

Ulceroprotective potential of ethyl acetate fractions of *Persea americana* seed and *Bryophyllum pinnatum* leaf binary combinations in indomethacin induced gastric ulcer

ABSTRACT

Aims: This study was aimed at investigating the protective potential of ethyl acetate fractions of *Persea americana* (PA) seed and *Bryophyllum pinnatum* (BP) leaf binary combinations in indomethacin-induced gastric ulcers.

Methodology: Fifty (50) male Wistar rats were used in this study; they were assigned into 10 groups of five animals each and respective groups received a standard rat diet and drinking water *ad libitum*. The groups were the normal control group (NC), ulcer control (UC), Omeprazole, 20mgkg⁻¹ (OMEP), PA 400 mgKg⁻¹ body weight, and BP 400 mgKg⁻¹ body weight. Groups receiving binary combinations were (PA + BP, 1:1), (PA + BP, 1:2), (PA + BP, 1:3), (PA + BP, 2:1), (PA + BP, 3:1) each pre-treated with 400 mgKg⁻¹ body weight/day of respective mixture by intubation for 21 days. On the 22nd day after overnight fasting, a gastric ulcer was induced with indomethacin (30mg/kg body weight) by intubation in a single dose. Ulcer markers, and histopathology were measured using standard methods.

Results: The present study revealed that the ethyl acetate fraction of the plants PA seeds and BP leaf possess potent gastric tissue protective effects. Analysis of gastric changes indicated a significant (P<0.05) reduction in ulcer index, gastric acid output, gastric mucus content and pepsin activity of ulcerated rats pretreated with PA and BP fractions. The effect of the binary combinations was significantly (p<0.05) higher than single plant fractions. The protective effect of the fractions was in the order OMEP > PA+BP (3:1) > PA+BP (2:1) > PA+BP (1:1), > PA+BP (1:2) > PA+BP (1:3) > PA > BP. The observed ulceroprotective effect of the binary combinations of *P. americana* seed and *B. pinnatum* leaf may be attributed to the synergy of the phytochemicals contained in the fractions.

Conclusion:

These findings suggest that *P. americana* and *B. pinnatum* ethyl acetate fraction combination favorably protect against indomethacin-induced gastric mucosal oxidative damage, reducing ulcer index, gastric acid output, gastric mucus content and pepsin activity. The ethyl acetate fractions exhibited a dose-dependent inhibition of ulcer formation in indomethacin gastric ulcer model.

Keywords: *P. americana*, *B. pinnatum*, indomethacin, gastric ulcer, ulcer index

1. INTRODUCTION

Gastric/stomach ulcers (GU) and duodenal ulcers form the two most common types of peptic ulcers. Gastric and duodenal ulcers are common digestive tract diseases highly prevalent in every part of the world. They cut across age barriers and social class. The prevalence of peptic ulcer disease and gastric cancer in Africa has been reported [1]. These disorders involve inflammation and ulceration of the digestive tract; and occupy a place only secondary to carcinoma in the field of gastroenterology [2, 3]. "However, gastric ulcer is the most prevalent gastrointestinal disorder known, affecting more than 10% of people and accounting for an estimated 15,000 mortality yearly" [4]. "The annual global incidence of GU perforation and hemorrhage are estimated at 3.8–14.0 and 19.4–57.0 per 100,000 persons, respectively" [5]. "The statistics for gastric ulcer disease in other countries are variable and are hinged primarily on the major causes of the disease, *Helicobacter pylori*, and non-steroidal anti-inflammatory drugs (NSAID)" [6].

Several factors have also been implicated in the pathogenesis of gastric ulcerations including sedentary lifestyle, alcohol intake, spicy food, NSAID, and various bacterial infections [7, 8]. Factors such as interleukin-1 β (IL-1 β), matrix metalloproteinases (MMP), and tumor necrosis factor- α (TNF- α) [9, 10] and reactive oxygen species (ROS) and reactive nitrogen species (RNS) are strongly linked to gastric ulcer [11, 12]. Homeostatic changes such as the generation of nitric oxide (NO) from the endothelial nitric oxide synthase (e-NOS) and growth factors are important in ulcer healing. They promote angiogenesis, regulate gastric mucosal blood flow, and stimulate gastric mucus secretion [13].

29 There is no single effective management therapy for gastric ulcers, therefore a combination of treatment strategies are
30 applied depending on the severity, identified causative agent, and presenting symptoms [14]. "Current treatments have
31 brought about remarkable changes in gastric ulcer recovery, but the efficacy is still debatable; there are incidences of
32 relapses, adverse effects, and the danger of drug interactions during ulcer therapy. Hence, the search for an ideal anti-
33 ulcer drug continues and has also been extended to medicinal plants/herbs in search of new and novel molecules, which
34 may afford better protection and decrease the incidence of relapse. Plants have been identified as an important source of
35 phytoactive compounds with verifiable medicinal properties and protective potential against toxicity" [15].

