

Original Research Article

Bamboo shoot processing - effects on nutritional and anti-nutritional quality

ABSTRACT

The present investigation was designed to find out the nutritional potential of some local edible bamboo shoot species and the impact of different processing methods on it. Sliced bamboo shoot pieces were treated with soaking in water for 30 min, boiling in water for 10 min, boiling in 1% NaCl solution for 10 min, boiling in 5% NaCl solution for 10 min and steaming for 10 min. There were significant variations in ash, crude protein, total carbohydrate, minerals, ascorbic acid, phenol, flavonoid, and tannin contents. Boiling in 1% NaCl solution for 10 min was found best in retaining the nutrients with a significant reduction in cyanide content. Overall, bamboo shoots confer numerous health beneficial activities and have the potential to be used as bioactives.

Keywords: *Anti-nutritional, Bamboo shoot, Cyanide, Nutritional.*

1. INTRODUCTION

Bamboo is a giant and fast-growing woody grass of the subfamily *Bambusoideae* (family *Poaceae*) and consists of approximately 1,250 species belonging to 75 genera. In terms of diversity of bamboo species, India is the third largest country in the world with about 125 species of bamboo belonging to 23 genera, China and Japan being the first and second with 300 and 237 species respectively [1]. Bamboo is one of the most economically important plants in the entire world with more than 1500 different uses - from construction materials to agricultural tools, from utensils and musical instrument to ornamental use etc.

The juvenile shoot of bamboo, commonly known as bamboo shoot is being consumed traditionally since time immemorial by the tribal communities worldwide, especially communities of South, South-East and East Asia in fresh, dried, shredded, canned, pickled or fermented and in the medicinal

form [2]. Fermented bamboo shoot is a common consumption pattern of NE India such as *kharisa* (in vernacular) in Assam. Bamboo shoots are regarded as an ideal vegetable because of its high nutrient contents in terms of dietary fibre, vitamins, carbohydrates, proteins, minerals, secondary metabolites and antioxidants [3, 4, 5, 6, 7]. Due to its high fibre content and nutritional value, it is commonly used in many food products like breakfast cereals, sauces, fruit juices, shredded cheeses, frozen desserts, pastas, bakery and meat products etc.[8].

Besides gastronomical practices, people have been using bamboo shoots traditionally to cure different diseases. Kalita and Dutta [9] have revealed that some ethnic tribes in Northeastern states of India, used bamboo shoots to treat cardiovascular ailments, control high blood pressure etc. Furthermore, South Asian countries were reported to utilize bamboo traditionally to relieve hypertension, sweating, and paralysis [10]. Overall, bamboo shoots confer numerous health beneficial activities such as improving appetite and digestion, curing cardiovascular disease and show antioxidant activity [11], controlling weight loss, anti-fatigue activity [12], antiallergic effects [13], anti-inflammatory [14] and anti-cancer [15] properties, antiviral activity [10]. Also, a regular consumer can indulge a youthful feeling, athletic energy, and longevity due to phytosterols present in the bamboo shoot [16].

Bamboo shoots are normally harvested when the shoot height is about 15-30 cm and generally 7-14 days after the emergence depending upon the species. The young shoots with overlapping sheaths are tightly clasped, which are removed to get the edible part. The freshly harvested bamboo shoot is soft, crispy, creamy yellow in color with a sweet taste that delivers a strong smell [17]. Shoots may contain as high as 90 per cent water at the time of harvest.

Not every species of bamboo shoot that can be found worldwide is edible. In India, economically important 18 bamboo species have been identified, out of which around 10 are edible [18]. The cyanogenic glycosides (taxiphyllin), a natural plant toxin is found in high amounts in bamboo shoots [19]. Different chemical constituents are responsible for the taste of bamboo shoots. While tannins increase the offensive taste of bamboo shoots, homogentisic acid provides pungent taste [20], and hydrogen cyanide (HCN) develops bitterness [17]. Due to high pungent smell and bitter acidic taste, many people avoid consuming bamboo shoots. There are reports that the cyanogenic glycosides of bamboo shoots

cause acute cyanide poisoning, in animals including humans. But cyanogens can be removed or reduced before consumption by adequate processing, thereby reducing the potential health risk [21]. This work was conducted to provide the nutritional profile of some locally available and widely used edible bamboo shoot species of Assam with the effects of some processing treatments in reducing cyanide content and analyze their effects on nutrient contents.

2. MATERIALS AND METHODS

2.1 Materials and Reagents

Three locally available edible bamboo shoot species of Assam *i.e.*, *Bambusa balcooa* (in vernacular, *bhaluka*), *Bambusa tulda* (in vernacular, *Jati*) and *Dendrocalamus hamiltonii* (in vernacular, *Kako*) were collected from Sivasagar district (Betbari) after attaining the height of 30 cm from the ground by each species separately. The shoots were cleaned by removing any dust and dirt after harvesting and wrapped with cling film (Oxywrap PVC cling film). The bamboo shoots were transported to the laboratory on the same day of the harvesting and kept in the refrigerator at 4 °C for further analysis. The chemicals and reagents used in the current study were of the analytical grade.

2.2 Sample preparation and processing treatments

The bamboo shoots were further cleaned by removing the outer covering sheaths and hard nodal portions while only the soft inner edible portions were taken for analysis. The cleaned shoots were cut into small pieces of equal size and subjected to five different treatments: (a) soaking in water for 30 minutes (T_1), (b) boiling in water for 10 minutes (T_2), (c) boiling in 1% NaCl for 10 minutes (T_3), (d) boiling in 5% NaCl for 10 minutes, (e) steaming for 10 minutes (T_5). Raw bamboo shoot without any treatment was taken as control (T_0).

2.3 Proximate composition

Selected raw and processed bamboo shoots were analyzed for the proximate composition and mineral composition by following standard protocols described in the Association of Official Analytical Chemists [22].

