

# Original Research Article

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Evaluation of total phenol content of some of the common paddy cultivars of odisha with the application of biofertilizer in assessing resistance to *Meloidogyne graminicola*.

## Abstract

**Comment [Ap1]:** Abstract has not described the contents of the research in detail so that readers do not get brief information about the results and benefits of the research

The quantitative and qualitative changes in plant-derived phenolic substances plays a vital role in induction of plant defense mechanism against various pathogens including rice root-knot nematode *Meloidogyne graminicola*. The total phenol content of four paddy cultivars were studied under pot culture condition in the net house following CRD design under different treatments using biofertilizer *A. brasilense*. The objective of the work was to study the quantitative changes in phenolic compounds content and the effect of biofertilizer in the induction of phenolic compounds on the test nematode. It was observed that there was a significant increase in total phenol content in both resistant and moderately resistant paddy cultivars Abhisek and Manik were seen higher than the susceptible variety Bas-12 and highly susceptible variety Lalat under five different treatments. There were increase in percentage of change ( $p \leq 0.05$ ) in  $T_3$  in all the four varieties followed by  $T_2$  and  $T_1$  with respect to control  $T_5$ . The  $T_4$  treatment in all the R, MR, S and HS varieties were recorded higher as compare to  $T_5$  but less than the treatments  $T_3$ ,  $T_2$ , and  $T_1$  in both roots and shoots against the test nematode, which could exacerbating for eco-friendly RKN management in India.

Key Words – Biofertilizer (*Azospirillum brasilense*), Paddy (*Oryza sativa*) cultivars, *Meloidogyne graminicola*, phenolic substances.

### 1. Introduction :

**Comment [Ap2]:** the references is too old, it would be better to use more up to date ones or no more than the last 10 years

In Asia, paddy is also known as rice belongs to Poaceae family, one of the main food grains of India as well as south-east Asian countries. It provides instant energy with a high carbohydrate content & is a staple food that is consumed by majority of Indian population. Paddy cultivation is an important part of Indian economy. India is the second largest producer of rice & largest

**Comment [Ap3]:** does paddy in other continents have different names?

33 exporter in the world. Rice production increased from 53.6Mt to 120Mt (2020-  
34 21). Odisha is a leading producer of rice of India & has contribution of a sizable  
35 amount of rice grains to the central pool of food stocks.

36 Being an adaptable crop, it can be cultivated in a variety of climates, be it  
37 plains, the mountains & hence is a kharif and rabi crop. In odisha, there are  
38 three systems of paddy cultivation such as dry, semi-dry & wet. The dry system  
39 accounts for 18% of the rice area and rest is shared by semi-dry and wet  
40 systems (Pani & Patra, 2004). Among ecto and endo plant parasitic nematodes,  
41 root-knot nematode *Meloidogyne spp.*, is an important polyphagous group of  
42 plant parasitic nematode which attack paddy crop grown extensively under  
43 various agro-climatic zones of India as well as in most of the south and east  
44 Asian countries of the world. Paddy is adversely affected by the rice root-knot  
45 nematode *Meloidogyne graminicola* and major constraints in agricultural  
46 production that results yield loss ranges from 20-80% & 11-73% respectively  
47 under upland or intermittently flooded condition (Plowright & Bridge, 1990;  
48 Soriano et. al. 2000). Damage in paddy plant consists of various degree of  
49 stunting, lack of vigour, chlorosis of leaf, wilting under moisture stress,  
50 secondary infection by other pathogens produces disease complexes,  
51 malfunctioning of root system. The eggs are laid in the root cortex. The newly  
52 parasitic J2s migrate intercellularly in the rice root cortex towards the root tip,  
53 where nematode invade the vascular cylinder (De Waele and Elsen, 2007) and a  
54 relatively fast life cycle on rice in red lateritic soil favouring soil temperature  
55 22-29 degree celcius (Bridge and Page, 1982; Yik and Birchfield, 1979). The  
56 sedentary nematode feeds from the giant cells, development of nematode  
57 especially pyriform shape of female nematode can result in tearing of the root  
58 and subsequent release of egg masses through this aperture into the soil and  
59 leads to a patchy appearances in paddy field. *M. graminicola* remain within  
60 their host for most of their life cycle and are thus protected from predators and  
61 potential pathogens by the immune system of the host.

