of Configuration of Chiral Centers of Linear Isomeric Aldohexoses. Found. Chem. 2018;20:31-41.
58. Fischer E. Synthesen in der Zuckergruppe II. Ber Deut Chem Ges. 1894;27:3189-232.
59. Bragg WL, Bragg WH. The diffraction of short electromagnetic waves by a crystal. Proc Roy Soc London Ser A. 1913;89:248-91.
60. Pauling L. The Shared-Electron Chemical Bond. Proc. Natl. Acad. Sci. USA 1928;14:359-62.
61. Pauling L. The Nature Of The Chemical Bond. Application Of Results Obtained From The Quantum Mechanics And From A Theory Of Paramagnetic Susceptibility To The Structure Of Molecules. J. Am. Chem. Soc. 1931;53:1367-400.
62. Fujita S. Chirality and RS-Stereogenicity as Two Kinds of Handedness Their Aufheben by Fujita's Stereoisogram Approach for Giving New Insights into Classification of Isomers. Bull Chem Soc Jpn. 2016a;89:987-1017.
63. Roberts JD, Caserio MC. Basic Principles of Organic Chemistry. Amsterdam: W A Benjamin Inc; 1977.
64. Schmid GH, Organic Chemistry. London: Mosby; 1996.
65. Yurkanis Bruice P. Organic Chemistry. 4th edition Prentice Hall: College Div; 2003.
66. Smith MB, March J. March's Advanced Organic Chemistry: Reactions, Mechanisms, and Structure. New York: Wiley; 2007.
67. McMurry J. Organic Chemistry. London: Thomson Learning; 2008.
68. Clayden J, Greeves N, Warren S. Organic Chemistry. Second Edition, Oxford UK: Oxford University Press; 2012.
69. Nonell S, Arbogast JW, Foote CS. Production of Fullerene C₆₀ Radical Cation by Photosensitized Electron Transfer. J. Phys. Chem. 1992; 96:4169-70.
70. Rassat A, Laszlo I, Fowler PW. Topological rotational strengths as chirality descriptors for fullerenes. Chem - A Eur J. 2003;9:644-51.
71. Finar IL. Organic Chemistry. Vol 1, London: Longmans Green and Co Ltd; 1963.
72. Fujita S. Half-Century Journey from Synthetic Organic Chemistry to Mathematical Stereochemistry through Chemoinformatics. Iran J Mathem Chem. 2016;7:155-221.
73. Polya G. Kombinatorische Anzahlbestimmungen für Gruppen Graphen und chemische Verbindungen. Acta Mathem. 1937;68:145-254.

74. Metzler DE, Metzler CM. Biochemistry: the chemical reactions of living cells. Amsterdam: Elsevier; 2003.
75. Pasteur L. Sur la transformation des acides tartriques en acide racémique. Découverte de l'acide tartrique inactif. Nouvelle méthode de séparation de l'acide racémique en acides tartriques droit et gauche. C R Séances Acad Sci. 1853;37:162-66.
76. Wagner G, Yang JC, Loewus FA. Stereoisomeric Characterization of Tartaric Acid Produced during L-Ascorbic Acid Metabolism in Plants. Plant Physiol. 1975;55:1071-73.
77. Meldola, R. The Chemical Synthesis of Vital Products and the Inter-Relations Between Organic Compounds, Vol. I. London: Edward Arnold; 1904.
78. Zhang L, Guo Z, Chen J, Xu Q, Lin H, Hu K, et al. Mechanism of 2,3-butanediol stereoisomers formation in a newly isolated *Serratia* sp. T241. Scient Rep. 2016;19257, 6:1-12.
79. Pigman WW, Goepf Jr RM. Chemistry of the Carbohydrates. New York: Academic Press Inc; 1948.
80. Hough L, Stacey BE. Variation in the allitol content of *Itea* plants during photosynthesis. Phytochem. 1966;5:171-75.
81. Lewis DH, Smith DC. Sugar alcohols (polyols) in fungi and green plants. II, Methods of detection and quantitative estimation in plant extracts. New Phytol. 1967;66:185-204.
