

Original Research Article

Exploration and assessment on the agronomic requirement of *Tavernieraabyssinica*A. Rich: a critically endangered medicinal plant of Ethiopia

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ABSTRACT

We determined the distribution and abundance of *Tavernieraabyssinica*A. Rich, a critically endangered endemic medicinal plant of Ethiopia, in the Shewa floristic region, Ethiopia. We also carried out a mesh-house experiment to know whether *T. abyssinica* is able to survive and grow in any soil. From the nine potential locations we made exploration, *T. abyssinica* populations were found only in the two, Lemen and Mojo. The abundance of mature individual was estimated to be more than 600/hectare. The one-way ANOVA and mean comparison (Tukey or Dunn-Bonferroni) results indicated that seedlings grown on soil collected from Lemen and Mojo were significantly ($p<0.05$) taller and heavier than those seedlings grown on Addis Ababa soil (where the species was never reported to grow). We report for the first time that *T. abyssinica* is N-fixing and arbuscular mycorrhizal, at least at the seedling stage. Seedling grown on the Addis Ababa soil were not colonized by arbuscular mycorrhizal fungi while those grown on Lemen and Mojo soils were. Although the estimated abundance of mature individuals was high, there is continued exploitation of the species and habitat loss is imminent. Therefore integrated conservation program by way of ex situ conservation, in situ conservation, and cultivation should be implemented. *Tavernieraabyssinica* could be cultivated in areas with leptosol and degraded vertisol soils with slightly acidic to basic pH. Arbuscular mycorrhizal fungi could have a significant role in the conservation and cultivation of the species.

Keywords: Analgesic property, antipyretic property, medicinal plants (MPs), leptosol, *Tavernieraabyssinica*, vertisol

1. INTRODUCTION

From the recorded 6000 vascular plant species of Ethiopia, the 887 are medicinal plants (MPs) of which, the 2.7% are endemic to Ethiopia [1]. *Tavernieraabyssinica* A. Rich is one of the most valuable endemic MP of Ethiopia [2,3]. Owing to its traditional use to treating sudden illnesses of all sorts, *T. abyssinica* is locally named "Dingetegna" to mean "sudden remedy". The dried

37 slender root (or sometimes stem) is chewed and the juice swallowed or the smoke of these plant
38 parts is inhaled to treat, among others, fever, stomach ache, Colic, and sudden illness due to evil
39 spirits [3]. Analgesic and antipyretic properties of the *T. abyssinica* root extract has been proved
40 to be significant [4]. Its potential effect against stomachache has been demonstrated [5]. It also
41 possesses strong nematicidal and weaker cytotoxic and antimicrobial effects [6]. The root extract
42 was found to contain isoflavonoids [7] which may have potential anticancer effects [8].

43

44 Unsustainable harvest and habitat loss due to agricultural expansion have resulted in the
45 significant decline of the *T. abyssinica* natural populations. Hence, *T. abyssinica* is considered to
46 be one of the critically endangered endemic plants of Ethiopia [9]. In light of its endemism,
47 conservation status, and current and future economic potential, *T. abyssinica* conservation should
48 therefore be among the primary biodiversity conservation priorities in Ethiopia. However, there
49 has been little effort of conserving the species. The current distribution and abundance of *T.*
50 *abyssinica* populations is also not well known.

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52 One of the viable mechanisms of conserving endangered or over-exploited MPs is to cultivate
53 them [10, 11]. Likewise, *T. abyssinica* is one of MPs recommended for cultivation [2]. In the past,
54 there have few research works relevant to the conservation and cultivation of *T. abyssinica*. The
55 seeds of *T. abyssinica* were determined to be orthodox and mechanical or chemical (98% sulfuric
56 acid) treatments were known to be effective methods to break seed dormancy [12]. Effective in-
57 vitro propagation protocols of the species have also been developed [13, 14]. However, the
58 agronomic requirement of the species is not known. Therefore, the objectives of this study

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59 wereto: 1) explore for the natural populations of *T. abyssinica* in the Shewa floristic region and 2)
60 to carry out preliminary assessment on the agronomic requirement of the species.

