

# EXOGENOUSLY APPLIED GIBBERELIC ACID ENHANCES GROWTH AND SALINITY STRESS TOLERANCE OF MAIZE THROUGH MODULATING THE MORPHO-PHYSIOLOGICAL AND PHOTOSYNTHETIC ATTRIBUTES

## ABSTRACT

Soil salinity is the major limiting factor restricting plant growth and development. Little is known about the comparative and combined effects of gibberellic acid (GA<sub>3</sub>) seed priming and foliar application on maize under salt stress. The current study determined the impact of different concentrations of GA<sub>3</sub> on morpho-physiological and photosynthetic attributes of maize seedlings under salinity stress treatments (no salinity and severe salinity-15 dSm<sup>-1</sup>). The GA<sub>3</sub> treatments consisted of 1mM, 2mM, 3mM, 4mM and 5mM GA<sub>3</sub> seed priming and exogenous application in salt condition. Salt stress particularly at 15 dSm<sup>-1</sup> reduced the length of shoots and roots, fresh and dry weights, chlorophyll, lycopene, beta-carotene and carotenoid contents in maize plants. Nevertheless, the application of GA<sub>3</sub> improved maize growth under salt stress. Compared with salt, the T4 recorded the highest increase in roots and shoots length, roots fresh and dry weights, shoots fresh and dry weights, chlorophyll content undersalt stress. These results indicated that 2mM GA<sub>3</sub> priming and exogenous application could be used as an effective tool for improving the maize growth and development in salt contaminated soils.

**Keywords:** salinity; gibberellic acid; maize; seed priming; exogenous application; growth; photosynthetic attributes

## 1. INTRODUCTION

*Zea mays* L., known as maize, is a common cereal crop that is grown all over the world for use as forage and as a source of food grains for both human and animal consumption. Additionally, it gives various industries access to raw materials[1]. Under field conditions, the maize crop is exposed to a number of abiotic challenges, including soil salinity, drought, light, and temperature, which can significantly reduce its production [2]. Soil salinity is one of the main abiotic stress that restricts crop growth and productivity. According to estimates, soil salinity affects around 6% of all arable land worldwide [3]. The growth, development, and yield of field crops are all significantly hampered by soil salinity, in addition to lowering seed emergence and germination rates [4, 5]. Additionally, high salt levels in the soil cause stomata to close and harm the photosynthetic apparatus and chlorophyll content [6–8]. Reactive oxygen species (ROS) are continuously created by metabolic processes in plants, with production increasing in response to environmental stress [4]. When ROS generation rises, lipids, membranes, nucleic acids, and proteins can be destroyed, which causes cellular machinery to fail [9–11]. Ion equilibrium is

disturbed as a result of the overproduction of ROS during salt stress [12, 13]. Plants have evolved a powerful antioxidant defense system, including the antioxidant enzymes superoxide dismutase (SOD), peroxidase (POD), ascorbate peroxidase (APX), and catalase (CAT), to limit the oxidative damage [14].

Chemical addition to plants, either as an exogenous foliar application or seed treatments, may stimulate their physiological systems, resulting in an improvement in plant growth [15]. The phenotypes of plants can change from seed germination through senescence, for instance, when seed priming with plant growth regulators is used [16]. Gibberellic Acid (GA<sub>3</sub>), a type of growth hormone that occurs naturally, controls how plants grow and develop [17]. The GA<sub>3</sub> are linked to numerous aspects of plant growth and development, including seed germination, hypocotyl lengthening, leaf expansion, floral initiation, uniform flowering, floral organ development, shortened flowering times, increased flower number and size, and induction of a few hydrolytic enzymes in the aleurone of cereal grains [18–20]. According to reports, growth regulators such as GA<sub>3</sub> can lessen the salinity-related germination inhibitory effect [21–24]. According to Kaur et al. [25], GA<sub>3</sub> at a concentration of 6 μM promotes enhanced seedling development under salt stress. According to Ashraf et al. [26], treatment of GA<sub>3</sub> increased wheat's nutrient uptake, dry weight, plant height, leaf area, and yield in saline conditions. Additionally, there is proof that GA<sub>3</sub> can significantly reverse the growth inhibition caused by NaCl in rice [27]. According to Starck and Kozinska [28], the GA<sub>3</sub> slightly changed the ion ratios in beans while increasing P and Ca<sup>2+</sup> absorption and decreasing Na<sup>+</sup> absorption. According to Bejaoui [29], the benefits of exogenously given GA<sub>3</sub> in reducing salt stress may be brought on by the activation of particular enzymes involved in RNA and protein synthesis. Aloni and Pressman [30] suggested a potential relationship between salt and the GA<sub>3</sub> effect on petiole elongation, cellular disintegration, and bolting in celery as a defense response to stressors in plants.

Due to high levels of salt in irrigation water, seed germination and stand establishment in maize farms are frequently poor. Therefore, the goal of this experiment was to investigate how priming and exogenous application of GA<sub>3</sub> treatments on maize affected their ability to maintain normal germination and growth under salinity stress.

## **2. MATERIALS AND METHODS**

### **2.1 Plant Materials and Chemicals**

The experiment was carried out at the Department of Seed Science and Technology of Bangladesh Agricultural University, Mymensingh, using maize, widely cultivated in many regions of Bangladesh. The chemicals, Gibberellic Acid (GA<sub>3</sub>), SA (Sigma-Aldrich), sodium hypochlorite (Sigma-Aldrich), and Hyponex (Osaka, Japan) nutrient solution were used as an analytical grade in this study.