2.4 Ascorbic acid

Ascorbic acid was determined by volumetric method [22]. Here, sample extracts were prepared by homogenizing bamboo shoots with 4% oxalic acid and the supernatants were titrated against the 2,6-dichlorophenolindophenol dye until pink colour appeared. The standard ascorbic acid solution was diluted with 5 ml of 4% oxalic acid and titrated with dye solution till pink color persisted for 10 seconds to get the dye factor. The calculation was done by the following formula: $\text{Ascorbic acid} = (\text{Titre value} \times \text{Dye factor} \times \text{Volume made up} \times 100) / (\text{Aliquot taken} \times \text{Weight of the sample})$

Dye standardization

Five ml of standard ascorbic acid solution was diluted with 5 ml of 4% oxalic acid and titrated with dye solution till pink color persisted for 10 seconds. Dye factor (mg of ascorbic acid per ml of dye) was calculated as follows: $\text{Dye factor (D.F)} = 5.0 / \text{titre value}$

2.5 Total phenol content

Total phenolic content was estimated using Folin-Ciocalteu reagent [23]. In this method, sample (1 g) was extracted with 80% methanol (10 ml) and centrifuged at 10000 rpm (25°C). The residue was re-extracted for 5 times and centrifuged. The supernatant was used for the analysis of total phenol content. The supernatant was dried, and the pellet was dissolved in 5 ml of distilled water. Aliquots of the solution (between 0.2-2 ml) were then taken, and the total volume was increased to 3 ml using distilled water. Each test tube received 0.5 ml of Folin-Ciocalteu reagent and 2 ml of 20 percent sodium carbonate after three minutes. The combination was maintained at 100°C for one minute, and the sample's absorbance after cooling was measured at 650 nm (Chemito UV 2100 spectrophotometer). Catechol was used as the standard. The total phenol was calculated as catechol equivalent, expressed as mg phenol per 100 g of the sample.

2.6 Alkaloids

The total alkaloids content in the sample were estimated using the method of Daniel [24]. Two gram sample was taken and about 10 times water was added to it. The sample was boiled for 30 min and filtered in hot condition. Then 10% lead acetate solution was added and centrifuged to separate the precipitate. The supernatant was boiled and then cooled. An equal volume of chloroform was added and transferred to a separating funnel and shook it. Separating funnel was then left undisturbed for half an

hour. The chloroform and aqueous layer were collected separately. The aqueous layer was again extracted with chloroform and pulled with previous chloroform extract. The chloroform was then evaporated, and the residue was then weighed. Alkaloid (%) = (Weight of residue × 100)/(Weight of the sample taken)

2.7 Flavonoid

Flavonoid content was determined by aluminium chloride colorimetric method [25]. Flavonoids were extracted with 80% ethanol in a ratio of 1:10 (w/v) and kept on a rotary shaker for 24 h. The extracts were filtered through Whatman No. 42 filter paper and the volume was adjusted with 80% ethanol. To 0.5 ml of extract, 1.5ml ethanol (95%), 0.1ml aluminium chloride (10%), 0.1ml potassium acetate (1M) and 2.8 ml of distilled water were added in order and incubated at room temperature for 30 min and then absorbance was measured at 415 nm in a UV-visible spectrophotometer (Varian Cary 50 Scan). Quercetin was taken as standard to make the calibration curve. The result was expressed as mg of quercetin equivalents (QE)/100g of dry weight.

2.8 Tannin

Tannin content was determined by the Folin Denis method [26]. For extraction, 0.5 g powder was boiled with 75 ml of distilled water for 30 min, and then centrifuged at 2000 rpm for 20 min. After centrifugation, the supernatant was collected in a volumetric flask and total volume was made to 100 ml with distilled water. A volumetric flask was filled with 1 ml of sample extract, 5 ml of Folin-Denis reagent, 10 ml of 35 percent sodium carbonate, and 100 ml by water in total. The solution was mixed well and after 30 minutes absorbance was read at 700 nm. The standard graph was prepared by using 0-100 µg tannic acid and expressed as tannic acid equivalent.

2.9 Cyanogens

The cyanogen content was determined by picrate paper method [27]. In a flat-bottomed glass bottle, 25 mg (z) grounded fresh bamboo shoot sample was placed and immediately 0.5 ml of 0.1M phosphate buffer (pH 6) was added to it followed by a yellow picrate paper attached to a plastic strip so that there is a gap between paper and the liquid of the bottle. Right away, the bottle was closed with the screw-capped lid. The blank was prepared by the same method without the addition of bamboo shoot sample. The bottle

was kept in a dark place for 16 to 24 hours at room temperature. Finally, the orange-brown picrate paper was immersed in 5.0 ml of distilled water for 30 min with gentle shaking occasionally. Absorbance was measured at 510 nm. The total cyanogens content (ppm) = $(396 \times \text{absorbance} \times 100)/z$

2.10 Statistical analysis

The experiment was performed by taking three replicates and the data obtained were subjected to statistical analysis using IBM SPSS Statistics version 25.0 for analysis and the results were statistically evaluated at significance level $p \leq 0.05$.

3. RESULTS AND DISCUSSION

The three local species of edible bamboo shoot *B. tulda*, *B. balcooa* and *D. hamiltonii* were processed with different treatments and their effects on nutritional quality along with cyanide reduction were analyzed. A fresh sample of bamboo shoots without any processing treatment was taken as control.

3.1 Processing effect on proximate parameters

The proximate parameters of fresh and processed bamboo shoots are presented in Table 1. The moisture content in the fresh bamboo shoots under study ranged from 90.76 to 92.29%. Such a high moisture content makes bamboo shoots highly perishable. Karanja *et al.* [28] also described the moisture content in the young shoots of a bamboo species ranging between 92.2-92.4% which is in agreement with the present findings. The young shoots of bamboo are rich in protein [29]. The shoots of *B. tulda*, *B. balcooa* and *D. hamiltonii* had 16.42%, 14.44% and 13.56% of crude protein respectively. Slightly higher crude protein content (21.1% to 25.8% on a dry weight basis) was reported by Kumbhare and Bhargava [30]. Kalita *et al.* [31] reported protein percentage in *B. balcooa*, *B. bambos*, *D. hamiltonii* and *B. vulgaris* as 28.52%, 33.54%, 33.73% and 22.30% respectively. Such variation in protein content might be because of the genetic make-up of the species used, pH of soil, nutrition status of the soil, climate, rainfall, and the growth stage at which bamboo shoots were harvested. Crude protein content was found to decrease in all the treatments in the present study with maximum reduction in the treatment where bamboo shoots were boiled in 5% NaCl solution for 10 minutes. The nutrients like protein, reducing sugar, are significantly influenced by temperature and blanching time [32]. Blanching at 95°C markedly reduced the protein content of bamboo shoot in comparison to 75°C and 85°C as most of the labile protein

gets denatured at high temperature [33]. The crude protein content might decrease because of boiling and soaking that might cause denaturation and solubilization of the nitrogenous substances [34].

The bamboo shoots can be considered an ideal vegetable due to low fat content, richness in minerals and dietary fibre [35]. The present study revealed that the fresh shoot of *B. tulda*, *B. balcooa* and *D. hamiltonii* had 1.48, 1.49 and 1.83% crude fat content respectively (Table 1). The crude fat content was almost in-line with those as reported by Bhatt *et al.* [36]. Badwaik *et al.* [32] observed that temperature and duration of blanching treatment significantly affected crude fat content. Blanching for 30 min resulted in the reduction of crude fat from 0.52 g/100g to 0.38, 0.28 and 0.19 g/100g at 75°C, 85°C and 95°C respectively. It was observed that blanching for 5-10 min retained more fat as compared to blanching for 20-30 minutes. In the present investigation, crude fat content was found reducing with each treatment with the highest reduction in all the samples steamed for 10 minutes. The reduction in fat might be due to oxidation and melting of fat at high temperature and longer duration, which may be lost with water during blanching [37].