36 "*Persea americana* Mill. and *Bryophyllum pinnatum* are plants widely known in the tropics for their food and medicinal
37 value. *P. americana* Mill. is an evergreen tree native to the Caribbean, commonly known as avocado and ube-beke,
38 orewépiá, and ganyen piya by Igbo, Yoruba, and Hausa tribes of Nigeria" [16]. "The reported medicinal properties of
39 different parts of *P. americana* include anti-inflammatory, antihypertensive, anti-ulcer, hypoglycaemic and
40 hypercholesterolaemic, and parasitic skin diseases treatment" [17, 18, 19]. "*B. pinnatum* is a member of the Crassulaceae
41 family, indigenous to Madagascar, but largely distributed with dominance in the rainforest belt and tropical countries. They
42 are known by names such as "air plant", "love plant", "miracle leaves", and "life plant", all attributable to their identified
43 characteristics" [20]. "*B. pinnatum* is known as 'odaa opue', and 'ewe abamoda' or 'odundun' among the Igbos and
44 Yorubas of South-eastern and western Nigeria respectively" [21]. "The Crassulaceae family has been widely investigated
45 due to their xeromorphic and medicinal properties. They are ethnopharmacologically applied in the management of
46 gastritis and bacterial infection and many diseases" [22, 23]. "Chemical composition, hepatoprotective activity, anti-
47 inflammatory effects, anti-ulcer activity, and free radical scavenging activities have also been reported" [24-27].

48 Furthermore, the seeds of *P. americana* and leaf of *B. pinnatum* have been widely touted to be wonder cures by
49 traditional practitioners; and applied locally in the management of gastric ulcers. It is therefore important to verify these
50 claims to create public awareness and ensure best practices. *P. americana* seed and *B. pinnatum* leaf may have been
51 studied for antidiabetic, hypotensive, and anti-inflammatory properties; but a comprehensive study on their ulcero-
52 protective effect using binary combinations has not been reported. It is necessary to study the combinatorial effect of
53 these plants which could enhance their effectiveness. The main objective of this study is to evaluate the ulcero-protective
54 properties of ethyl acetate fractions of the binary combination of *P. americana* seed and *B. pinnatum* leaf in indomethacin-
55 induced gastric ulcers.

57 2. MATERIAL AND METHODS

59 2.1 Chemicals/Reagents

60 Indomethacin (Sigma-Aldrich Mo USA), Ethanol (JHD, China), ethyl acetate (JHD, China), Sodium dihydrogen
61 orthophosphate (JHD, China), Disodium hydrogen orthophosphate (JHD, China), Omeprazole extended-release capsules
62 (Sanofi-aventis, Switzerland). All other chemicals and reagents used were of analytical grade.

63 2.2 Plant Materials

64 Fresh leaves of the plant *B. pinnatum* were collected from the bush around the Department of Biochemistry, Federal
65 University of Technology Owerri, Nigeria, while fresh fruits of *P. americana* were harvested from a farm in Ugiri-ike
66 Autonomous community, Ikeduru L.G.A. Imo State. The plant materials were authenticated by Prof. F. N. Mbagwu a plant
67 taxonomist at the Department of Plant Science and Biotechnology, Imo State University, Owerri, Nigeria. Plant specimens
68 were deposited in the institution herbarium with voucher numbers IMSUH 0225 and IMSUH 0226 respectively.

69 2.3 Experimental animals

70 Fifty (50) apparently healthy male Wistar rats (*Rattus norvegicus*) weighing 80-120g (averaging 8 weeks old) were used in
71 this study. They were purchased from the animal house of the Department of Veterinary Medicine, University of Nigeria,
72 Nsukka, and housed in stainless steel cages under standard laboratory conditions of light, temperature ($25\pm 2^{\circ}\text{C}$), and
73 relative humidity ($55 \pm 5\%$). The animals were fed standard rat pellets (Vital finisher, Nigeria) and portable water *ad*
74 *libitum*.

75 2.4 Preparation of ethyl acetate fraction

76 The fresh leaves of the plants were shed, sorted, and washed, while the fresh seeds were collected by carefully cutting
77 the fruits open; the seeds were peeled and cut into cubes for easy drying. Both samples were air-dried at 30°C , and
78 reduced to a coarse powder in a mill (Kenwood BL357). Each of the plant powders (500g) was extracted with 2 L of 80%
79 ethanol using a Soxhlet extractor. The ethanol extract was further partitioned between ethyl acetate and water to recover
80 the ethyl acetate soluble component of the extract. The ethyl acetate fraction was recovered by distillation under reduced

pressure at 49°C in a Buchi rota vapor (Switzerland), then dried to solid forms in vacuum desiccators, and stored in a freezer ($\leq 4.0^{\circ}\text{C}$) until when needed.