62 Among the number of management strategies of rice root-knot  
63 nematodes, though the easy, traditional and quick knock down effects of  
64 nematode control is mainly based on use of chemical nematicides, yet the  
65 harmful impacts on environment due to residual effects of toxic compounds,  
66 creates health hazard in human, increases risk of pollutions, imbalance between  
67 various ecosystems and pest resurgence after prolonged use. So a classical  
68 biological approach, which may emphasize on the non-conventional method of

69 nematode management by using cost effective sources like Biofertilizer such as  
70 *Azospirillum brasilense* is an eco-friendly alternatives to stabilize production for  
71 low cost production. The notable biofertilizers of different strains are known to  
72 have a significant role in suppression of rice root-knot nematode disease and  
73 their effectiveness was reported in paddy by inducing the increasing in total  
74 phenol content of all types of paddy cultivars under different treatments  
75 conditions. Therefore the present experiment work was concentrated to exploit  
76 effectiveness of biomanagement in suppression of *Meloidogyne graminicola*  
77 infecting different paddy varieties by using biofertilizer.

## 78 **2. Materials and Methods :**

79 The experiment was conducted in the net house of the Department of  
80 Nematology, College of Agriculture, Orissa University of Agriculture and  
81 Technology (OUAT), Bhubaneswar during 2020 and 2021 in kharif seasons.  
82 The entire experiment was done under natural environmental condition and was  
83 designed in complete randomized design(CRD) comprising of five treatments  
84 with three replications. After screening and evaluation of different growth  
85 parameters of one hundred ten paddy cultivars by following RKI index (0-  
86 5scale), which were collected from different sources of odisha, four varieties  
87 were selected for biochemical analysis of total phenol content against the  
88 pathogen. In order to understand the basis of nematode resistance four varieties  
89 of paddy cultivars namely Abhisek (Resistant), Manik (Moderately resistant),  
90 Bas-12 (Susceptible) & Lalat (Highly susceptible).

### 91 2.1 : Preparation of soil and sowing of seeds -

92 Five to six healthy seeds of each four varieties were grown in earthen pots  
93 of 15 cm height x 15 cm dia, sterilized with formaldehyde solution (1.0 %) and  
94 filled with autoclaved soil (15 lbs/20min). Before sowing the seeds were surface  
95 sterilized by treating them with 0.1 % HgCl<sub>2</sub> for 5 minutes, washed thoroughly  
96 with sterile water and air dried.

97 The soil to be filled in the pots were pulverised, mixed with N, P, K  
98 fertilizers @ 150:50:60 per hectare on soil weight basis and filled into the pots @  
99 1 kg/pot. The surface sterilized seeds were sown @ 4 to 5 seeds per pot. Each  
100 variety was replicated 3 times. Watering was done regularly after the emergence  
101 of seedlings. At 15 days after sowing the plants –were thinned keeping one  
102 seedling per pot. Biofertilizer *A. brasilense* and nematodes were applied as per  
103 treatment details mentioned below. A small glass tube (2 cm long, 0.5 cm bore)

Comment [Ap4]: Line 92; 5-6 seeds

104 was inserted into the soil near the base of each of the surviving seedlings. Two  
105 weeks after seedling emergence nematodes were counted under a stereoscopic  
106 microscope and released into the holes after removal of the glass tube @ 1000 J<sub>2</sub>  
107 ± 20 per seedling in 10 ml. Sterile water in treatments T<sub>1</sub>, T<sub>2</sub> at 7days prior or  
108 after the application of biofertilizer and at 15DAT in T<sub>4</sub>. For analysis of total  
109 phenol content the following treatments were followed. These pots were  
110 arranged on green house benches according to the treatments and replications.  
111 During the period of investigation, following Complete Randomised Design  
112 (CRD) with five treatments, each replicated thrice. The treatments are as  
113 follows: T<sub>1</sub> - Nematode @ 1000J<sub>2</sub> at 15DAT + *Azospirillum brasilense* @  
114 12kg/ha (after 7days of nematode inoculation) T<sub>2</sub> – *Azospirillum brasilense* @  
115 12kg/ha at 15DAT + Nematode @1000J<sub>2</sub> (after 7days of *A. brasilense*  
116 application), T<sub>3</sub> – *Azospirillum brasilense* @ 12kg/ha only at 15DAT, T<sub>4</sub> –  
117 Inoculated (1000J<sub>2</sub>/plant) and T<sub>5</sub> – Control. Recording of observation after 45  
118 days of transplanting and fresh root and shoot samples were collected for  
119 estimation of total phenol content.