The crude fibre content in *B. tulda*, *B. balcooa* and *D. hamiltonii* shoots was 5.28%, 5.57% and 4.09% respectively (Table 1). The almost similar crude fibre content of 6.90% was reported by Rajyalakshmi and Geervani [38]. Higher crude fibre content ranging from 23.1% to 35.5% in shoots of different bamboo species was reported by Bhatt *et al.* [36]. Variation in fibre content in bamboo shoots might arise from species difference as well as the growth stage and period of the harvest of bamboo shoots. Chongtham *et al.* [4] reported an approximately three-fold increase in dietary fibre in shoots after keeping for 10 days. Different treatments used in this study had no significant effect on the crude fibre content in the shoots of all the three bamboo species. Badwaik *et al.* [32] also reported that crude fibre content in bamboo shoots was almost unaffected against blanching temperatures and time.

Carbohydrate contents were recorded as 6.58%, 7.00% and 5.37% on dry weight basis in *B. tulda*, *B. balcooa* and *D. hamiltonii* shoots respectively (Table 1). Singhal *et al.* [39] reported the carbohydrate content in the raw bamboo shoots ranged from 2.0% to 9.94%. There was a significant increase in the total carbohydrate contents in all the three bamboo species upon boiling in water for 10 minutes which might be due to the hydrolysis of complex polysaccharides of bamboo shoots. Kumbhare

and Bhargava [30] also observed an increase in carbohydrate contents ranging from 3.1% to 5.1% after boiling. However, a decrease in carbohydrate contents in bamboo shoots was found when boiled in 1% and 5% salt solution separately for 10 minutes. During wet processing in presence of salt, heating might help in leaching out of carbohydrates in water resulting in the decrease of total carbohydrate content [30]. Pandey and Ojha [35] reported similar observation when bamboo shoots were boiled in solution with different salt concentrations.

The ash content of a food is a measurement of the overall quantity of minerals present. Bamboo shoots are enriched with minerals like K, Na, Ca, Mg, P, and Fe. The present study revealed that shoots of *B. tulda* had 1.10%, *B. balcooa* had 1.54% and *D. hamiltonii* had 0.89% of ash content on a dry weight basis (Table 1). The result was found to be in-line with results reported earlier. Rawat *et al.* [40] found 1.03% ash in bamboo shoots. Badwaik *et al.* [32] reported ash content in the range of 0.82-0.90 g/100g of fresh shoots of *D. hamiltonii*, *B. balcooa* and *B. pallida*. However, there were reports of high ash content of 14.2-17.1% in bamboo shoots [6, 28]. Such variation in ash content in bamboo shoots could be due to the age of the shoots, samples used for analysis and genetic variations of the bamboo varieties. Ash content was decreased significantly with all the processing treatments with highest reduction of ash content in *B. tulda* with boiling the shoots in water for 10 min and boiling the shoots in 1% NaCl solution for 10 min. Reduction in ash content was found lowest in soaking the bamboo shoots in water for 30 minutes. The decrease in ash content upon boiling was reported by several workers [30, 4, 41] which might be due to the leaching out of minerals in water during the processing treatment [40].

3.2 Processing effects on mineral contents

In the present study, minerals such as calcium, phosphorus, iron and potassium were found in the range of 4.83 to 6.70 mg, 572.0 to 654.0 mg, 9.34 to 10.94 mg and 645.33 to 890.33 mg/100g on dry weight basis respectively (Table 2). The three bamboo species varied significantly in their contents in these mineral elements. The treatments used did not have significant effect on calcium contents in bamboo shoots under study. However, treatments showed significant effects on phosphorus, iron and potassium contents in bamboo shoots. The contents of these mineral elements were found to decrease significantly with all the treatments in all the three species of bamboo shoots. The highest retention was found in

samples soaked in water for 30 minutes and the lowest was found in boiling in water for 10 minutes. According to wet processing (soaking and boiling) and heat treatment was reported to be the major cause for the leaching out of mineral or ash content leading to the reduction in their contents [35].

3.3 Processing effects on ascorbic acid and secondary metabolite contents

Ascorbic acid contents in *B. tulda*, *B. balcooa* and *D. hamiltonii* shoots ranged between 2.47-3.31 mg/100g fresh sample (Table 3). These values were found lower than those reported earlier [35, 36]. All the treatments in the present study significantly reduced the ascorbic acid content in the shoots of all the bamboo species. Badwaik *et al.* [32] found that blanching temperature significantly reduced the ascorbic acid content of bamboo shoot. Blanching for short time (5-10 min) retained more ascorbic acid while at 20-30 min the losses were high. Zhang *et al.* [37] reported that ascorbic acid retention in boiled bamboo shoots was 47.37% while in steamed ones; it was 57.83%. Such variation might be due to the high solubility and heat labile character of ascorbic acid. Soaking and boiling, therefore, might cause leaching of ascorbic acid into water and destruction by oxidation during boiling [42, 43]. The reduction was lowest in the treatment soaking with water for 30 minutes.

There were significant variations in the total alkaloid, phenol and flavonoid contents of *B. tulda*, *B. balcooa* and *D. hamiltonii* shoots used in the study (Table 3). The alkaloid content varies with growing conditions, environmental factors, storage time and temperature, harvest season, age of shoot as well as part of shoot used [44]. No significant effect of the treatments was observed on the alkaloid content of bamboo shoots under study. The total phenol content varied between 493.64 and 506.62 mg Catechol Equivalent/100g on a dry weight basis. The phenolic content was found increasing in *B. tulda* and *D. hamiltonii* with steaming the bamboo shoots for 10 minutes. In other treatments, phenol content was found decreasing. Zhang *et al.* [37] reported after boiling total phenolic content was significantly reduced while total phenolic content increased by 3.98% after steaming. Badwaik *et al.* [32] also observed loss of phenolic content with an increase in temperature and duration of blanching. Heat treatment might inactivate polyphenol oxidases, thereby preventing decomposition of phenols [45]. Stewart *et al.* [46] also reported that dietary fibre bound polyphenol decomposed into free phenolic compounds due to heat treatment which made the detected phenolics value higher.