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62 2. MATERIAL AND METHOD

63 2.1. Description of *Taverniera abyssinica*

64 *Taverniera abyssinica* A. Rich (Fabaceae/Papilionoideae) is a single foliolate or very rarely,
65 pinnately 3-foliolate shrub or shrublet reaching up to 2 m in height. Leaflets are glabrous above
66 while appressed-pubescent beneath. Racemes 2-8-flowered; rhachis and peduncle together c 3-25
67 mm long. Calyx appressed-pubescent outside; lobes equaling or longer than tube. Corolla 12-17
68 mm long, dark pink to purplish red. Pods are stipitate (stalk bearing), 1-3 segmented, finely
69 pubescent and spiny (c 1.5 mm long). Young stems are covered with densely appressed-fine
70 hairs [15, Fig. 1].



71
72 Figure 1: *Taverniera abyssinica* A. Rich. (Photo by Fisseha Asmelash). The flower of *T.*
73 *abyssinica* is Papilionaceous with diadelphous stamen.
74

75 **2.2. Exploration of *Taverniera abyssinica* natural populations**

76 Based on the Flora of Ethiopia and Eritrea [15], herbarium records, and interview with
77 *Dingetegna* vendors in Merkato (the biggest market in Addis Ababa and Ethiopia), we
78 determined that *T. abyssinica* populations were found in Shewa and Tigray floristic regions of
79 Ethiopia. Hence, to assess the current conservation status of *T. abyssinica*, we made
80 explorations in the Shewa floristic region; particularly in the localities of Adadi Mariam, Aliyu
81 Amba, Butajira, Debralibanos, Deneba, Ensaro, Gohatsion, Lemen, and Mojo.

82 At each locality, relevant government offices and experts were contacted and together,
83 local key informants (farmers and traditional healers) were identified for interview. The species
84 was described to the key informants by also telling them the local name and showing an
85 illustration of the species based on [15]. When key informants say they recognize the species,
86 they were asked to locate *T. abyssinica* populations. For those places where the key informants
87 were able to locate *T. abyssinica* populations, the name of the locality, geographic coordinate,
88 altitude, and *T. abyssinica* abundance were recorded. Abundance was determined by counting *T.*
89 *abyssinica* individuals in 30 by 30 plots laid at each site where a *T. abyssinica* population was
90 located.

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92 **2.3. Assessment on the agronomic requirement of *Taverniera abyssinica***

93 **2.3.1. Mesh-house experiment design**

94 To know if *T. abyssinica* can survive and grow on soils other than where its natural populations
95 are found, a mesh-house experiment was carried out in the Ethiopian Biodiversity Institute,
96 Shewa floristic region, Addis Ababa. Based on the *T. abyssinica* exploration result (Table 2),
97 Lemen and Mojo were identified to be the locations where *T. abyssinica* natural populations are

98 found. Hence, soil from Lemen and Mojo was collected for the mesh-house experiment. Another
99 soil from Addis Ababa where no *T. abyssinica* population has ever been reported was also
100 collected. After four months of seedlings growth, survival rate, growth, and root traits
101 (nodulation and mycorrhization) of *T. abyssinica* were compared. Hence, a preliminary
102 assessment on the agronomic requirement of the species was made.

103
104 Seeds collected from the Mojo population were germinated on filter paper according to [12].
105 Germinated seedlings were then transplanted on eighteen 1-liter plastic pots filled with the Addis
106 Ababa, Lemen, and Mojo soil (six pots each). On each of the pot, two *T. abyssinica* seedlings
107 were transplanted making the total number of seedlings in the experiment 36. Hence, in this
108 experiment, the treatment was soil (Table 1) and the three soil types were arranged in a
109 completely random design. The experiment lasted for four months from March 31, 2022 to July
110 31, 2022.