### **2.2 Experiment at pre-seedling stage**

Uniform in appearance maize seeds were sorted out and surface sterilized with 1% sodium hypochlorite for 5 min and then washed 3-4 times with dH<sub>2</sub>O. For the seed priming experiment, the seeds were soaked in 1mM, 2mM, 3mM, 4mM and 5mM GA<sub>3</sub> for 60 minutes, and the control experiment seeds were washed in distilled water for several times in normal laboratory (the room temperature was 25±1°C and relative humidity was 95%) conditions. After that, treated seeds were placed in a petri-dish (150×20 mm diameter) having three layers of wetted-Whatman filter papers and kept for 7 days for the germination study. Fifteen treated seeds were placed in each petri-dish. Thus the treatments in this study was presented below

Table 1 : List of treatments and Concentration of priming agent

Treatment	Concentration of priming agent
T1	Control
T2	Salt
T3	Salt+1mM GA <sub>3</sub>
T4	Salt+2mM GA <sub>3</sub>
T5	Salt+3mM GA <sub>3</sub>
T6	Salt+4mM GA <sub>3</sub>
T7	Salt+5mM GA <sub>3</sub>

The experiment was conducted with a completely randomized block design having three replicates. Germination percentage (GP) and seed vigor index (Rauf et al. 2020) were computed with the following equations [31]:

$$\text{Germination percentage GP} = \frac{\text{Total number of seeds germinated}}{\text{Total number of seeds placed in germination}} \times 100 \dots\dots(1)$$

$$\text{Seed vigor index SVI} = \text{GP} \times \text{seedling length (cm)} \dots\dots\dots(2)$$

### 2.3 Experiment at seedling stage

Uniformly germinated seeds were placed in plastic pots (22 cm in height and 25 cm in diameter) filled with soil (4 seedlings per pot). The water was mixed with nutrient solution Hyponex (Osaka, Japan) containing nitrogen, phosphorous, potassium, and other micronutrients. The nutrient solution (2 ml mixed with water for per pot) was applied twice in a week in the pots. After 12 days, seedlings were exogenously treated with different concentrations of GA<sub>3</sub> for four times in four days (8 ml per plant per spray). After 4 days of GA<sub>3</sub> treatment, morphological and physiological data were collected from three plants.

## 2.4 Relative water content measurement

Relative water content (RWC) was determined followed by the standard procedure of Mostofa & Fujita [32]. In the case of RWC measurement, leaf samples were collected after 14 days of transplanting and then fresh weight (FW) of leaves were taken and immersed in dH<sub>2</sub>O and kept for 4 hr. After that, excess water was removed from the turgid leaves with a paper towel and turgid weight (TW) was recorded instantly. After that leaves were oven dried at 70 °C for 48hrs and dry weight (DW) was recorded. The RWC was calculated according to the following formula:  $RWC (\%) = (FW - DW)/(TW - DW) \times 100$ .....(3)

## 2.6 Measurement of leaf chlorophyll contents

Based on the procedure outlined by Lichtenthaler [33], the concentrations of the photosynthetic leaf pigments chlorophyll, lycopene, beta carotene, and carotenoids were measured spectrophotometrically. Fresh leaves weighing 0.5g were picked and placed in a tiny vial with 10 mL of 80% ethanol. For the purpose of extracting the pigments, the containers were covered with aluminum foil and kept in the dark for 7 days. A spectrophotometer was used to measure the absorbance from leaf extraction at wavelengths of 663, 645, 505, and 453 nm for the concentrations of chlorophyll, lycopene, beta carotene, and carotenoids (Shimadzu UV-2550, Kyoto, Japan). The following formulae were used to compute the photosynthesis pigments:

$$\text{Total Chlorophyll} = \text{Chlorophyll a} + \text{Chlorophyll b} \dots \dots \dots (4)$$

$$\text{Chlorophyll a} = 0.999 \times A_{663} - 0.0989 \times A_{645} \dots \dots \dots (5)$$

$$\text{Chlorophyll b} = -0.328 \times A_{663} + 1.77 \times A_{645} \dots \dots \dots (6)$$

$$\text{Lycopene} = 0.0458 \times A_{663} + 0.204 \times A_{645} + 0.372 \times A_{505} - 0.0806 \times A_{453} \dots \dots \dots (7)$$

$$\text{Beta-carotene} = 0.216 \times A_{663} - 1.22 \times A_{645} - 0.304 \times A_{505} + 0.452 \times A_{453} \dots \dots \dots (8)$$

$$\text{Carotenoids} = A_{480} + (0.114 \times A_{663} - 0.638 \times A_{645} \dots \dots \dots (9)$$

**2.8 Statistical Analysis** Data collected for each parameter were subjected to one way ANOVA using Minitab 17 statistical software (Minitab Inc., State College, PA, USA). The statistical differences among the mean values of different treatments were compared using Tukey's pair-wise comparisons ( $P < 0.05$ ).

## 3. RESULT

### 3.1. Priming boosts germination indices and traits of seedlings under salt stress

The impacts of GA<sub>3</sub> priming and exogenous application on the germination indices of maize under salt stress are displayed in Figure 1. The findings show that salt stress significantly

reduced GP by 17.39% compared to the control condition. Priming and exogenous application of GA<sub>3</sub> showed a significant effect on GP (Figure 1a). While the highest GP (90%) was recorded for T2 (2mM GA<sub>3</sub>) treatment, the lowest GP (63.33%) was recorded for salt stressed seeds (Figure 1a). In the case of shoot and root length, significant variations were found for different priming treatments compared to salt stress. The highest shoot and root length under stress condition was observed in treatment T4 (Salt+2mM GA<sub>3</sub>) 43.1cm and 18.1cm, respectively. Shoot length decreased by 24.61% in stress condition while GA<sub>3</sub> treatments increased shoot length by 46.59, 66.67, 40.54, 35.03 and 23.37%, respectively for 1mM, 2mM, 3mM, 4mM and 5mM GA<sub>3</sub> treated seeds under stress condition (Figure 1b). Salt stress significantly reduced root length while priming with GA<sub>3</sub> increased root length under salt settings. Root length decreased by 56.98% in stress condition while GA<sub>3</sub> treatments increased shoot length by 58.77, 94.73, 53.51, 34.21 and 6.14% respectively, for 1mM, 2mM, 3mM, 4mM and 5mM GA<sub>3</sub> treated seeds under stress condition (Figure 1c). Similarly, different priming conditions increased SVI but salt stress significantly reduced SVI (48.54%) compared with the control. The results indicated that SVI increased by 89.47, 147.98, 97.28, 77.37 and 49.75%, respectively, for 1mM, 2mM, 3mM, 4mM and 5mM GA<sub>3</sub> priming compared to salt stress (Figure 1d).