82. Podwysotzki WV. Adonis vernalis adonitol. Arch Pharm. 1889;141:227-30.
83. Fischer E. Ueber Adonit einen neuen Pentit. Ber Deut Chem Ges. 1893;26(1):633-639.
84. Schmidt RR, Lieberknecht A. Funktionelle D- and L-ribose-derivate über eine racematspaltung mit rückführung. Angew. Chem. 1978;90:821-22.
85. Kiely DE, Hash KR, Sr. Method of oxidation using nitric acid. US 7692041 B2. 2010.
86. Levy DE, Fügedi P, (eds.) The Organic Chemistry of Sugars. London: Taylor and Francis; 2006.
87. Liu C, Zhong SM, Chen RY, Wu Y, Zhu XJ. Two new compounds from the dried tender stems of *Cinnamomum cassia*. J. Asian Nat. Prod. Res. 2009;11(9):845-49.
88. Fischer E, Passmore F. Ueber kohlenstoffreichere Zuckerarten aus d Mannose. Ber. deut. chem. Ges. 1890;23:2226-39.
89. Hann RM, Maclay WD, Knauf AE, Hudson CS. Relations between Rotatory Power and Structure in the Sugar Group XXXI The Configuration of D- α -Mannooctose D-Manno-L-manno-octose. J. Am. Chem. Soc. 1939;61:1268-69.
90. Hudson CS. Emil Fischer's Discovery of the Configuration of Glucose. J Chem Ed. 1941;18:353-357.
91. Hirs CHW, Moore S, Stein, WH. The Chromatography of Amino Acids on Ion Exchange Resins. Use of Volatile Acids for Elution. J. Am. Chem. Soc. 1954;76:6063-65.
92. Work E, Birnbaum SM, Winitz M, Greenstein JP. Separation of the three isomeric components of synthetic α,ϵ -diaminopimelic acid. J Am chem Soc. 1955;77(7):1916-18.
93. Hoare DS, Work, E. The Stereoisomers of $\alpha\epsilon$ -Diaminopimelic Acid: their Distribution in Nature and Behaviour towards certain Enzyme Preparations. Biochem. J. 1957;65:441-447.
94. Meadow PM, Work E. Biosynthesis of diaminopimelic acid and lysine in *Escherichia coli*. I. The incorporation of ^{14}C from various organic precursors into the diaminopimelic acid of a lysine-requiring mutant. Biochem J. 1959;72:396-400.
95. Richaud C, Higgins W, Mengin-Lecreulx D, Stragier P. Molecular Cloning, Characterization, and Chromosomal Localization of *dapF*, the *Escherichia coli* Gene for Diaminopimelate Epimerase. J Bacteriol. 1987; 169(4):1454-59.
96. Uehara A, Fujimoto Y, Kawasaki A, Kusumoto S, Fukase K, Takada, H. *Meso*-Diaminopimelic acid and *meso*-lanthionine, amino acids specific to bacterial peptidoglycans, activate human epithelial cells through NOD1. J Immunol. 2006;177:1796-804.
97. Brown GB, du Vigneaud V. The stereoisomeric forms of lanthionine. J Biol Chem. 1941;140:767-71.

98. Kellner R, Jung G, Horner T, Zahner H, Schnell N, Entian KD, Gotz F. Gallidermin: a new lanthionine-containing polypeptide antibiotic. *Eur J Biochem.* 1988;177:53-59.
99. Chiku T, Padovani D, Zhu W, Singh S, Vitvitsky V, Banerjee, R. H₂S Biogenesis by Human Cystathionine γ -Lyase Leads to the Novel Sulfur Metabolites Lanthionine and Homolanthionine and Is Responsive to the Grade of Hyperhomocysteinemia. *J Biol Chem.* 2009;284(17):11601-12.
100. Nonappa K, Ahonen M, Lahtinen F, Kolehmainen E. Cyclic dpeptides: catalyst/promoter-free, rapid and environmentally benign cyclization of free aminoacids. *Green Chem. Issue 5* (2011).
101. Garg NK, Stoltz BM. The formal total synthesis of dragmacidin B, trans-dragmacidin C, and cis- and trans-dihydrohamacanthins A. *Tetrahedr. Lett.* 2005;46:2423-26.