**Comment [Ap5]:** preferably the dose of bacteria can be mentioned only once

**Comment [Ap6]:** When?

## 120 **2.2** : Estimation of total phenolic substances (Bray and Thrope, 1954) -

121 Exactly 0.5 g plant samples were ground with a pestle and mortar in 10  
122 ml of 80 per cent ethanol until it became a pulp. The homogenate was  
123 centrifuged at 5000 rpm for 20 minutes . The process was repeated with another  
124 5 ml of 80 per cent ethanol. Both the supernatants were pooled and evaporated  
125 to dryness. The residue was dissolved in 10 ml distilled water. The aliquot was  
126 pipetted into test tubes with 0.5 ml each. The volume was made up to 3 ml with  
127 distilled water. Exactly 0.5 ml of folin-ciocalteu reagent was added into it.  
128 After 3 minutes 2ml of 20 per cent Na<sub>2</sub> CO<sub>3</sub> solution was added into each tube.  
129 The contents were mixed thoroughly, placed in boiling water for 1 minute and  
130 then cooled. Absorbance was measured at 650 nm in a colorimeter and  
131 compared with a blank. The above procedure was followed for extraction of  
132 phenol from both shoot and root samples of paddy varieties. A standard curve  
133 was prepared using different concentrations of catechol. Then the  
134 concentrations of the phenol in test samples was calculated by comparing with  
135 the standard curve and expressed as mg/g material (catechol) on fresh weight  
136 basis.

## 137 **3. Result**

138 It was observed that there were increase in total phenol content of both  
 139 the roots and shoots due to infestation caused by *M. graminicola* in all the four  
 140 varieties of paddy. The post-infection phenol content increase percentage were  
 141 recorded highest in roots of MR varieties Manik and Abhisek 27.77% and  
 142 30.76% wrt control T<sub>5</sub>. In roots, the increased phenol content in T<sub>3</sub> were  
 143 recorded 0.26, 0.19, 0.22, and 0.20mg/100g samples in Abhisek, Manik, Bas-12  
 144 and Lalat. In treatments T<sub>1</sub> (N→A) and T<sub>2</sub> (A→N) in all R, MR, S & HS  
 145 varieties were observed increased resistance against the nematode as compared  
 146 to the control. The phenolic compound content were recorded higher in  
 147 resistance varieties than susceptible and highly susceptible varieties. Basically,  
 148 shoots of control plants in all four cultivars were observed more content of  
 149 phenolic compounds than their roots. But after *M. graminicola* invasion the  
 150 amount increased in all the varieties in both the roots and shoots.

#### 151 4. Discussion :

152 Patel et. al, 2018 studied that the resistant cultivars of tomato infected  
 153 with *M. incognita* showed maximum increase in the amount of total phenol  
 154 content than the susceptible cultivars. There is a distinct correlation between the  
 155 degree of plant resistance to nematode pathogens and the amount of phenolic  
 156 compounds present in plant tissues, which is the best known factors involved in  
 157 the susceptible-resistance response (Giebel, 1974). Several studies have been  
 158 found that resistant cultivars with nematode infection had higher amount of  
 159 phenolic compounds (Ganguly and Dasgupta, 1982) and the infection with  
 160 *Meloidogyne javanica* stunted the growth of *Carica papaya* and enhanced the  
 161 level of O-dihydric phenols, total phenols and IAA in infected plants (Goel  
 162 et.al, 1982). The resistance inducing plant defense mechanism were being  
 163 activated due to the secretion of phenolic substances against nematode attack  
 164 (Ohri and Pannu,2010).