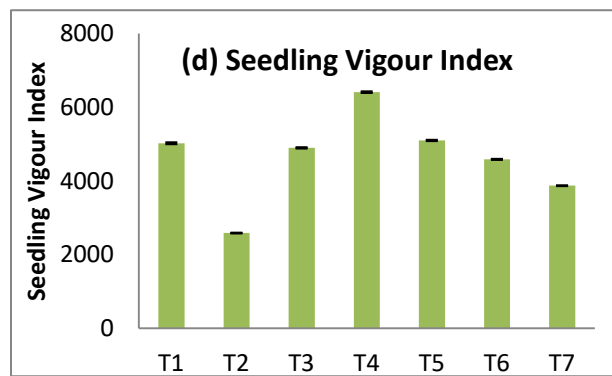
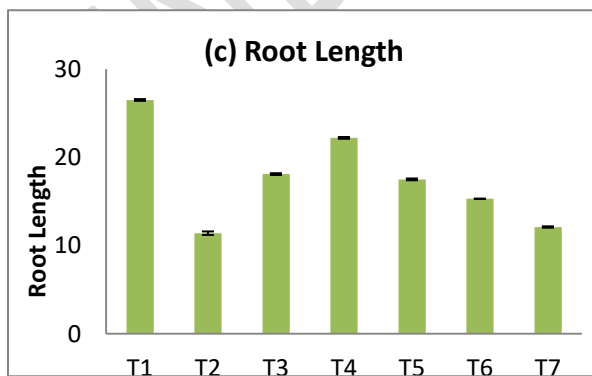
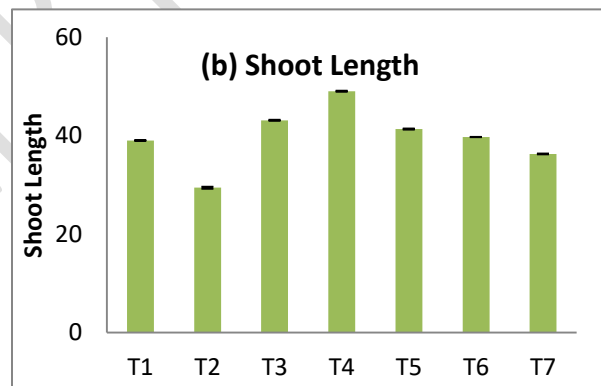
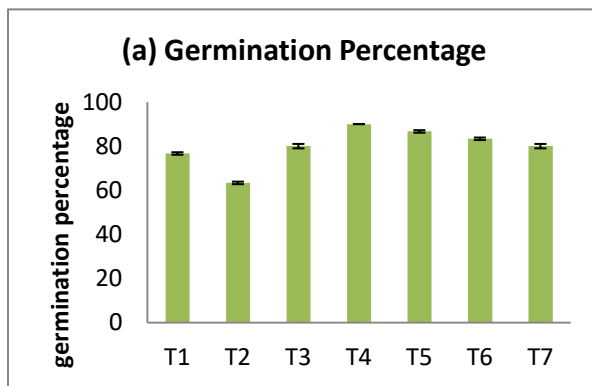


Figure 1. Effects of GA<sub>3</sub> priming and exogenous application on the germination and growth indices of maize under salt stress. (a) Germination percentage; (b) Shoot Length; (c) Root Length; (d) Seed vigor index. The error bar represents standard error.

### 3.2 Priming increases fresh and dry weight of seedlings under salt stress

To assess the effects of salt stress and stress-decreasing acts of GA<sub>3</sub> on the weight of maize seedlings, we recorded the fresh and dry weight of seedlings. All the priming conditions significantly increased fresh and dry weight of shoot and root under salt stress. Shoot fresh weight (SFW), shoot dry weight (SDW), and root fresh and dry weight (RFW, RDW) were decreased by salt stress (Figure 2). In case of shoot fresh and dry weight was highest at T4 treatment 2.78g and 0.17g, respectively under stress condition (Figure 1a, c). Similarly, root fresh and dry weight was highest at T4 treatment 0.75g and 0.11g, respectively under salt stress (Figure 1b, d). All other treatments increased shoot and root fresh and dry weight under stress condition when seeds were treated with GA<sub>3</sub> (Figure 2).