102. Kozlovsky AG, Vinokurova NG, Adanin VM, Burkhardt G, Dahse HM, Grafe U. New diketopiperazine alkaloids from *Penicillium fellutanum*. *J Nat Prod.* 2000;63:698-700.
103. Yang SW, Chan TM, Terracciano J, Loebenberg D, Chen G, Patel M, et al. Structure Elucidation of a New Diketopiperazine Sch 725418 from *Micromonospora* sp. *J Antibiot.* 2004;57:345-47.
104. Kozlovsky AG, Zhelifonova VP, Antipova TV. Biologically active metabolites of *Penicillium* fungi. Signpost Open Access J. Org. Biomol. Chem. 2013;1:11-21. Article ID 010302.
105. Li SM. Melanin Biosynthesis Inhibitors from the Bark of *Machilus thunbergii*. *Nat Prod Rep.* 2010;27:57-78.
106. Overman LE, Paone DV, Stearns BA. Direct Stereo- and Enantiocontrolled Synthesis of Vicinal Stereogenic Quaternary Carbon Centers Total Syntheses of meso- and (-)-Chimonanthine and (+)-Calycanthine. *J. Am. Chem. Soc.* 1999;121:7702-03.
107. Ishikawa H, Takayama H, Aimi N. Dimerization of indole derivatives with hypervalent iodines(III): a new entry for the concise total synthesis of rac- and meso-chimonanthines. *Tetrahedr Lett.* 2002;43:5637-39.
108. Barbeau K, Zhang G, Live DH, Butler A. Petrobactin, a Photoreactive Siderophore Produced by the Oil-Degrading Marine Bacterium *Marinobacter hydrocarbonoclasticus*. *J Am Chem Soc.* 2002;124:378-79.
109. Bergeron RJ, Huang G, Smith RE, Bharti N, McManis JS, Butler A. Total synthesis and structure revision of petrobactin. *Tetrahedron*, 2003;59:2007-14.
110. Yun BS, Ryoo IJ, Kim WG, Kim JP, Koshino H, Seto H, et al. Structures of phenazostatins A and B, neuronal cell protecting substances of microbial origin. *Tetrahedron Lett.*, 1996;37:8529-30.
111. Maskey RP, Kock I, Helmke E, Laatsch H. Isolation and Structure Determination of Phenazostatin D, a New Phenazine from a Marine Actinomycete Isolate *Pseudonocardia* sp. B6273. *Z Naturforsch.* 2003;58b:692-94.
112. Rüttimann A, Mayer H. Synthesis of optically active natural carotenoids and structurally related compounds. V: synthesis of (3R,3'R)-, (3S, 3'S)-, and (3R,3'S meso)-zeaxanthin by asymmetric hydroboration: a new approach to optically active carotenoid building units. *Helv Chim Acta.* 1980;63:1456-62.
113. Rüttimann A, Schiedt K, Vecchi, M. Separation of (3R,3'R)-, (3R,3'S; meso)-, (3S,3'S)-zeaxanthin, (3R,3'R,6'R)-, (3R,3'S,6'S)- and (3S,3'S,6'S)-lutein via the dicarbamates of (S)-(+)- α -(1-naphthyl) ethyl isocyanate. *J High Resol Chromatogr Chromatogr Commun.* 1983;6:612-16.
114. Maoka T, Arai A, Shimizu M, Matsuno T. *Comp Biochem Physiol.* The first isolation of enantiomeric and meso-zeaxanthin in nature. 1986;83B:121-24.
115. Thurnham DI. Macular zeaxanthins and lutein – a review of dietary sources and bioavailability and some relationships with macular pigment optical density and age-related macular disease. *Nutr Res Rev.* 2007;20:163-79.

116. Kayser H, Aareskjöld, K. Borch G, Liaaen-Jensen S. Partly racemized 2-hydroxy- β -type carotenoids from the insects *Cerura vinula* and *Ectatosoma tiaratum*. *Insect Biochem.* 1984;14:51-54.
117. Matsuno T, Yamashita E. Tunaxanthin stereoisomers. *Nippon Suisan Gakkaishi*, 1989; 55:1667.
118. Bingham A Jr, Wilkie DW, Mosher HS. Tunaxanthin: occurrence and absolute stereochemistry. *Comp Biochem Physiol B.* 1979;62:489-95.