111
112 Table 1: Soil type and physiochemical property of the potting soil used (Based on World Soil
113 Information Service data base available online: <https://soilgrids.org/>)

Soil characteristics	Potting soil type		
	Addis Ababa	Lemen	Mojo
Soil type	Luvisol	Vertisol	Vertisol
Bulk density (cg/cm ³)	128	128	132
Sand content (g/kg)	121	208	219
Silt content (g/kg)	489	307	401
Clay content (g/kg)	376	490	385
pH	6.2	6.7	7.1
Organic carbon (dg/kg)	299	188	174
Total nitrogen (cg/kg)	209	164	162
CEC (mmol(c)/kg)	320	397	513

Note: soil property is for the 5-15 cm of soil depth

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116 **2.3.2. Growth and root traits measurement**

117 After four months of growth, seedlings survival rate and growth, viz., leaf number, shoot height,
118 shoot fresh weight, and root fresh weight were determined. Survival was computed per pot as
119 (number of living individuals/ total number transplanted)*100. Leaf number was determined by
120 counting leaves per plant. Shoot height was measured by a ruler while shoot fresh weight and
121 root fresh weight were measured by analytical balance. Since, part of the root was delicate, some
122 fine root have remained attached to the soil. Hence, the root fresh weight measured was mainly
123 on the root part that was effectively pulled out from the soil matrix and which comprises the
124 major proportion of the root. Two of the most important root traits (nodulation and mycorrhizal
125 association) were also determined by counting the number of root nodules and by measuring root
126 arbuscular mycorrhizal fungi (AMF) colonization. Since nodule size is an important predictor of
127 N-fixation potential [16], we counted the nodules that were easily visible with the necked eyes.
128 Very small nodules that were not developed well were not counted. Root AMF colonization was
129 determined by the gridline intersection method on 100 intersection points [17]by using a
130 NOVEX light stereomicroscope (45x). Roots were first cleared in 10% KOH[18] and stained and
131 de-stained by the ink and vinegar technique [19] using black Hero ink as stain [20].

132

133 **2.3. Data analysis**

134 One-way ANOVA was computed to know the soil preference of *Tavernieraabyssinica*. The data
135 were first checked for the equality of variances and parametric ANOVA or Kruskal–Wallis test
136 were computed to know the effect of soil. When significant effect ($p<0.05$) was found, means
137 were computed using Tukey honestly significant difference (HSD) orDunn–Bonferroni tests
138 ($p<0.05$) respectively for the parametric ANOVA orthe Kruskal–Wallis test. All the statistical
139 analysis was carried out using the R software version 4.1.1.

140 **3. RESULTS**

141 **3.1. The conservation status of *Taverieraabyssinica***

142 From the nine locations where exploration was carried out, *Tavernieraabyssinica* populations
143 were found only in Lemen and Mojo (Table 2). In Mojo, three populations were located and *T.*
144 *abyssinica* grows abundantly (more than 500/hectare). In Lemen also, three populations were
145 located. However, the *T. abyssinica* abundance was much lower than Mojo (more than
146 70/hectare). The *T. abyssinica* populations were found in highly degraded sites with heavy clay
147 soil. In Lemen, the species is mostly found growing solitary. However, in Mojo area, it was
148 found growing in thickets with different plant species (Fig. 2).



149
150 Figure 2: *Taverneria abyssinica* grows in highly degraded sites; *Acacia saligna* planted to reclaim
151 the site is visible (A) and it can grow together with other plant species forming a thicket. Those
152 plants with flowers (B) are *T. abyssinica* individuals.

153 Table 2: The location and abundance of *Tavernieraabyssinica* populations in parts of the Shewa
 154 floristic region