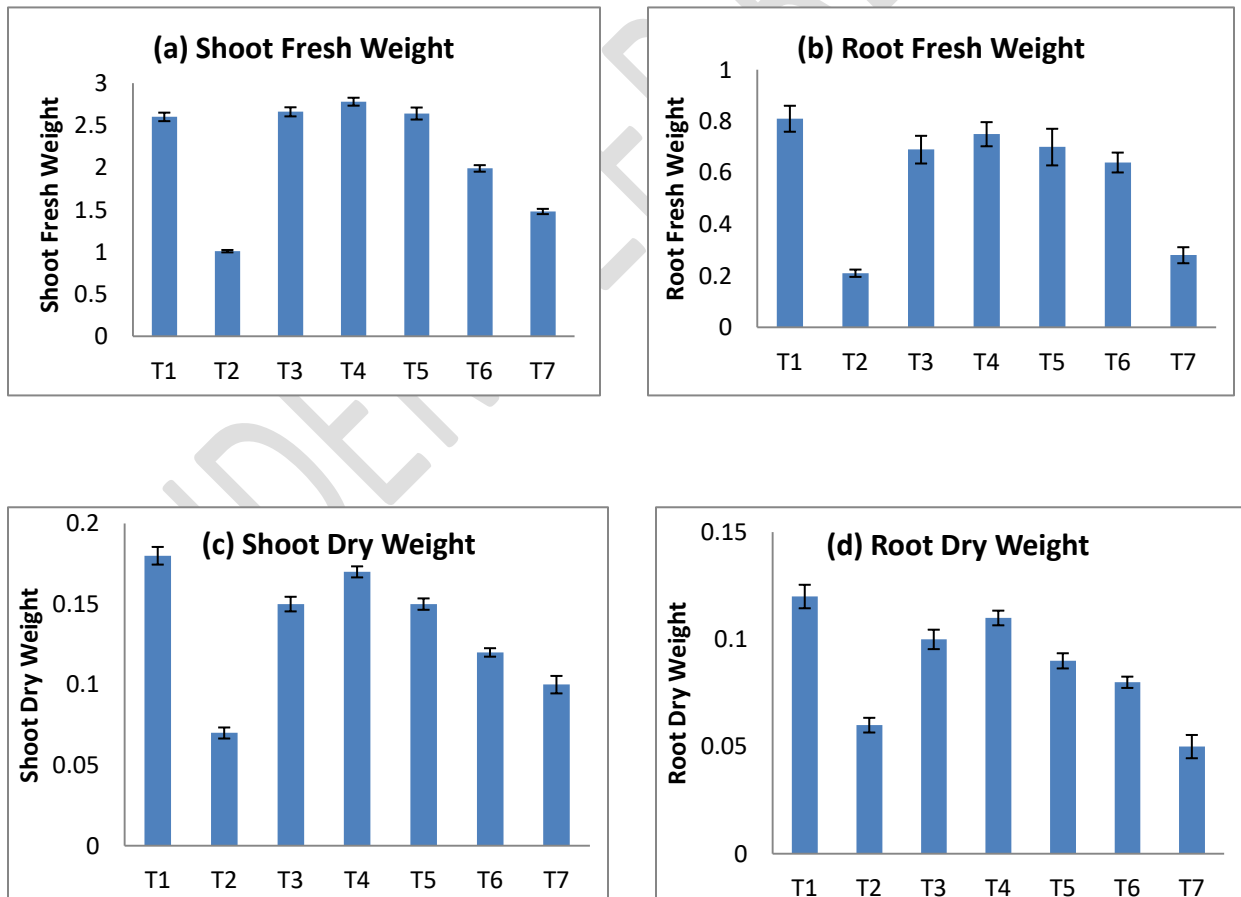


Figure 2. Effects of GA<sub>3</sub> priming and exogenous application on the germination and growth indices of maize under salt stress. (a) Shoot fresh weight; (b) Root fresh weight; (c) Shoot dry weight; (d) Root dry weight. The error bar represents standard error.

### 3.3 Exogenous GA<sub>3</sub> enhance RWC of plants under salt stress

The water status of maize plants was studied in this work by measuring RWC with and without salt stress using priming agents. The results showed that salt stress significantly reduced RWC by 10%.29 (Figure 3). The application of priming agents responded strongly to RWC at both times in comparison to salt conditions. The highest RWC increased by T4 treatment by 11%.

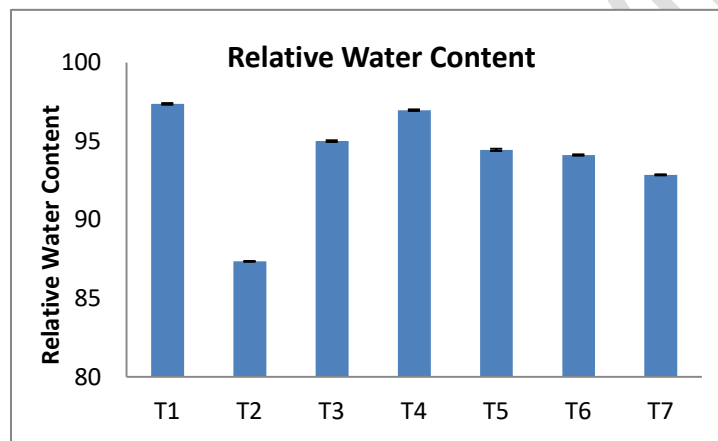


Figure 3. Effects of GA<sub>3</sub> priming and exogenous application on the relative water content of maize under salt stress. The error bar represents standard error.

### 3.4. Pretreatment of GA<sub>3</sub> regulate photosynthetic pigment of maize under salt stress

A significant fluctuation of Chl pigment contents was detected due to applied salt stress (Figure 4). A considerable decline in total Chl content (34.54%) including Chl a (29.42%) and Chl b (41.51%) in maize leaves due to salt stress compared to the control (Figure 4a–c). The supplementation of different concentrations of GA<sub>3</sub> remarkably augmented Chl a, Chl b, and total Chl contents. Pigment analysis also revealed that lycopene (52.26%), beta-carotene (43.93%) and carotenoids (31.10%) were also reduced due to salt stress, and the supplementation of GA<sub>3</sub> significantly increased the pigments (Figure 4c–f). The supplementation of 2mM GA<sub>3</sub> under salt stress increased the maximum lycopene (96.68%), beta-carotene (57.71%) and carotenoids (34.92%) contents.