119. Ikuno Y, Shimizu M, Koshino Y, Maoka T, Matsuno T. Stereochemical investigation of carotenoids from yellow-tail rockfish *Sebastes flavidus*. *Bull Jap Soc Sci Fish.* 1985;51:2033-35.
120. Matsuno T, Katsuyama M, Ikuno Y, Yamashita E, Ha BS. The occurrence of eight stereoisomers of Tunaxanthin from the fresh-water fish *Siniperca scherzeri*. *Bull Jap Soc Sci Fish.* 1990;56(4):651-54.
121. Vecchi M, Englert G, Mayer, H. Chromatographische Trennung und Identifizierung diastereomerer Carotinoide mit grossem räumlichem Abstand der chiralen Zentren. *Helv Chim Acta.* 1982; 65:1050-58.
122. Ikuno Y, Maoka T, Shimizu M, Komori T, Matsuno T. Direct diastereomeric resolution of carotenoids. II. All ten stereoisomers of tunaxanthin (ϵ,ϵ -carotene-3,3'-diol). *J Chromatogr.* 1985;328:387-91.
123. Ronneberg H, Renstrom B, Aareskjold K, Liaaen-Jensen S, Vecchi M, Leuenberger FJ, et al. Naturally occurrence of enantiomeric and *meso*-astaxanthin 1. Ex lobster eggs (*Homarus gammarus*). *Helv Chim Acta.* 1980;63:711-15.
124. Matsuno T, Maoka T, Katsuyama M, Ookubo M, Katagiri K, Jimura H. The occurrence of enantiomeric and *meso*-astaxanthin in aquatic animals. *Nippon Suisan Gakkaishi* 1984;50:1589-92.
125. Andrewes AG, Kjoson H, Liaaen-Jensen S, Weisgraber KH, Lousberg RJJ, Weiss U. Animal Carotenoids. 7. Carotenes of Two Colour Variants of the Aphid *Macrosiphum liriodendri* – Identification of Natural γ,γ -Carotene. *Acta Chem Scand B.* 1971;25(10):3878-80.
126. Morimoto Y, Iwai T, Kinoshita T. Effective Combination of Two-Directional Synthesis and Rhenium(VII) Chemistry: Total Synthesis of *meso* Polyether Teurilene. *J. Am. Chem. Soc.* 1999;121:6792-97.
127. Brocks JJ, Love GD, Summons RE, Knoll AH, Logan GA, Bowden SA. Biomarker evidence for green and purple sulphur bacteria in a stratified Palaeoproterozoic sea. *Nature.* 2005;437:866-70.
128. Brocks JJ, Schaeffer P. Okenane, a biomarker for purple sulfur bacteria (Chromatiaceae), and other new carotenoid derivatives from the 1640 Ma Barney Creek Formation. *Geochim Cosmochim Acta.* 2008;72:1396-414.
129. Smith JHC. Carotene: III. Hydrogenation and Optical Properties of Carotene And its Hydrogenated Derivatives. *J Biol Chem.* 1931;90:597-605.
130. Karrer P, Jucker E. Carotenoids, Amsterdam: Elsevier; 1950.
131. Koopmans MP, Schouten IS, Kohlen LMEL, Sinninghe Damste JS. Restricted utility of aryl isoprenoids as indicators for photic zone anoxia. *Geochim Cosmochim Acta.* 1996;60(23):4873-76.
132. Schouten S, Sinninghe Damste JS, De Leeuw JW. A novel triterpenoid carbon skeleton in immature sulphur-rich sediments. *Geochim Cosmochim Acta.* 1995;59(5):953-58.
133. Schwarzbauer J, Jovančićević B. Main Types of Organic Matter in Geosphere. In: Fossil Matter in the Geosphere. Fundamentals in Organic Geochemistry. Cham: Springer; 2015.
134. Cho M-H, Moinuddin SGA, Helms GL, Hishiyama S, Eichinger D, Davin LB et al. (+)-Larreatricin hydroxylase an enantio-specific polyphenol oxidase from the creosote bush (*Larrea tridentata*). *Proc Natl Acad Sci USA.* 2003;100:10641-46.