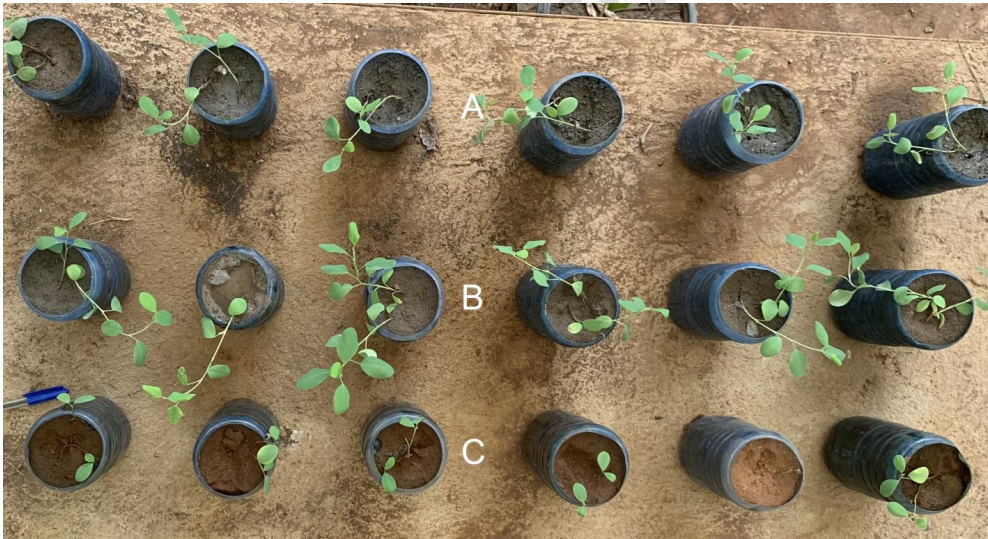
Location	Region	Zone	<i>Tavernieraabyssinica</i> presence and estimated abundance	
			Presence	Abundance
Adadi Mariam	Oromia	South West Shewa	No	Nil [§]
Aliyu Amba	Amhara	North Shewa	No	Nil
Butajira	South	Butajira	No	Nil
Debralibanos	Oromia	North Shewa	No	Nil
Deneba	Amhara	North Shewa	No	Nil
Ensaro	Amhara	North Shewa	No	Nil
Gohatsion	Oromia	North Shewa	No	Nil
Lemen	Oromia	South West Shewa	Yes	78/hectare
Mojo	Oromia	East Shewa	Yes	556/hectare

155 [§]local informants described the species correctly and insisted the species is present; however they
 156 were not able to locate a single site where *Tavernieraabyssinica* is growing.
 157

158 3.2. The agronomic requirement of *Tavernieraabyssinica*

159 Generally, seedlings grew better on the Mojo and Lemen soil and in a descending order while
 160 they grew poorly on the Addis Ababa soil (Fig. 3). After four months of growth, the survival rate
 161 of *T. abyssinica* seedlings was 100%, 92%, and 83%, respectively for Mojo, Lemen, and Addis
 162 Ababa soils. According to the one-way ANOVA result, these survival rates were not
 163 significantly ($p>0.05$) different (Table 3). However, significant ($p<0.05$) effect of soil was found
 164 for all the remaining seedling variables measured, viz., leaf number, shoot height, shoot fresh
 165 weight, root fresh weight, root nodule number, and root AMF colonization (Table 3). The
 166 seedlings grown on the Mojo soil produced the highest leaf number which was significantly
 167 ($p<0.05$) and 23.48% and 305.71% more than the mean leaf number of the seedlings grown on
 168 the Lemen and Addis Ababa soils respectively. Similarly, the highest shoot height, shoot fresh
 169 weight, and root fresh weight was also recorded for seedlings grown on the Mojo soil to be
 170 followed by the seedlings grown on the Lemen and Addis Ababa soils respectively. Whereas the
 171 highest nodule number was recorded for seedlings grown on the Lemen soil, it was for the
 172 seedlings grown on Mojo soil that the highest root AMF colonization (RC) was recorded. The

173 nodule number of seedlings on the Lemen soil was significantly ($p<0.05$) and 236.64% and
 174 1233.33% greater than the mean nodule number of seedlings grown on Mojo and Addis Ababa
 175 soils respectively. The RCoF of seedlings on the Mojo soil was not significantly ($p>0.05$) greater
 176 than the mean RC of the seedlings grown on the Lemen soil. The RC of seedlings grown on both
 177 Mojo and Lemen soil were however, significantly ($p<0.05$) greater than the mean RC of
 178 seedlings grown on the Addis Ababa soil. The roots of the seedlings grown on the Addis Ababa
 179 soil were not colonized by AMF (Table 3). Root nodules recorded were mostly oval but there
 180 were also elongate/cylindrical and spherical/globose nodules. In the case of RC Vesicles were
 181 the frequently observed structures (Fig. 4).