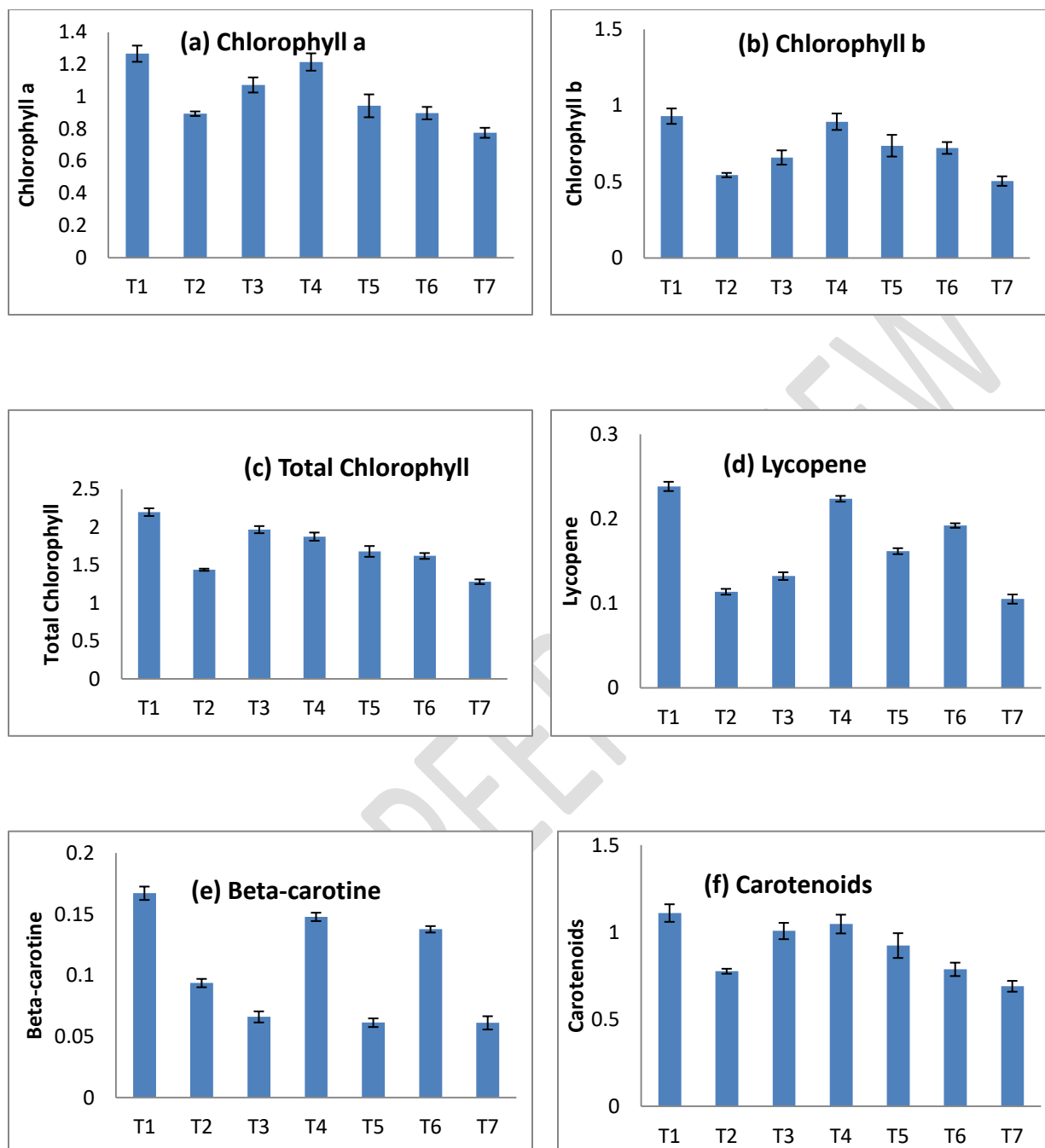


Figure 4. Effects of GA3 priming and exogenous application on the photosynthetic pigments of maize under salt stress. (a) Chlorophyll a; (b) chlorophyll a; (c) total chlorophyll; (d) lycopene; (e) beta-carotene; (f) carotenoids. The error bar represents the standard error.

#### 4. DISCUSSION

For seedling development and subsequent productivity, successful seed germination is the most important and fundamental stage in the plant growth cycle [34]. According to a number of

studies, seed priming is a widely used method for encouraging germination, boosting morphological traits, and speeding up plant development in both stress-free and stressful conditions [35, 36]. One of the main types of stress that significantly reduces seed germination and crop establishment is salt stress. According to the literature, many crops like wheat, faba beans, and rice can acquire salinity resistance through priming and the addition of various signaling molecules [37, 38, 39]. The priming of maize seeds with GA<sub>3</sub> improved germination and seedling characteristics under salt stress, as we recently observed [40]. The goal of the current study was to understand GA<sub>3</sub>'s function in the priming of maize seeds under salt stress. The findings showed that salt stress significantly reduced GP, SL, RL, and SVI (Figure 1). Additionally, salt stress decreased SFW, SDW, RFW, and RDW (Figure 2). The results showed that under salt stress, GA<sub>3</sub> priming increased the GP, GI, SL, RL, and SVI of maize. These findings are in line with other studies that discovered that a number of priming agents considerably lessened the detrimental effects of salt stress on parameters linked to seed germination in wheat [41], maize [42], and rice [43]. Plant growth may be facilitated by the exogenous input of GA<sub>3</sub>, which may increase its endogenous accumulation [44]. The higher seedling length and better growth of the GA<sub>3</sub>-treated maize in the current study (Figures 1 and 2) may be attributable to the increased stem and cell elongation because GA<sub>3</sub> is thought to be a key hormone for cell elongation [45, 46].

A crucial physiological step for preserving normal growth progression in plants under salt stress is maintaining an adequate water level [41]. Since RWC is a water-related property, it is well recognized as a water status indicator in plants [42]. Salinity lowers the soil's water potential, which has been associated with a decline in the RWC of leaves and a drop in photosynthesis [43]. Numerous plant groups' hydration status has been shown to be improved by exogenous chemical supplementation [47–49]. The study's findings revealed that RWC decreased as a result of the salt stress (Figure 3), and this was because the salt's damage to the leaves' cell walls altered their structure and reduced their ability to absorb water [50]. The outcomes also showed that the addition of these drugs and reduced GA<sub>3</sub> priming inhibit RWC when it is hindered by salt (Figure 3). This finding suggests that the uptake of extra water from the soil to modify the water level within plant organs may include a number of priming and exogenous substances.