135. Li G, Ju HK, Chang HW, Jahng Y, Lee SH, Son JK. Melanin Biosynthesis Inhibitors from the Bark of *Machilus thunbergii*. Biol Pharm Bull. 2003;26(7):1039-41.
136. Miyazawa M, Okuno Y, Oshiro K, Kasahara H, Shimamura H, Nakamura SI, Kameoka H. Suppression of the SOS-Inducing Activity of Trp-P-1 and Aflatoxin B1 by Meso-dihydroguaiaretic Acid from *Machilus thunbergii* in the *Salmonella typhimurium* TA1535/pSK1002 *umu* Test. Biosci Biotechnol Biochem. 1998;62(7):1425-27.
137. Xiao W, Peng B, Peng Y, Xiao PG. Advances in studies on *Saururus chinensis*. Chin Tradit Herb Drugs. 2010;41:12-15.
138. Ikeya Y, Taguchi H, Yosioka I. The constituents of *Schisandra chinensis* Baill. The structures of two new lignans, pre-gomisin and Gomisin J Chem Pharm Bull. 1978;26(2):682-84.
139. Xue YB, Zhang YL, Yang JH, Du X, Pu JX, Zhao W, et al. Nortriterpenoids and lignans from the fruit of *Schisandra chinensis*. Chem Pharm Bull. 2010;58(12):1606-11.
140. Yamauchi S, Masuda T, Sugahara T, Kawaguchi Y, Ohuchi M, Someya T, et al. Antioxidant Activity of Butane Type Lignans, Secoisolariciresinol, Dihydroguaiaretic Acid, and 7,7'-Oxidihydroguaiaretic Acid. Biosci Biotechnol Biochem. 2008;72(11):2981-86.
141. Li G, Lee CS, Woo MH, Lee SH, Chang HW, Son JK. Lignans from the Bark of *Machilus thunbergii* and Their DNA Topoisomerases I and II Inhibition and Cytotoxicity. Biol Pharm Bull. 2004;27(7):1147-50.
142. Nguyen PH, Le TVT, Kang HW, Chae J, Kim SK, Kwon KI, et al. AMP-activated protein kinase (AMPK) activators from *Myristica fragrans* (nutmeg) and their anti-obesity effect. Bioorg Med Chem Lett. 2010;20:4128-31.
143. Hughes GK, Ritchie, E. The chemical constituents of *Himantandra* species. I. The Lignins of *Himantandra baccata* Bail. and *H. belgraveana* F. Muell. Austral. J. Chem. 1954;7(1):104-12.
144. Blears JG, Haworth RD. The constituents of natural phenolic resins. 23. A synthesis of galgravin. J Chem Soc. 1958; 1985-87.
145. Park BY, Min BS, Kwon OK, Oh SR, Ahn KS, Kim TJ, et al. Increase of Caspase-3 activity by lignans from *Machilus thunbergii* in HL-60 cells. Biol Pharm Bull. 2004;27(8):1305-07.
146. Cui H, Xu B, Wu T, Xu J, Yuan Y, Gu Q. Potential Antiviral Lignans from the Roots of *Saururus chinensis* with Activity against Epstein-Barr Virus Lytic Replication. J Nat Prod. 2014;77:100-10.
147. Fang JM, Lee CK, Cheng YS. Lignans from Leaves of *Juniperus chinensis*. Descr: meso-secoisolariciresinol. Phytochem. 1992;31(10):3659-61.
148. Sugahara T, Yamauchi S, Kondo A, Ohno F, Tominaga S, Nakashima Y, et al. First stereoselective synthesis of meso-secoisolariciresinol and comparison of its biological activity with (+)- and (-)-secoisolariciresinol Biosci. Biotechnol. Biochem. 2007;71:2962-68.
149. Ruolin Y, Zhonghong Y, Congying C, Jianhua L. Constituents and Activities of *Acorus tatarinowii*. Med Res Arch. 2017;5(7):1-14.
150. Davis RA, Caroll AR, Duffy S, Avery VM, Guymer GP, Forster PI, et al. Endiandrin A, a potent glucocorticoid receptor binder isolated from the Australian plant *Endiandra anthropophagorum*. J Nat. Prod. 2007;70(7):1118-21.