Flavonoid contents in bamboo shoots were found to be 370.00, 445.00 and 352.67 mg QE/100g in *B. tulda*, *B. balcooa* and *D. hamiltonii* respectively (Table 3). Kalita *et al.* [31] reported much higher flavonoid content in *B. balcooa*, *B. bambos*, *D. hamiltonii* and *B. vulgaris* as 5.19%, 7.22%, 5.32% and 4.17% respectively. The highest reduction in flavonoid content was found in *B. tulda* and *B. balcooa* shoots, boiled in 5% NaCl solution for 10 minutes. *D. hamiltonii* shoot was affected most by boiling in 1% NaCl solution. Flavonoid is water-soluble and can easily be reduced by heat treatment and this might be the probable cause for such reduction.

3.4 Processing effects on anti-nutritional contents

Tannin contents were found to be 204.20, 184.30 and 244.00 mg tannic acid equivalent/100g on a dry weight basis (Table 3) for *B. tulda*, *B. balcooa* and *D. hamiltonii* shoots respectively. Processing caused a reduction in the tannin contents in bamboo shoots. Tannin content was affected most by boiling bamboo shoots in 5% NaCl for 10 minutes. The minimum loss was recorded in soaking in water for 30 minutes. As tannin is water-soluble and heat-sensitive, during hot and wet processing loss was found maximum.

Bamboo shoots contain taxiphyllin, a cyanogenic glycosides which upon maceration gets hydrolyzed by intracellular enzyme β -glycosidase to hydrogen cyanide (HCN). Cyanide can cause toxicity in humans, thus necessitating its removal from the shoots before human consumption. In the present study, cyanide contents in *B. tulda*, *B. balcooa* and *D. hamiltonii* were found to be 875.16, 866.91 and 1067.87 ppm respectively (Table 3). Mina *et al.* [47] reported distribution of cyanide in the succulent bamboo shoot of *B. tulda* as 354.45, 598.26 and 900.00 ppm in the base, middle and a tip portion. Cyanogen glycoside content is highest in the shoot tips of most of the edible bamboo shoots species [16]. High cyanogenic content in bamboo shoots can be mitigated by boiling in water for about 2 hours and by NaCl treatment. Anti-nutrient content can be reduced significantly by fermenting bamboo shoot slices [48]. In the present study, cyanide contents were reduced with each treatment and the maximum reduction (96-98%) was found with boiling the bamboo shoot at 5% NaCl for 10 minutes. Rana *et al.* [49] also treated bamboo shoots with NaCl and reported 95.4-98.1% reduction in cyanide content. The bamboo shoot's cell walls broke down during boiling, allowing the release of its cell content along with poisonous and antinutritional elements [50]. Jaiwunglok *et al.* [51] reported that sodium chloride might

facilitate the leaching of taxiphyllin from bamboo shoot through exosmosis reaction. Therefore, optimized boiling of bamboo shoots may be an effective way to make them suitable for human consumption.

4. CONCLUSION

This study was conducted to find out the nutritional potential of some local edible bamboo shoot species (*Bambusa tulda*, *B. balcooa* and *Dendrocalamus hamiltonii*) and to find out the effect on the nutritional and anti-nutritional components by different processing methods. Bamboo shoots were found to differ in their nutritional and anti-nutritional contents. The processing methods were found effective in reducing the anti-nutrients, but also affected the nutrient contents. The highest reduction of cyanide was found in boiling in 5% NaCl for 10 minutes. Soaking in water for 30 minutes showed the highest retention of nutrients and cyanide content. Boiling in 1% NaCl for 10 minutes could be considered best with a significant reduction in cyanide content and better retention of the nutrients. Bamboo shoot is a highly nutritious food along with the medicinal value. Adapting pre-treatments and proper cooking methods, the toxic chemicals in bamboo shoots can easily be eliminated.

REFERENCES

1. Bal LM, Singhal P, Satya S, Naik SN, Kar A. Bamboo shoot preservation for enhancing its business potential and local economy: a review. *Critical Reviews in Food Science and Nutrition*. 2012;52(9):804-814.
2. Choudhury D, Sahu JK, Sharma GD. Bamboo shoot based fermented food products: a review. *Journal of Scientific and Industrial Research*. 2011;70:199-203.
3. Tripathi YC. Food and nutrition potential of bamboo. *Minor Forest Products (MFP)-News*. 1998;8(1):10-11.
4. Chongtham N, David E, Sharma ML. Changes in nutrient components during ageing of emerging juvenile bamboo shoots. *International Journal of Food Sciences and Nutrition*. 2007;58(8):612-618.
5. Rai S. Edible Bamboo Shoot- A Review. *Bulletin of Arunachal Forest Research*. 2007;23(1&2):39-44.

6. Awol A. Nutrient, mineral and bioactive constituent evaluation of bamboo shoots grown in Masha area, South-West of Ethiopia. *American Scientific Research Journal for Engineering, Technology, and Sciences*. 2014;7(1):15-25.
7. Goyal AK, Pradhan S, Basistha BC, Sen A. Micropropagation and assessment of genetic fidelity of *Dendrocalamus strictus* (Roxb.) using RAPD and ISSR markers. *3 Biotech*. 2014;5:473-482.
8. Chongtham N, Sheena H, David E. Bamboo shoots: a rich source of dietary fibres. In: Klein F, Moller G, editors *Dietary fibres, fruit and vegetable consumption and health*. USA: Nova Science Publishers, 2009, p. 15–30.
9. Kalita T, Dutta U. A comparative study on indigenous usage of bamboo shoot in the health care practices in NE India. *The Clarion*. 2012;1(2):130–141.
10. Mustafa U, Naeem N, Masood S, Farooq Z. Effect of bamboo powder supplementation on physicochemical and organoleptic characteristics of fortified cookies. *Food Science and Technology*. 2016;4(1):7-13.
11. Goyal AK, Middha SK, Sen A. *In vitro* antioxidative profiling of different fractions of *Dendrocalamus strictus* (Roxb.) leaves extracts. *Free Radicals and Antioxidants*. 2011;1(2):42-48.
12. Akao Y, Seki N, Nakagawa Y, Yi H, Matusumoto K, Funaoka M, Maruyama W, Nozawa Y. A highly bioactive lignophenol derivative from bamboo lignin exhibit a potent activity to suppress apoptosis induced by oxidative stress in human neuroblastoma SH-SY5Y cells. *Bioorganic and Medicinal Chemistry*. 2004;12:4791-801.
13. Zhang JJ, Liu M, Li Y, Zhou T, Xu DP, Li HB. Nutrition values and biological activities of bamboo shoots and leaves. *International Journal of Food Nutrition and Safety*. 2016;7(2):98-108.
14. Singh SA, Bora TC, Singh NR. Preliminary phytochemical analysis and antimicrobial potential of fermented *Bambusa balcooa* shoots. *The Bioscan*. 2012;7(3):391-394.
15. Shi QT, Yang KS. Study on relationship between nutrients in bamboo shoots and human health, *Proceedings of the International Symposium on Industrial Use of Bamboo*. International Timber