182 Figure 3: *Taverniera abyssinica* seedlings after four months of growth in a mesh-house in the
 183 Ethiopian Biodiversity Institute. Row A=Lemen soil, Row B=Mojo soil, and Row C=Addis
 184 Ababa soil. Seedlings seem to grow poorly on the Addis Ababa soil.
 185
 186

187 Table 3: One-way ANOVA results and mean comparison

Variables	Mean (\pm SE) values on the different soil			ANOVA	
	Addis Ababa	Lemen	Mojo	F	Chi.sq.
Survival rate (%)	83.0(\pm 17) ^{ns}	100.0(\pm 0.0) ^{ns}	92.0(\pm 8) ^{ns}	0.6	-
Leaf number	1.75(\pm 0.25) ^c	5.75(\pm 0.39)	7.1(\pm 0.82) ^a	-	24.312***

		^b			
Shoot height (cm)	5.77(±0.31) ^c	9.44(±0.5) ^b	12.25(±0.9) ^a	25.421***	-
Shoot fresh weight (g)	0.15(±0.01) ^c	0.43(±0.04) ^b	0.65(±0.06) ^a	32.617***	-
Root fresh weight (g)	0.11(±0.01) ^b	0.18(±0.02) ^a	0.24(±0.04) ^a	5.5685*	-
Root nodule number	0.6(±0.6) ^b	8.0(±1.1) ^a	2.2(±1.0) ^b	17.796***	-
Root AMF colonization (%)	0.0(±0.0) ^b	26.17(±7.1) ^a	45.0(±13.3) ^a	-	10.753**

188 *significant at $p < 0.05$, ** significant at $p < 0.01$, and *** significant at $p < 0.001$. Means labeled
 189 with different letters across columns are significantly different for TukeyHSD or Dunn-
 190 Bonferroni tests ($p < 0.05$), "ns" indicates means are not significantly different ($p > 0.05$).
 191



192 Figure 4: Nodulated roots of *Taverniera abyssinica* seedlings above and arbuscular mycorrhizal
 193 fungi (AMF) colonization below. E=elongate nodule, O=oval nodule, S=spherical nodule, V=
 194 vesicles (root colonized with AMF), NC=root not colonized with AMF. Photos are not with
 195 scale.
 196

197 **4. DISCUSSION**

198 *Tavernieraabyssinica* was categorized to be an IUCN “critically endangered” species not based
199 on abundance of mature individuals but by its distribution, experts’ judgment of population
200 decline, and actual or potential levels of exploitation [9]. Here, we have determined an estimated
201 abundance of *T. abyssinica* mature individuals (250-1000) which if used alone, will put the
202 species under the “vulnerable” category. Since we found *T. abyssinica* populations in two
203 locations (six sites), one of the criteria, i.e., existing only at a single location, that was used
204 previously to designate the species as critically endangered [9] is also not valid currently. This
205 could be more so if the number of mature individuals were to be accounted throughout Ethiopia
206 (Table 4). However, we observed the species is still under exploitation for its root and in Adadi
207 Mariam, we noticed its habitat has been changed to cultivated land. The Lemen populations were
208 also found interspersed within agricultural lands, which indicate agricultural expansion in to the
209 specie’s habitat have taken place. Moreover, we have observed frequent uprooting of the species
210 particularly in the Lemen area. Hence, although the number of mature individuals could
211 potentially be significantly high, considering the current and potential future exploitation and
212 considering the high risk of habitat loss, the current critically endangered designation could still
213 be appropriate. In Mojo area, we observed some of the *T. abyssinica* populations growing in
214 thickets. This corroborates the flora record that described the species as a bush land species [15].
215 Therefore, *T. abyssinica* could be conserved in situ in protected bushlands/forests.

216
217 Available data indicate that *T. abyssinica* grows on leptosols and vetrisols (Table 4). Our results
218 corroborate this data. Hence, in the mesh-house condition in the Ethiopian Biodiversity Institute
219 (2400 m altitude), *T. abyssinica* although survived well, it did not grow fit on the

220 Luvisol compared to the vertisol. Hence, soil type could be an important agronomic requirement
 221 of *T. abyssinica*. Leptosols and vertisols could have better CEC and copper content compared to
 222 Luvisols [21]. Hence, soil type in general and CEC and essential nutrients such as copper in
 223 particular, could be important factors to determine *T. abyssinica* growth. The soil pH could be
 224 another important factor. Seedlings grew better on the Mojo soil with a slightly basic pH
 225 compared to the Lemen and Addis Ababa soils with slightly acidic and acidic pH values. This
 226 may indicate that the species prefers slightly basic soils than acidic ones. The comparative better
 227 growth of *T. abyssinica* seedlings on the Mojo soil could also be due to home soil advantage.
 228 This is because, the seeds used in this study were collected from the Mojo populations.