151. Davis RA, Barnes EC, Longden J, Avery VM, Healy PC. Isolation, structure elucidation and cytotoxic evaluation of endiandrin B from the Australian rainforest plant *Endiandra anthropophagorum*. Bioorg Med Chem. 2009;17(3):1387-92.
152. Yamamura S, Niwa M, Yukimasa T, Nonoyama M. The isolation and structures of novel neolignans and neosessquilignans from *Heterotropa takaai* M. Bull Chem Soc Jpn. 1982;55:3573-79.

153. Li CC, Wang TL, Zhang ZQ, Yang WQ, Wang YF, Chai X, et al. Phytochemical and Pharmacological Studies on the Genus *Psoralea*: A Mini Review. *Evid-Based Complem Altern Med*. Volume 2016, Article ID 8108643.
154. Wang YH, Morris-Natschke SL, J. Yang, Niu HM, Long CL, Lee KH. Anticancer Principles from Medicinal *Piper* (胡椒 Hú Jiāo) Plants. *J Tradit Complem Med*. 2014;4(1):8-16.
155. Mallette JR, Casale JF. Rapid determination of the isomeric truxillines in illicit cocaine via capillary gas chromatography/flame ionization detection and their use and implication in the determination of cocaine origin and trafficking routes. *J Chromatogr A*. 2014;1364:234-40.
156. Fabricant DS, Nikolic D, Lankin DC, Chen SN, Jaki BU, Kronic A, et al. Cimipronidine, a Cyclic Guanidine Alkaloid from *Cimicifuga racemosa*. *J Nat Prod*. 2005;68(8):1266-70.
157. Martin TS, Kikuzaki H, Hisamoto M, Nakatani N. Constituents of *Amomum tsao-ko* and Their Radical Scavenging and Antioxidant Activities. *J Am Oil Chem Soc*. 2000;77:667-73.
158. Uehara SI, Yasuda I, Akiyama K, Morita H, Takeya K, Itokawa H. Diarylheptanoids from the rhizomes of *Curcuma Xanthorrhiza* and *Alpinia officinarum*. *Chem Pharm Bull*. 1987;35:3298-304.
159. Tabata N, Sunazuka T, Tomoda H, Nagamitsu T, Iwai Y, Omura, S. Diolmycins, new anticoccidial agents produced by *Streptomyces* sp. II. Structure elucidation of diolmycins A1, A2, B1 and B2, and synthesis of diolmycin A1. *J Antibiot*. 1993;46(5):762-69.
160. Nicolaou KC, Gray DLF. Total Synthesis of Hybocarpone and Analogues Thereof A Facile Dimerization of Naphthazarins to Pentacyclic Systems. *J. Am. Chem. Soc*. 2004;126:607-12.
161. Kinoshita K, Usuniwa Y, Yamamoto Y, Koyama K, Takahashi K. Red pigments from the cultured mycobiont of a lichen, *Sphaerophorus fragilis* (L.) Pers. *Lichenol*. 2009;8(1):1-4.
162. Dragan SV, Borisova KL, Pelageev DN, Anufriev VPh. Concerning the Stereoselectivity of the Oxidative Dimerization of 3-Alkyl-2-Hydroxy-1,4-Naphthoquinones in the Synthesis of Hybocarpone. *Nat Prod. Commun*. 2019:1-5.
163. Choudhary A, Naughton LM, Montánchez I, Dobson ADW, Rai DK. Current Status and Future Prospects of Marine Natural Products (MNPs) as Antimicrobials. *Mar Drugs*. 2017;15:1-42.
164. Li WDZ, Ma BC. A Simple Biomimetic Synthesis of dl-Chamaejasmine, a Unique 3,3'-Biflavanone. *Org Lett*. 2005;7(2):271-74.
165. Chang HS, Chen, IS. Chemical constituents and bioactivity of Formosan lauraceous plants. *J Food Drug Anal*. 2016;24:247-63.
166. Nocquet PA. Vers la synthèse d'une nouvelle classe d'iminosucre conformationnellement contraints: ouverture d'azétidines cyclisation 4-exo-trig et C-H amination catalytique. *Autre. Université de Strasbourg, Français NNT: 2013STRAF047*; 2013.