- Organisation and Chinese Academy of Forestry, Beijing, China: Bamboo and its Use, 1992, p. 338-346.
16. Nongdam P, Tikendra L. The nutritional facts of bamboo shoots and their usage as important traditional foods of northeast India. *International Scholarly Research Notices*. 2014;2014(1):1-17.
 17. Choudhury D, Sahu JK, Sharma GD. Biochemistry of bitterness in bamboo shoots. *Assam University Journal of Science and Technology*. 2010;6(2):105-111.
 18. Chandramouli S, Viswanath S. Bamboo Shoots – An emerging new age health food. *Forestry Bulletin*. 2012;12(2):21-28.
 19. Haque MR, Bradbury JH. Total cyanide determination of plants and foods using the picrate and acid hydrolysis methods. *Food Chemistry*. 2002;77(1):107-114.
 20. Bhargava A, Kumbhare V, Srivastava A, Sahai A. Bamboo parts and seeds for additional source of nutrition. *Journal of Food Science and Technology*. 1996;33(2):145-146.
 21. Ferreira VLP, Marsaiolo AJ, Iaderoza M. Simplified methods for identification and detection of taxiphyllin in bamboo shoots. *Coletanea do Instituto de Tecnologia de Alimentos*. 1991;21(1):57-63.
 22. AOAC. *Official Methods of Analysis*. Association of Official Analytical Chemists, Washington, DC, 2000.
 23. Bray HG, Thorpe WV. Analysis of phenolic compounds of interest in metabolism. *Methods of Biochemical Analysis*. 1954;1:27-52.
 24. Daniel M. *Methods in Plant Chemistry and Economic Botany*, Kalyani Publishers, New Delhi, 1991, p. 74.
 25. Kalita P, Barma TK, Pal TK, Kalita R. Estimation of total flavonoids content (TFC) and antioxidant activities of methanolic whole plant extract of *Biophytum sensitive* Linn. *Journal of Drug Delivery and Therapeutics*. 2013;3:33-37.
 26. Schanderi SH. *Method in Food Analysis*. Academic Press, New York, 1970, p. 709.

27. Bradbury MG, Egan SV, Bradbur JH. Determination of all forms of cyanogens in cassava roots and cassava products using picrate paper kits. *Journal of the Science of Food and Agriculture*. 1999;79:593-601.
28. Karanja PN, Kenji GM, Njoroge SM, Sila DN, Onyango CA, Koaze H, Baba N. Variation of nutrients and functional properties within young shoots of a bamboo species (*Yushania alpina*) growing at Mt Elgon region in Western Kenya. *Journal of Food and Nutrition Research*. 2015;3:675-680.
29. Chongtham N, Bisht MS, Haorongbam S. Nutritional properties of bamboo shoots: potential and prospects for utilization as a health food. *Comprehensive Reviews in Food Science and Food Safety*. 2011;10(3):153-168.
30. Kumbhare V, Bhargava A. Effect of processing on nutritional value of central Indian bamboo shoots. Part I. *Journal of Food Science and Technology*. 2007;44(1):29-31.
31. Kalita C, Ganguly M, Devi A. Evaluation of antioxidant capacity and antimicrobial properties of ethnic *Bambuseae* species and identification of the active components. *International Journal of Pharmaceutical and Biological Archive*. 2016;7:61-71.
32. Badwaik LS, Gautam G, Deka SC. Influence of blanching on antioxidant, nutritional and physical properties of bamboo shoot. *Journal of Agricultural Sciences*. 2015;10(3):140-150.
33. Wei ZY, Wang JS, Cao ZY, Zhang ZY, Li XJ. Effect of high temperature and high humidity on contents of protein, amino acids and sugars in wheat grains. *Journal of Chinese Cereals and Oils Association*. 2009;24:5-11.
34. Udensi AE, Arisa UN, Ikpa E. Effects of soaking and boiling and autoclaving on the nutritional quality of *Mucuna flagellipes* ("ukpo"). *African Journal of Biochemistry Research*. 2010;4:47-50.
35. Pandey AK, Ojha V. Precooking processing of bamboo shoots for removal of anti-nutrients. *Journal of Food Science and Technology*. 2014;51:43-50.
36. Bhatt BP, Singh K, Singh A. Nutritional values of some commercial edible bamboo species of the North Eastern Himalayan region. *Indian Journal of Bamboo and Rattan*. 2005;4:111-124.

37. Zhang JJ, Ji R, Hu YQ, Chen JC, Ye XQ. Effect of three cooking methods on nutrient components and antioxidant capacities of bamboo shoot (*Phyllostachys praecox* C.D. Chu et C.S. Chao). *Journal of Zhejiang University Science*. 2011;12(9):752-759.
38. Rajyalakshmi P, Geervani P. Nutritive value of the foods cultivated and consumed by the tribals of South India. *Plant Foods for Human Nutrition*. 1994;46:53-61.
39. Singhal P, Bal LM, Satya S, Sudhakar P, Naik SN. Bamboo shoots: a novel source of nutrition and medicine. *Critical Reviews in Food Science and Nutrition*. 2013;53(5):517-534.
40. Rawat K, Sharma V, Saini N, Chongtham N, Bisht MS. Impact of different boiling and soaking treatments on the release and retention of antinutrients and nutrients from the edible shoots of three bamboo species. *American Journal of Food Science and Nutrition Research*. 2016;3:31-41.
41. Chongtham N, Sharma ML, David E. A comparative study of nutrient components of freshly harvested, fermented and canned bamboo shoots of *Dendrocalamus giganteus* Munro. *The Journal of the American Bamboo Society*. 2008;21(1):33-39.
42. Rumm-Kreuter D, Demmel I. Comparison of vitamin losses in vegetables due to various cooking methods. *Journal of Nutritional Science and Vitaminology*. 1990;36:7-15.
43. Afify AM, El-Beltagi HS, El-Salam SM, Omran AA. Biochemical changes in phenols flavonoids tannins vitamin E β -carotene and antioxidant activity during soaking of three white sorghum varieties. *Asian Pacific Journal of Tropical Biomedicine*. 2012;2:203-209.
44. Bajwa HK, Chongtham N, Koul A, Bisht MS. Effects of processing and preservation on phenols and phytosterols content in bamboo shoots. 10th World Bamboo Congress, Korea, 2015.
45. Yamaguchi T, Katsuda M, Oda Y, Terao J, Kanazawa K, Oshima S, Inakuma T, Ishiguro Y, Takamura H, Matoba T. Influence of polyphenol and ascorbate oxidases during cooking process on the radical scavenging activity of vegetables. *Food Science and Technology Research*. 2003;9:79-83.
46. Stewart AJ, Bozonnet S, Mullen W, Jenkins GI, Lean MEJ, Crozier A. Occurrence of flavonols in tomatoes and tomato based products. *Journal of Agricultural and Food Chemistry*. 2000;48:2663-2669.