Chlorophyll is the main pigment of plant photosynthesis and plays an important role in different The primary pigment used in photosynthesis in plants, chlorophyll, is crucial to numerous physiological processes in plants [51]. Salinity stress dramatically decreased the amount of leaf chlorophyll in this study (measured as Chla, Chlb, and total chlorophyll) compared to untreated plants (Figure 4). Additionally, it has been demonstrated in numerous earlier studies that salt stress can decrease the activity of photosynthetic pigments [52, 53]. The development of proteolytic enzymes at high salt concentrations is the cause of the decrease in chlorophyll content [45]. These enzymes also cause the degradation of chlorophyll [54] and the loss of photosynthesis in salty environments [55]. The decrease in chlorophyll levels (Figure 4) and the rate of photosynthesis [56] may possibly be contributing factors to the decrease in maize biomass

under salt stress (Figures 3). The chlorophyll content of maize leaves exposed to salt stress rose with the application of GA<sub>3</sub>, reaching its highest rise with the T4 treatment (Figure 4). In maize seedlings exposed to salinity stress, higher chlorophyll content accumulation may be related to decreased Na<sup>+</sup> buildup, reduced oxidative damage, and enhanced antioxidant defense. These results are in line with those of earlier research [57–59], which showed that the application of GA<sub>3</sub> raised the chlorophyll content of leaves. Under salt stress, foliar application of GA<sub>3</sub> considerably increased the chlorophyll content in maize [45].

## 5. CONCLUSIONS

According to the research, salt stress lowers the germination indices, growth features, leaf hydration status and photosynthetic pigments of maize. The germination %, seed vigor index, shoot and root length, shoot and root fresh and dry weight, leaf hydration status, and photosynthetic pigments of maize under salt stress are all improved by priming and exogenous application of GA<sub>3</sub>. Our findings also imply that maize production might be successful with lower GA<sub>3</sub> concentrations under salt stress condition. In spite of this, it is recommended that future research conducts a trial at the field level to validate our findings.

## REFERENCES

1. Jiang, C.; Zu, C.; Lu, D.; Zheng, Q.; Shen, J.; Wang, H.; Li, D. Effect of exogenous selenium supply on photosynthesis, Na<sup>+</sup> accumulation and antioxidative capacity of maize (*Zea mays* L.) under salinity stress. *Sci. Rep.* **2017**, *7*, 42039. [CrossRef] [PubMed]
2. Ahmad, P. Growth and antioxidant responses in mustard (*Brassica juncea* L.) plants subjected to combined effect of gibberellic acid and salinity. *Arch. Agron. Soil Sci.* **2010**, *56*, 575–588. [CrossRef]
3. Munns, R.; Tester, M. Mechanisms of salinity tolerance. *Annu. Rev. Plant Biol.* **2008**, *59*, 651–681. [CrossRef]
4. Gill, S.S.; Anjum, N.A.; Hasanuzzaman, M.; Gill, R.; Trivedi, D.K.; Ahmad, I.; Pereira, E.; Tuteja, N. Glutathione and glutathione reductase: A boon in disguise for plant abiotic stress defense operations. *Plant Physiol. Biochem.* **2013**, *70*, 204–212. [CrossRef]
5. Saade, S.; Maurer, A.; Shahid, M.; Oakey, H.; Schmöckel, S.M.; Negrão, S.; Pillen, K.; Tester, M. Yield-related salinity tolerance traits identified in a nested association mapping (NAM) population of wild barley. *Sci. Rep.* **2016**, *6*, 1–9. [CrossRef] [PubMed]
6. Anuradha, S.; Rao, S.S.R. Effect of brassinosteroids on salinity stress induced inhibition of seed germination and seedling growth of rice (*Oryza sativa* L.). *Plant Growth Regul.* **2001**, *33*, 151–153. [CrossRef]
7. Tuteja, N.; Sahoo, R.K.; Garg, B.; Tuteja, R. Os SUV 3 dual helicase functions in salinity stress tolerance by maintaining photosynthesis and antioxidant machinery in rice (*Oryza sativa* L. cv. IR 64). *Plant J.* **2013**, *76*, 115–127.
8. Deinlein, U.; Stephan, A.B.; Horie, T.; Luo, W.; Xu, G.; Schroeder, J.I. Plant salt-tolerance mechanisms. *Trends Plant Sci.* **2014**, *19*, 371–379. [CrossRef]

9. Pérez-López, U.; Robredo, A.; Lacuesta, M.; Sgherri, C.; Muñoz-Rueda, A.; Navari-Izzo, F.; Mena-Petite, A. The oxidative stress caused by salinity in two barley cultivars is mitigated by elevated CO<sub>2</sub>. *Physiol. Plant.* **2009**, *135*, 29–42. [CrossRef]
10. Suzuki, N.; Koussevitzky, S.; Mittler, R.; Miller, G. ROS and redox signalling in the response of plants to abiotic stress. *Plant Cell Environ.* **2012**, *35*, 259–270. [CrossRef]
11. Miller, G.; Suzuki, N.; Ciftci-Yilmaz, S.; Mittler, R. Reactive oxygen species homeostasis and signalling during drought and salinity stresses. *Plant Cell Environ.* **2010**, *33*, 453–467. [CrossRef]
12. Chaparzadeh, N.; D'Amico, M.L.; Khavari-Nejad, R.-A.; Izzo, R.; Navari-Izzo, F. Antioxidative responses of *Calendula officinalis* under salinity conditions. *Plant Physiol. Biochem.* **2004**, *42*, 695–701. [CrossRef]
13. Chawla, S.; Jain, S.; Jain, V. Salinity induced oxidative stress and antioxidant system in salt-tolerant and salt-sensitive cultivars of rice (*Oryza sativa* L.). *J. Plant Biochem. Biotechnol.* **2013**, *22*, 27–34. [CrossRef]
14. Parveen, A.; Liu, W.; Hussain, S.; Asghar, J.; Perveen, S.; Xiong, Y. Silicon Priming Regulates Morpho-Physiological Growth and Oxidative Metabolism in Maize under Drought Stress. *Plants* **2019**, *8*, 431. [CrossRef] [PubMed]
15. Rahman IU, Ali S, Alam M, Basir A, Adnan M, Malik MFA, Shah AS, Ibrahim M (2016) Effect of seed priming on germination performance and yield of okra (*Abelmoschus esculentus* L.). *Pakistan Journal of Agricultural Research* 29:250-259.
16. Vwioko ED (2013) Performance of soybean (*Glycine max* L.) in salt-treated soil environment following salicylic acid mitigation. *NISEB Journal* 13:44–49.
17. Jaleel, C.A., R. Gopi, P and R. Pannerselvam, 2009. Alterations in non-enzymatic antioxidant components of *Catharanthus roseus* exposed to paclobutrazol, gibberellic acid and *Pseudomonas fluorescens*. *Plant Omics Journal.*, 2(1): 30-40.
18. Akazawa, T., J. Yamaguchi and M. Hayashi, 1990. Rice-Amylase and gibberellin action a person view. In *gibberellin* (Eds), B.N. Takahashi, B.O. Phineey and J. Mac Millan. Springer-Varlag., pp: 114-124.
19. Swain, S.M. and D.P. Singh, 2005. Tale tales from slydwarves: novel functions of gibberellins in plant development. *Trends plant Sci.*, 10: 123-129.
20. Khassawneh, A.N.M., N.S. Karam, R.A. Shibli, 2006. Growth and following of black iris (*Iris nigricans* Dinsm). following treatment with plant growth regulators. *Sci. Horti.*, 107: 187-193.
21. Ismail, A.M.A., 1990. Germination ecophysiology in population of *Zygophyllum qatarenses* Hadidi from contrasting habitats. *J. Arid Environ.*, 18: 185-194.
22. Khan, M.A., B. Gul and D.J. Weber, 2002. Improving seed germination of *Salicornia rubra* (Chenopodiaceae) under saline conditions using germination regulating chemicals. *West. North Am. Nat.*, 62: 101-105.
23. Khan, M.A., B. Gul and D.J. Weber, 2004. Action of plant growth regulators and salinity on the seed germination of *Ceratoides lanata*. *Can. J. Bot.*, 82: 37-42.