167. Pfaltz A. Design of Chiral Ligands for Asymmetric Catalysis: from C₂-Symmetric Semicorrins and Bisoxazolines to Non-Symmetric Phosphinooxazolines. *Acta Chem. Scand*. 1996;50:189-94.
168. Ghosh AK, Mathivanan P, Cappiello J. C₂-Symmetric chiral bis(oxazoline)-metal complexes in catalytic asymmetric synthesis. *Tetrahedr. Asymm*. 1998;9:1-45.
169. Bach RD, Dmitrenko O. The Effect of Geminal Substitution on the Strain Energy of Dioxiranes The Origin of the Low Ring Strain of Dimethyldioxirane. *J Org Chem*. 2002;67:3884-96.
170. Ellis AV, Kannangara GSK, Wilson MA. Chemistry of Sodium Lactate Formation under Simulated Alumina Refinery Conditions. *Ind. Eng. Chem. Res*. 2003;42:3185-89.

171. Wilson MA, Kannangara GSK, Ellis AV. Carbohydrate rearrangements in humic solutions. In Combined national conference of the Australian Organic Geochemists and the International Humic Substances Society. 16-19 February, Blue Mountains, New South Wales, Australia, Cameron McIntyre ed. Published by CSIRO Petroleum, Australia; 2004.
172. Mendkovich AS, Leibzon VN, Mairanovski SG, Krayushkin MM, Klimova TA, Novikov SS et al. Electroreduction of polyhedrane derivatives 1 Structural effect of keto derivatives of bicyclo[3.3.1]nonane adamantane and noradamantane on electrochemical reduction. Russ. Chem. Bull. 1978;27:1639-43.
173. Steiner GW, Strobel GA. Helminthosporoside a Host-specific Toxin from *Helminthosporium sacchari*. J. Biol. Chem. 1971;246:4350-57.
174. Asselineau C, Asselineau J, Laneelle G, Laneelle MA. The biosynthesis of mycolic acids by mycobacteria. Curr Alternat Hypoth Prog Lipid Res. 2002;41:501-23.
175. Skrela BC. Synthesis and Coordination Chemistry of New Multidentate Ligands for Applications in Olefin Polymerization and Dinitrogen Activation. A Thesis Submitted to the Faculty of Graduate Studies in Partial Fulfilment of the Requirements for the Degree of Master of Science Graduate Program in Chemistry, York University Toronto, Ontario: August; 2012. © Barbara C Skrela; 2012.
176. Robinson RW, Harary F, Balaban AT. The Numbers of Chiral and Achiral Alkanes and Monosubstituted Alkanes. Tetrahedr. 1976;32:355-61.
177. Balaban AT. Chemical Graphs. XXXII. Constitutional and Steric Isomers of Substituted Cycloalkanes. Croat Chem Acta. 1978;51:35-42.
178. Shimshoni JA, Bialer M, Wlodarczyk B, Finnell RH, Yagen B. Potent Anticonvulsant Urea Derivatives of Constitutional Isomers of Valproic Acid. J. Med. Chem. 2007;50:6419-27.
179. Fischer E. Synthese der Mannose und Lävulose. Ber deut chem Ges. 1890;23:370-94.
180. Fischer E. Ueber die Configuration des Traubenzuckers und seiner Isomeren. Ber deut chem Ges. 1891;24:1836-45.
181. Britton G, Liaaen-Jensen S, Pfander H. Carotenoids. Basel: Springer AG; 2004.
182. Bräse S, Encinas A, Keck J, Nising CF. Chemistry and Biology of Mycotoxins and Related Fungal Metabolites. Chem Rev. 2009; 109:3903-3990.
183. Borthwick AD. 2,5-Diketopiperazines: Synthesis, Reactions, Medicinal Chemistry, and Bioactive Natural Products. Chem. Rev., 2012;112:3641-716.
184. Poplata S, Tröster A, Zou Yq, Bach T. Recent Advances In The Synthesis Of Cyclobutanes By Olefin [2+2] Photocycloaddition Reactions. Chem. Rev. 2016;116:9748-815.