47. Mina T, Giri S, Shantibala. Contents of antinutritional substances of bamboo shoots and relation to these to sensory characters of their intact and fermented forms. *The Bioscan*. 2014;9:1123-1126.
48. Sarangthem K, Singh T. Fermentation decreases the antinutritional content in bamboo shoots. *International Journal of Current Microbiology and Applied Sciences*. 2013;2(11):361-369.
49. Rana B, Awasthi P, Kumbhar BK. Optimization of processing conditions for cyanide content reduction in fresh bamboo shoot during NaCl treatment by response surface methodology. *Journal of Food Science and Technology*. 2012;49:103-109.
50. Ogbadoyi EO, Makun HA, Bamigbade RO, Oyewale AO, Oladiran JA. The effect of processing and preservation methods on the oxalate levels of some Nigerian leafy vegetables. *Biokemistri*. 2006;18:121-125.
51. Jaiwunglok P, Tia S, Yoovidhya T. Effects of temperature and pH on taxiphyllin degradation in bamboo shoots. In: *Food Innovation Asia Conference, Indigenous Food Research and Development to Global Market*, BITEC Bangkok, Thailand, June 17–18, 2010.

Table 1. Proximate composition of fresh and processed bamboo shoots (g/100g on dry weight basis).

Proximate analysis	Variety	Treatments						Means \pm SD
		Control	Soaking in H ₂ O for 30 min.	Boiling in H ₂ O for 10 min.	Boiling in 1% NaCl for 10 min.	Boiling in 5% NaCl for 10 min.	Steaming for 10 min.	
Moisture	<i>Bambusatulda</i> (Jati)	90.76 \pm 1.211	90.79 \pm 1.220	91.90 \pm 0.956	90.41 \pm 1.232	90.06 \pm 1.204	89.27 \pm 1.331	90.53 \pm 1.298 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	92.29 \pm 1.176	92.32 \pm 1.192	93.24 \pm 0.932	91.98 \pm 1.175	91.83 \pm 1.180	91.46 \pm 1.050	92.19 \pm 1.100 ^b
	<i>Bambusabalcooa</i> (bhaluka)	91.65 \pm 0.498	91.66 \pm 0.502	92.46 \pm 0.477	91.43 \pm 0.426	91.18 \pm 0.348	91.08 \pm 0.640	91.58 \pm 0.620 ^b
	Mean	91.56 \pm 1.103 ^{ab}	91.59 \pm 1.108 ^{ab}	92.53 \pm 0.918 ^a	91.27 \pm 1.117 ^{ab}	91.02 \pm 1.158 ^b	90.61 \pm 1.361 ^b	
Crude protein	<i>Bambusatulda</i> (Jati)	16.42 \pm 0.015	16.49 \pm 0.029	14.66 \pm 0.017	14.65 \pm 0.029	14.66 \pm 0.032	14.63 \pm 0.026	15.24 \pm 0.856 ^a
	<i>Dendrocalamus hamiltonii</i> (Kako)	13.56 \pm 0.025	13.54 \pm 0.030	11.76 \pm 0.030	11.76 \pm 0.030	11.75 \pm 0.035	11.75 \pm 0.057	12.35 \pm 0.871 ^c
	<i>Bambusabalcooa</i> (bhaluka)	14.44 \pm 0.036	14.42 \pm 0.026	12.66 \pm 0.040	12.66 \pm 0.040	12.66 \pm 0.045	12.65 \pm 0.040	13.25 \pm 0.859 ^b
	Mean	14.81 \pm 1.272 ^a	14.79 \pm 1.272 ^a	13.03 \pm 1.285 ^c	13.02 \pm 1.284 ^b	13.02 \pm 1.286 ^b	13.01 \pm 1.278 ^b	
Crude Fat	<i>Bambusatulda</i> (Jati)	1.48 \pm 0.457	1.45 \pm 0.447	1.31 \pm 0.492	1.30 \pm 0.490	1.30 \pm 0.489	1.29 \pm 0.508	1.35 \pm 0.412 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	1.83 \pm 0.563	1.80 \pm 0.569	1.75 \pm 0.060	1.75 \pm 0.056	1.74 \pm 0.055	1.78 \pm 0.101	1.77 \pm 0.281 ^b
	<i>Bambusabalcooa</i> (bhaluka)	1.49 \pm 0.091	1.48 \pm 0.093	1.39 \pm 0.154	1.38 \pm 0.150	1.37 \pm 0.150	1.36 \pm 0.149	1.41 \pm 0.126 ^b
	Mean	1.60 \pm 0.404 ^a	1.58 \pm 0.401 ^a	1.48 \pm 0.332 ^b	1.48 \pm 0.331 ^b	1.470 \pm 0.327 ^b	1.48 \pm 0.355 ^a	
Crude fibre	<i>Bambusatulda</i> (Jati)	5.28 \pm 0.253	5.17 \pm 0.203	5.07 \pm 0.206	5.00 \pm 0.232	4.97 \pm 0.215	5.07 \pm 0.258	5.09 \pm 0.221 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	4.09 \pm 0.038	4.02 \pm 0.036	3.97 \pm 0.030	4.27 \pm 0.586	4.25 \pm 0.577	4.26 \pm 0.560	4.15 \pm 0.365 ^c
	<i>Bambusabalcooa</i> (bhaluka)	5.57 \pm 0.091	5.54 \pm 0.061	5.45 \pm 0.060	5.39 \pm 0.110	5.34 \pm 0.125	5.47 \pm 0.100	5.46 \pm 0.114 ^a
	Mean	4.98 \pm 0.692 ^a	4.91 \pm 0.694 ^a	4.83 \pm 0.676 ^a	4.89 \pm 0.584 ^a	4.85 \pm 0.572 ^a	4.94 \pm 0.190 ^a	
Total Carbohydrate	<i>Bambusatulda</i> (Jati)	6.58 \pm 0.298	6.52 \pm 0.306	8.39 \pm 0.212	6.12 \pm 0.231	6.01 \pm 0.231	6.17 \pm 0.286	6.63 \pm 0.864 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	5.37 \pm 0.252	5.07 \pm 0.208	7.63 \pm 0.062	4.82 \pm 0.040	4.66 \pm 0.076	4.72 \pm 0.076	5.38 \pm 1.073 ^c
	<i>Bambusabalcooa</i> (bhaluka)	6.97 \pm 0.049	6.89 \pm 0.015	9.03 \pm 0.155	6.61 \pm 0.170	6.47 \pm 0.144	6.59 \pm 0.202	7.09 \pm 0.918 ^a
	Mean	6.30 \pm 0.749 ^b	6.16 \pm 0.854 ^b	8.35 \pm 0.623 ^a	5.85 \pm 0.816 ^c	5.71 \pm 0.828 ^c	5.82 \pm 0.868 ^c	
Ash	<i>Bambusatulda</i> (Jati)	1.10 \pm 0.021	1.07 \pm 0.030	0.94 \pm 0.036	0.94 \pm 0.042	0.95 \pm 0.032	0.98 \pm 0.040	1.00 \pm 0.033 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	0.89 \pm 0.021	0.86 \pm 0.026	0.79 \pm 0.032	0.78 \pm 0.023	0.79 \pm 0.045	0.81 \pm 0.040	0.83 \pm 0.031 ^b
	<i>Bambusabalcooa</i> (bhaluka)	1.54 \pm 0.020	1.52 \pm 0.021	1.43 \pm 0.030	1.44 \pm 0.021	1.44 \pm 0.023	1.47 \pm 0.012	1.47 \pm 0.048 ^b
	Mean	1.18 \pm 0.286 ^a	1.16 \pm 0.287 ^a	1.05 \pm 0.290 ^b	1.05 \pm 0.296 ^b	1.06 \pm 0.294 ^b	1.08 \pm 0.297 ^b	