24. Khan, M.A. and B. Gul, 2006. Halophyte seed germination. In: Eco-physiology of High Salinity Tolerant Plants. (Eds.): M.A. Khan and D.J. Weber, pp: 11-30. Springer Publication, Netherlands.
25. Kaur, S., A.K. Gupta and N. Kaur, 1998. Giberellin Aeverse the effect of salt stress in chickpea (*Cicer arietinum* L.) seedling by enhancing amylase activity and mobilization of starch in cotyledons, Plant Growth Reg., 26: 85-90.
26. Ashraf, M., F. Karim and E. Rasul, 2002. Interactive effects of gibberellic acid (GA3) and salt stress on growth, ion accumulation and photosynthetic capacity of two spring wheat (*Triticum aestivum* L.) cultivars differing in salt tolerance. Plant Growth Regul., 36(1): 49-59.
27. Wen, F., Z. Zhang, T. Bai, Q. Xu and Y. Pan, 2010. Proteomics reveals the effects of gibberellic acid (GA3) on salt-stressed rice (*Oryza sativa* L.) shoots. Plant Sci., 178: 170-175.
28. Starck, Z. and M. Kozinska, 1980. Effect of phytohormones on absorption and distribution of ions in the salt stressed bean plants. Acta Soc. Bot. Poloniae., 49: 111-125.
29. BeJaoui, M., 1985. Interaction between NaCl and some phytohormones on soybean growth. J. Plant Physiol., 120: 95-110.
30. Aloni, B. and E. Pressman, 1980. Interaction with salinity of GA3-induced leaf elongation, petiole pithiness and bolting in celery. Sci. Hortic., 13(2): 135-142.
31. Rauf F, Sagar A, Rahman T, Hossain MA, Kabir H, Hossain AK. Seed Quality Status of Rice Varieties Based on Physical Properties, Seed Health and Proximate Composition. Int. J. Plant Soil Sci. 2020;32:14-24.
32. Mostofa, M.G., Fujita, M. Salicylic acid alleviates copper toxicity in rice (*Oryza sativa* L.) seedlings by up-regulating antioxidative and glyoxalase systems. *Ecotoxicology* **22**, 959–973 (2013).
33. Lichtenthaler, H.K. Chlorophylls and carotenoids: Pigments of photosynthetic bio-membranes. Meth. Enzymol. 1987, 148, 350–382.
34. Shiade, S.R.; Boelt, B. Seed germination and seedling growth parameters in nine tall fescue varieties under salinity stress. ActaAgric. Scand.-B Soil Plant Sci. 2020, 17, 485–494.
35. Rhaman, M.S.; Imran, S.; Rauf, F.; Khatun, M.; Baskin, C.C.; Murata, Y.; Hasanuzzaman, M. Seed priming with phytohormones:An effective approach for the mitigation of abiotic stress. Plants 2021, 10, 37.
36. Muhei, S.H. Seed priming with phytohormones to improve germination under dormant and abiotic stress conditions. Adv. CropSci. Technol. 2018, 6, 403–409.
37. Abdel Latef, A.A.H.; Tahjib-Ul-Arif, M.; Rhaman, M.S. Exogenous auxin-mediated salt stress alleviation in faba bean (*Vicia faba*L.). Agronomy 2021, 11, 547.
38. Roy, P.R.; Tahjib-Ul-Arif, M.; Akter, T.; Ray, S.R.; Sayed, M.A. Exogenous ascorbic acid and hydrogen peroxide alleviates salt-induced oxidative stress in rice (*Oryza sativa* L.) by enhancing antioxidant enzyme activities and proline content. Adv. Environ.Biol. 2016, 10, 148–155.
39. Tania, S.S.; Rahaman, M.; Rauf, F.; Afroj Suborna, M.; Humayun Kabir, M.; Hoque, A.; Rhaman, M.S. Seed priming with SalicylicAcid (SA) and Hydrogen Peroxide (H2O2) Improve

Germination and Seedling Growth of Wheat (*Triticum aestivum* L.) under Salt Stress. *Asian J. Res. Crop Sci.* 2021, 6, 60–69.