165 **Table 1- : Percentage of change in total starch content in four different paddy**  
 166 **cultivars under different treatments against *Meloidogyne graminicola* :**

167 **Table-a :** (Average of three replications)

Varieties	Total phenol content in mg/g on fresh weight basis							
	Abhisek (R)				Manik (MR)			
Treatments	Root	% of change	Shoot	% of change	Root	% of change	Shoot	% of change
T <sub>1</sub>	0.24	33.33	0.66	34.69	0.18	38.46	0.67	36.73
T <sub>2</sub>	0.25	38.88	0.68	38.77	0.18	38.35	0.67	36.58

T <sub>3</sub>	0.26	44.44	0.70	42.85	0.19	46.15	0.68	44.68
T <sub>4</sub>	0.23	27.77	0.60	22.44	0.17	27.65	0.60	27.65
T <sub>5</sub>	0.18		0.49		0.13		0.47	
SE (±m)	0.0060		0.0276		0.0054		0.0259	
CD (0.05)	0.0219		0.0531		0.0148		0.0507	

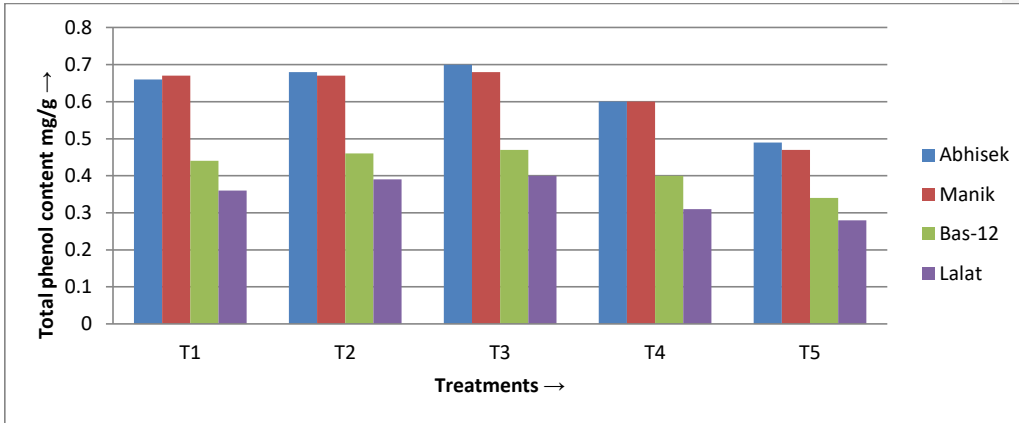
168 **Table-b :**

Varieties	Total phenol content in mg/g on fresh weight basis							
	Bas-12 (S)				Lalat (HS)			
Treatments	Root	% of change	Shoot	% of change	Root	% of change	Shoot	% of change
T <sub>1</sub>	0.20	25.00	0.44	29.41	0.18	20.00	0.36	28.57
T <sub>2</sub>	0.21	31.25	0.46	35.02	0.19	26.66	0.39	39.28
T <sub>3</sub>	0.22	37.50	0.47	38.23	0.20	33.33	0.40	42.85
T <sub>4</sub>	0.19	18.75	0.40	17.64	0.17	13.33	0.31	10.71
T <sub>5</sub>	0.16		0.34		0.15		0.28	
SE (±m)	0.0057		0.0138		0.0055		0.0073	
CD (0.05)	0.0162		0.0398		0.0156		0.0193	