Means in the same column with different letters are significantly ($p < 0.05$) different.

Means \pm standard deviation of three determinations.

Table 2. Mineral composition of fresh and processed bamboo shoots (mg/100g on dry weight basis).

Mineral analysis	Variety	Treatments						Means \pm SD
		Control	Soaking in H ₂ O for 30 min.	Boiling in H ₂ O for 10 min.	Boiling in 1% NaCl for 10 min.	Boiling in 5% NaCl for 10 min.	Steaming for 10 min.	
Potassium	<i>Bambusatulda</i> (Jati)	813.33 \pm 5.859	812.33 \pm 4.933	789.33 \pm 7.767	803.67 \pm 6.429	803.67 \pm 6.658	797.67 \pm 2.082	803.33 \pm 9.840 ^a
	<i>Dendrocalamus hamiltonii</i> (Kako)	645.33 \pm 5.508	644.67 \pm 6.028	629.67 \pm 9.074	636.33 \pm 6.506	637.00 \pm 5.000	636.00 \pm 5.568	638.17 \pm 7.748 ^a
	<i>Bambusabalcooa</i> (bhaluka)	840.33 \pm 4.509	840.00 \pm 4.583	819.33 \pm 5.686	833.67 \pm 2.887	833.33 \pm 4.933	832.33 \pm 6.506	833.17 \pm 8.291 ^a
	Mean	766.33 \pm 91.616 ^a	765.67 \pm 91.649 ^{ab}	746.11 \pm 88.542 ^a	757.89 \pm 92.212 ^{bc}	758.00 \pm 91.782 ^{bc}	755.33 \pm 90.857 ^c	
Phosphorous	<i>Bambusatulda</i> (Jati)	654.00 \pm 4.000	608.00 \pm 7.000	420.00 \pm 3.000	422.00 \pm 3.000	422.00 \pm 2.000	590.00 \pm 3.000	519.00 \pm 102.787 ^a
	<i>Dendrocalamus hamiltonii</i> (Kako)	572.00 \pm 4.000	540.00 \pm 5.000	320.00 \pm 5.000	460.00 \pm 3.000	470.00 \pm 5.000	480.00 \pm 6.000	473.00 \pm 81.905 ^c
	<i>Bambusabalcooa</i> (bhaluka)	620.00 \pm 5.000	560.00 \pm 2.000	450.00 \pm 3.000	460.00 \pm 8.000	465.00 \pm 5.000	5.20 \pm 2.000	512.00 \pm 65.535 ^b
	Mean	615.33 \pm 35.878 ^a	569.33 \pm 30.586 ^b	396.67 \pm 59.040 ^b	447.33 \pm 19.532 ^d	452.33 \pm 23.146 ^d	530.00 \pm 48.345 ^c	
Iron	<i>Bambusatulda</i> (Jati)	9.34 \pm 0.025	9.32 \pm 0.031	8.81 \pm 0.023	9.22 \pm 0.025	9.23 \pm 0.025	9.02 \pm 0.015	9.16 \pm 0.193 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	10.93 \pm 0.020	10.92 \pm 0.015	10.37 \pm 0.020	10.87 \pm 0.010	10.87 \pm 0.021	10.81 \pm 0.012	10.80 \pm 0.201 ^a
	<i>Bambusabalcooa</i> (bhaluka)	9.07 \pm 0.067	8.99 \pm 0.015	8.59 \pm 0.052	8.99 \pm 0.061	9.00 \pm 0.064	8.95 \pm 0.020	8.93 \pm 0.167 ^c
	Mean	9.78 \pm 0.871 ^a	9.74 \pm 0.897 ^{ab}	9.26 \pm 0.842 ^d	9.69 \pm 0.889 ^b	9.70 \pm 0.887 ^b	9.59 \pm 0.911 ^c	
Calcium	<i>Bambusatulda</i> (Jati)	4.83 \pm 0.036	4.82 \pm 0.046	4.72 \pm 0.035	4.76 \pm 0.038	4.76 \pm 0.038	4.75 \pm 0.025	4.77 \pm 0.051 ^e
	<i>Dendrocalamus hamiltonii</i> (Kako)	5.76 \pm 0.060	5.75 \pm 0.050	5.74 \pm 0.075	5.74 \pm 0.075	5.75 \pm 0.080	5.75 \pm 0.066	5.75 \pm 0.058 ^b
	<i>Bambusabalcooa</i> (bhaluka)	6.70 \pm 0.086	6.69 \pm 0.090	6.59 \pm 0.085	6.60 \pm 0.085	6.60 \pm 0.081	6.63 \pm 0.066	6.64 \pm 0.083 ^a
	Mean	5.76 \pm 0.813 ^a	5.75 \pm 0.813 ^a	5.69 \pm 0.813 ^a	5.70 \pm 0.801 ^a	5.70 \pm 0.803 ^a	5.71 \pm 0.815 ^a	

Means in the same column with different letters are significantly ($p < 0.05$) different.

Means \pm standard deviation of three determinations.

Table 3. Vitamin C content, secondary metabolites, and antinutritional composition of fresh and processed bamboo shoots.