229 Table 4: Soil type and pH of sites where herbarium record *Taverniera abyssinica* was found
 230 (Based on <https://soilgrids.org/>).

	Location	Floristic region	Soil type	pH
8	Mekelle city	Tigray	Leptosol	7.9
6	Adigudem-B	Tigray	Vertisol	7.8
1	Adigudem-A	Tigray	Vertisol	7.7
3	Hagereslam	Tigray	Leptosol	7.7
7	Adi Amedy	Tigray	Leptosol	7.7
5	Mekelle/Giba plane	Tigray	Leptosol	7.5
2	Gijet	Tigray	Leptosol	7.3
4	Abala	Tigray	Leptosol	7.2
11	Gebraguracha	Shewa	Leptosol	6.4

231 Note: exploration to all these locations was not possible due to security reasons

232
 233 Leptosols are problematic soils for crops growth [22]. *Taverniera abyssinica* thrives well on
 234 such soils and degraded vertisols. Hence, the species must have effective soil resource acquisition
 235 mechanisms. Therefore, the assessment we made on its root traits, i.e., nodulation and
 236 mycorrhization, was important to better understand its agronomic requirement. Particularly,
 237 understanding its association with arbuscular mycorrhizal fungi (AMF) is very important [23,
 238 24]. Based on our results, we report for the first time that *T. abyssinica* is N-fixing and
 239 arbuscular mycorrhizal at least at the seedling stage. Mature individuals in the field were

240 observed to be without nodules. The fact that the species is a nodulating species is to be
241 expected. This is because, its roots were found to produce isoflavonoides [7], a biochemical
242 required for nodulation and typical of the Papilionoideae subfamily [25]. The nodule number we
243 record for seedlings grown on Mojo was not significantly ($p < 0.05$) greater than the nodule
244 number recorded for seedlings grown on soil Addis soil. However, seedlings grown on the Addis
245 Ababa soil were not colonized by AMF. Hence, the significantly ($p < 0.05$) higher growth
246 recorded for *T. abyssinica* seedlings on the Mojo soil and the opposite on the Addis Ababa soil
247 could indicate that *T. abyssinica* forms association with a selected AMF communities that are
248 lacking in the Addis Ababa soil [26]. Rhizobia (nodulation) could have no or less significant role
249 in this regard.

250

251 5. CONCLUSION

252 One of our objectives was to determine the distribution and abundance of *T. abyssinica*, a
253 critically endangered plant species, in Shewa floristic region, Ethiopia. The other objective was
254 to assess the agronomic requirement of the species. We have fulfilled both our objectives. From
255 the nine locations where exploration was carried out, *T. abyssinica* populations were found only
256 in the two, viz., Lemen and Mojo. The abundance of *T. abyssinica* was much greater than what
257 we initially expected. However, the treatment levels that were primarily used to designate its
258 conservation status seem to persist. Hence, to improve the conservation status of the species,
259 integrated conservation program by way of ex situ conservation, in situ conservation, and most
260 importantly, cultivation is crucial. Based on the mesh-house experiment results, and relevant data
261 we gathered, *T. abyssinica* could be cultivated in leptosol and degraded vertisol sites with
262 slightly acidic to basic pH. We report for the first time that *T. abyssinica* is N-fixing and

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263 arbuscular mycorrhizal. Arbuscular mycorrhizal fungi (AMF) could play key role in the future
264 conservation/cultivation program of the species. Hence, the AMF communities associated with
265 *T. abyssinica* should be identified.

266

267 **CONSENT:** It is not applicable.

268 **ETHICAL APPROVAL:** It is not applicable.

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270 REFERENCES

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