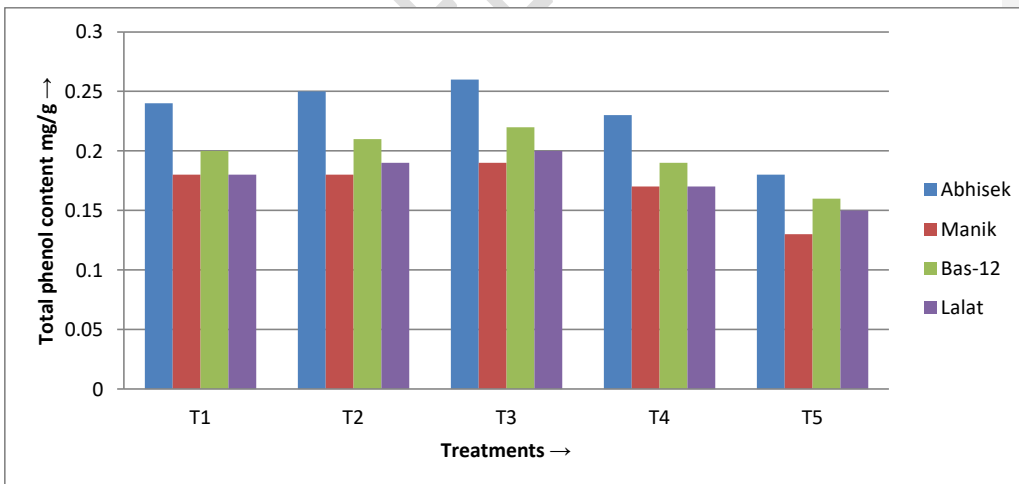
169 T<sub>1</sub> -N @1000J2→Azsp @12kg/ha, T<sub>2</sub> -Azsp @ 12kg/ha→ N @1000J2, T<sub>3</sub> - Azsp @  
170 12kg/ha, T<sub>4</sub> -Inoculated (N @1000J2), T<sub>5</sub> - Control

**Fig.1 - Total phenol content in shoots (mg/g) of different paddy cultivars under five different treatments on fresh weight basis**

**Comment [Ap7]:** It is better add standard deviation



**Fig.2 - Total phenol content in roots (mg/g) of different paddy cultivars under five different treatments on fresh weight basis**



The mode of action of phenolic compounds may be related to the modification in the nematode physiology. RKNs secrete a pool of substances into the plant cell membrane that to induce nematode feeding site formation. (Williamson and Gleason 2003, Cailaud *et.al.*2008). It has been studied that such secretion may be induced by some phenolic compounds such as resorcinol, catechol, hydroquinone, p-coumaric acid and caffeic acid (McClure and Mende, 1998; Jaubert *et.al.* 2002; Bellafiore *et.al.*2008). Pathogen invasion enhances the transcription of m-RNA resulting increase amounts of PAL enzymes that helps in synthesis of more number of phenolic compounds (Taiz and Zeiger, 2002). Barkovskii A., 1995 noticed that *Azospirillum* strains were able o sustain some of its activities using phenolics as an alternative electrons acceptor under low oxygen content caused by the pathogen. Inoculation of *Azospirillum brasilense* showed positive outcomes on the growth of tomato plant against pathogen (Licea and Quiroz, 2020).It helps in reduction of nitrite in mineral decomposition processes and the plant growth promoting bacteria PGPB induces phenolic compounds such as IAA results plant growth. *A. brasilense* has been found in the intercellular spaces of vascular tissues of stems and roots (Robson & Robson, 2015) and also enhances plant hormone synthesis (Baars *et.al.*, 2018).

## 5. Conclusion –

However, it is yet to study that those types of ISR in plant defense mechanism whether the phenolic compounds could lead the nematode to secrete their substances elsewhere rather than roots by preventing heir establishment of feeding sites in such types of plant roots. Biofertilizer enhances the nematode control potential of the plant varieties by inducing the phenol content that unlikely to increase the nematode mortaliy. The nematicidal effects of *Azospirillum brasilense* were seen significantly effective in R & MR varieties such as Abhisek and Manik as well as in another two varieties like Bas-12(S) and HS variety Lalat under different treatment conditions. Biological management approach is a promising and active area of nematological research at the present time. It is an essential step towards improving the level of reliability and biocontrol activity of bio-inoculants that enhances the natural regulations of plant parasitic nematodes below threshold damage level. In *M. graminicola* prone field it could be a cost effective eco-friendly nematode management approach in sustainable agriculture era.

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