40. Shahzad, K.; Hussain, S.; Arfan, M.; Hussain, S.; Waraich, E.A.; Zamir, S.; Saddique, M.; Rauf, A.; Kamal, K.Y.; Hano, C.; et al. Exogenously Applied Gibberellic Acid Enhances Growth and Salinity Stress Tolerance of Maize through Modulating the Morpho-Physiological, Biochemical and Molecular Attributes. *Biomolecules* **2021**, 11, 1005. <https://doi.org/10.3390/biom11071005>.

41. Farooq, M.; Basra, S.M.A.; Rehman, H.; Saleem, B.A. Seed priming enhances the performance of late sown wheat (*Triticum aestivum* L.) by improving chilling tolerance. *J. Agron. Crop. Sci.* 2008, 194, 55–60.

42. Ashraf, M.; Rauf, H. Inducing salt tolerance in maize (*Zea mays* L.) through seed priming with chloride salts: Growth and ion transport at early growth stages. *Acta Physiol. Plant.* 2001, 23, 407–414.

43. Tahjib-Ul-Arif, M.; Afrin, S.; Polash, M.A.S.; Akter, T.; Ray, S.R.; Hossain, M.; Hossain, M.A. Role of exogenous signaling molecules in alleviating salt-induced oxidative stress in rice (*Oryza sativa* L.): A comparative study. *Acta Physiol. Plant.* 2019, 41, 1–14.

44. Aldesuquy, H.; Ibrahim, A. Interactive effect of seawater and growth bioregulators on water relations, abscisic acid concentration and yield of wheat plants. *J. Agron. Crop Sci.* **2001**, 187, 185–193.

45. Tuna, A.L.; Kaya, C.; Dikilitas, M.; Higgs, D. The combined effects of gibberellic acid and salinity on some antioxidant enzyme activities, plant growth parameters and nutritional status in maize plants. *Environ. Exp. Bot.* **2008**, 62, 1–9.

46. Muhammad, I.; Kolla, M.; Volker, R.; Günter, N. Impact of nutrient seed priming on germination, seedling development, nutritional status and grain yield of maize. *J. Plant Nutr.* 2015, 38, 1803–1821.

47. Saboon, R.I.; Ahmad, N.; Ilyas, N.; Batool, N.; Gul, S. Salicylic acid enhances wheat plant growth under water stress conditions. *Int. J. Biol. Biotech.* 2015, 12, 329–336.

48. Tahjib-Ul-Arif, M.; Siddiqui, M.N.; Sohag, A.A.; Sakil, M.A.; Rahman, M.M.; Polash, M.A.; Mostofa, M.G.; Tran, L.S. Salicylic acid-mediated enhancement of photosynthesis attributes and antioxidant capacity contributes to yield improvement of maize plants under salt stress. *J. Plant Growth Regul.* 2018, 37, 1318–1330.

49. Hernández, J.A.; Díaz-Vivancos, P.; Acosta-Motos, J.R.; Barba-Espín, G. Potassium nitrate treatment is associated with modulation of seed water uptake, Antioxidative Metabolism and Phytohormone Levels of Pea Seedlings. *Seeds* 2022, 1, 5–15. [CrossRef]

50. Parvin, K.; Hasanuzzaman, M.; Bhuyan, M.H.; Nahar, K.; Mohsin, S.M.; Fujita, M. Comparative physiological and biochemical changes in tomato (*Solanum lycopersicum* L.) under salt stress and recovery: Role of antioxidant defense and glyoxalase systems. *Antioxidants* 2019, 8, 350.

51. Gu, X.; Li, Y.; Du, Y. Compensative impact of winter oilseed rape (*Brassica napus* L.) affected by water stress at re-greening stage under different nitrogen rates. *Chin. J. Eco Agric.* **2016**, *24*, 572–581.
52. El-Esawi, M.A.; Al-Ghamdi, A.A.; Ali, H.M.; Alayafi, A.A.; Witzczak, J.; Ahmad, M. Analysis of Genetic Variation and Enhancement of Salt Tolerance in French Pea (*Pisum Sativum* L.). *Int. J. Mol. Sci.* **2018**, *19*, 2433. [CrossRef]
53. Qu, C.; Liu, C.; Gong, X.; Li, C.; Hong, M.; Wang, L.; Hong, F. Impairment of maize seedling photosynthesis caused by a combination of potassium deficiency and salt stress. *Environ. Exp. Bot.* **2012**, *75*, 134–141. [CrossRef]
54. Sabater, B.; Rodriguez, M.T. Control of chlorophyll degradation in detached leaves of barley and oat through effect of kinetin on chlorophyllase levels. *Physiol. Plant.* **1978**, *43*, 274–276.
55. Yasseen, B. An Analysis of the Effects of Salinity on Leaf Growth in Mexican Wheats. Ph.D. Thesis, University of Leeds, Leeds, UK, 1983. *Biomolecules* **2021**, *11*, 1005 16 of 17
56. Xia, S.; Wang, X.; Su, G.; Shi, G. Effects of drought on cadmium accumulation in peanuts grown in a contaminated calcareous soil. *Environ. Sci. Pollut. Res.* **2015**, *22*, 18707–18717.
57. Bose, S.K.; Yadav, R.K.; Mishra, S.; Sangwan, R.S.; Singh, A.; Mishra, B.; Srivastava, A.; Sangwan, N.S. Effect of gibberellic acid and calliterpenone on plant growth attributes, trichomes, essential oil biosynthesis and pathway gene expression in differential manner in *Mentha arvensis* L. *Plant Physiol. Biochem.* **2013**, *66*, 150–158. [CrossRef]
58. Shah, S. Effects of salt stress on mustard as affected by gibberellic acid application. *Gen. Appl. Plant Physiol.* **2007**, *33*, 97–106.
59. Zang, Y.-X.; Chun, I.-J.; Zhang, L.-L.; Hong, S.-B.; Zheng, W.-W.; Xu, K. Effect of gibberellic acid application on plant growth attributes, return bloom, and fruit quality of rabbiteye blueberry. *Sci. Hortic.* **2016**, *200*, 13–18.