Biochemical parameters	Variety	Treatments						Means \pm SD
		Control	Soaking in H ₂ O for 30 min.	Boiling in H ₂ O for 10 min.	Boiling in 1% NaCl for 10 min.	Boiling in 5% NaCl for 10 min.	Steaming for 10 min.	
Ascorbic acid (mg/100g fresh wt)	<i>Bambusatulda</i> (Jati)	2.60 \pm 0.200	2.45 \pm 0.154	1.83 \pm 0.076	1.86 \pm 0.070	1.86 \pm 0.070	1.92 \pm 0.065	2.09 \pm 0.337 ^d
	<i>Dendrocalamus hamiltonii</i> (Kako)	2.47 \pm 0.025	2.38 \pm 0.070	1.82 \pm 0.072	1.88 \pm 0.074	1.88 \pm 0.068	1.93 \pm 0.074	2.06 \pm 0.276 ^d
	<i>Bambusabalcooa</i> (bhaluka)	3.31 \pm 0.338	3.23 \pm 0.331	2.53 \pm 0.208	2.58 \pm 0.208	2.56 \pm 0.204	2.72 \pm 0.227	2.82 \pm 0.397 ^d
	Mean	2.79 \pm 0.437 ^a	2.69 \pm 0.450 ^a	2.06 \pm 0.372 ^b	2.11 \pm 0.376 ^b	2.10 \pm 0.362 ^b	2.19 \pm 0.418 ^b	
Alkaloid (%) (on dry weight basis).	<i>Bambusatulda</i> (Jati)	20.38 \pm 1.370	20.37 \pm 1.380	19.58 \pm 0.887	19.55 \pm 0.890	19.17 \pm 0.707	19.60 \pm 0.873	19.78 \pm 0.996 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	20.25 \pm 0.983	20.23 \pm 0.972	20.07 \pm 0.918	20.05 \pm 0.921	20.02 \pm 0.924	20.06 \pm 0.933	20.11 \pm 0.797 ^{bc}
	<i>Bambusabalcooa</i> (bhaluka)	20.70 \pm 0.262	20.69 \pm 0.258	20.56 \pm 0.231	20.56 \pm 0.242	20.50 \pm 0.289	20.55 \pm 0.211	20.60 \pm 0.224 ^a
	Mean	20.45 \pm 0.876 ^a	20.43 \pm 0.878 ^a	20.07 \pm 0.776 ^b	20.05 \pm 0.783 ^b	19.90 \pm 0.836 ^b	20.07 \pm 0.767 ^b	
Phenol (mg Catechol Equivalent/100 g dry wt).	<i>Bambusatulda</i> (Jati)	493.64 \pm 5.178	493.27 \pm 5.136	399.34 \pm 0.610	400.45 \pm 0.558	400.30 \pm 0.520	493.83 \pm 5.184	446.80 \pm 48.231 ^c
	<i>Dendrocalamus hamiltonii</i> (Kako)	506.62 \pm 4.447	506.35 \pm 4.511	470.90 \pm 2.513	470.55 \pm 2.211	470.41 \pm 2.554	507.06 \pm 4.233	488.65 \pm 18.791 ^a
	<i>Bambusabalcooa</i> (bhaluka)	504.70 \pm 4.019	504.28 \pm 3.823	441.70 \pm 5.451	441.63 \pm 5.084	441.70 \pm 5.057	441.72 \pm 5.446	462.62 \pm 30.734 ^b
	Mean	501.65 \pm 7.247 ^a	501.30 \pm 7.240 ^a	437.31 \pm 31.30 ^b	437.55 \pm 30.635 ^b	437.47 \pm 30.655 ^b	480.87 \pm 30.22 ^b	
Flavonoid contents (mg QE/100g dry wt).	<i>Bambusatulda</i> (Jati)	370.00 \pm 2.000	370.00 \pm 3.000	347.00 \pm 2.000	347.00 \pm 1.000	344.00 \pm 2.000	367.33 \pm 2.517	357.56 \pm 12.108 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	352.67 \pm 1.528	351.33 \pm 1.528	330.67 \pm 1.155	329.00 \pm 2.00	327.00 \pm 1.000	350.33 \pm 1.528	340.17 \pm 11.744 ^c
	<i>Bambusabalcooa</i> (bhaluka)	445.00 \pm 3.000	444.67 \pm 3.055	413.67 \pm 0.577	413.33 \pm 1.155	313.33 \pm 2.517	442.00 \pm 2.646	412.00 \pm 47.560 ^a
	Mean	389.22 \pm 42.546 ^a	388.67 \pm 42.831 ^a	363.78 \pm 38.09 ^b	363.11 \pm 38.485 ^b	328.11 \pm 13.411 ^c	386.56 \pm 42.27 ^b	
Tannin (mg TAE/100g dry wt.)	<i>Bambusatulda</i> (Jati)	204.20 \pm 0.200	204.10 \pm 0.265	184.70 \pm 0.700	144.37 \pm 0.306	104.94 \pm 0.040	124.48 \pm 0.030	177.73 \pm 28.072 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	204.20 \pm 0.200	204.10 \pm 0.265	184.40 \pm 0.400	184.40 \pm 0.400	124.77 \pm 0.751	164.51 \pm 0.040	187.53 \pm 47.888 ^b
	<i>Bambusabalcooa</i> (bhaluka)	184.30 \pm 0.300	184.23 \pm 0.208	144.70 \pm 0.700	144.37 \pm 0.306	104.94 \pm 0.040	124.48 \pm 0.030	147.84 \pm 29.889 ^c
	Mean	210.83 \pm 26.402 ^a	210.67 \pm 26.233 ^a	164.53 \pm 17.21 ^b	157.59 \pm 20.114 ^b	118.17 \pm 9.933 ^c	164.40 \pm 34.52 ^b	
Cyanide (ppm fresh wt)	<i>Bambusatulda</i> (Jati)	875.16 \pm 13.527	463.28 \pm 5.631	20.41 \pm 1.534	17.31 \pm 2.085	16.09 \pm 1.935	457.02 \pm 8.890	308.21 \pm 330.936 ^b
	<i>Dendrocalamus hamiltonii</i> (Kako)	1067.87 \pm 18.326	563.64 \pm 45.169	44.17 \pm 1.187	43.45 \pm 1.202	42.96 \pm 0.987	530.01 \pm 41.67	382.02 \pm 392.226 ^b
	<i>Bambusabalcooa</i> (bhaluka)	866.91 \pm 5.166	468.89 \pm 9.049	27.51 \pm 1.808	25.27 \pm 3.312	25.17 \pm 3.267	461.36 \pm 14.89	312.52 \pm 325.532 ^b
	Mean	936.65 \pm 99.175 ^a	498.60 \pm 54.073 ^b	30.70 \pm 10.642 ^c	28.68 \pm 11.781 ^c	28.07 \pm 11.997 ^c	482.80 \pm 42.03 ^b	

Means in the same column with different letters are significantly ($p < 0.05$) different.

Means \pm standard deviation of three determinations.