2.5 Experimental design

The ulcero-protective study of each ethyl acetate fraction was carried out in ten (10) groups of five (5) animals each grouped according to body weight (80-120mg/kgbw.). The groups and administration regimen were organized as follows:

Group 1: Group 1 served as the normal control group (NC). They received a standard rat diet and drank water *ad libitum* for 21 days. On the 22nd day, the animals were given indomethacin vehicle distilled water.

Group 2: Group 2 served as ulcer control (UC). Ulcer control group received a standard rat diet and drank water *ad libitum* for 21 days. On the 22nd day after overnight fasting, animals in this group were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

Group 3: Omeprazole standard group (OMEP) received a standard rat diet and drinking water *ad libitum* and pre-treatment with 20mg/kg body weight of Omeprazole by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

Group 4: *P. americana* group (PA) received a standard rat diet and drinking water *ad libitum* and pre-treatment with 400mg/kg body weight of *P. americana* ethyl acetate fraction by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with Indomethacin 30mg/kg body weight by intubation in a single dose.

Group 5: *B. pinnatum* group (BP) group received a standard rat diet and drinking water *ad libitum* and pre-treatment with 400mg/kg body weight of *B. pinnatum* ethyl acetate fraction by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

Group 6: *P. americana* + *B. pinnatum* group (PA + BP, 1:1). This group received a standard rat diet and drinking water *ad libitum* and pre-treatment with 400mg/kg body weight of *P. americana* + *B. pinnatum* ethyl acetate fraction (50%: 50% (1:1)) combination by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

Group 7: *P. americana* + *B. pinnatum* group (PA + BP, 1:2). This group received a standard rat diet and drinking water *ad libitum* and pre-treatment with 400mg/kg body weight of *P. americana* + *B. pinnatum* ethyl acetate fraction (33%: 67% (1:2)) combination by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

Group 8: *P. americana* + *B. pinnatum* group (PA + BP, 1:3). This group received a standard rat diet and drinking water *ad libitum* and pre-treatment with 400mg/kg body weight of *P. americana* + *B. pinnatum* ethyl acetate fraction (25%: 75% (1:3)) combination by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

Group 9: *P. americana* + *B. pinnatum* group (PA + BP, 2:1). This group received a standard rat diet and drinking water *ad libitum* and pre-treatment with 400mg/kg body weight of *P. americana* + *B. pinnatum* ethyl acetate fraction (67%: 33% (2:1)) combination by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

Group 10: *P. americana* + *B. pinnatum* group (PA + BP, 3:1). This group received a standard rat diet and drinking water *ad libitum* and pre-treatment with 400mg/kg body weight of *P. americana* + *B. pinnatum* ethyl acetate fraction (75%: 25% (3:1)) combination by intubation for 21 days. On the 22nd day after overnight fasting, they were treated with indomethacin (30mg/kg body weight) by intubation in a single dose.

2.6 Sacrificing, Isolation of the stomach, and collection of gastric juice

On the twenty-second day (4 hrs post ulcer induction), the animals were humanely sacrificed under light anesthesia with dichloromethane. The animal stomach was carefully excised; thereafter opened along the greater curvature and gastric content was drained into a centrifuge tube. Ten milliliters of distilled water was added and the resultant solution was

126 centrifuged at 3000 rpm for 10 min. The supernatant obtained was used for biochemical analyses. The cleaned stomach
127 tissues were preserved in ice-cold 0.1M phosphate buffer saline (1:4 (w/v), pH 7.4) before macroscopic examination.

128 **2.7 Determination of ulcer index**

129 The ulcer index was determined using the method described by Goyal [28]. The excised rat stomach tissues were
130 carefully opened along the greater curvature. Debris was cleaned with a running stream of water. The stomach tissue was
131 then stretched out on cardboards, with the luminal surface facing up. The ulcer index was calculated from the glandular
132 portion of the stomach, with the aid of a magnifying glass and measuring tape.

133 The ulcer index was calculated as:

$$134 \text{ Ulcer index} = \frac{10}{x}$$

$$135 \text{ Where } x = \frac{\text{Total mucosal surface}}{\text{Total ulcerated area}}$$

136 **Measurement rules:** ulcer lesions were measured along the greatest length. Five identified petechial were considered
137 equivalent to 1 sq-mm of ulcer area. The total area of the glandular portion of the stomach and that of ulcerated mucosa
138 were measured and used for the determination of the ulcer index [28].

139 **2.8 Determination of free acidity, total acidity, and gastric pH**

140 Free and total acidity were determined in the gastric content according to the method described by Kulkarni and Varghese
141 [29]. The gastric content was collected, and made up to 10 ml with sterile deionized water. This was centrifuged and the
142 supernatant was taken in a conical flask and titrated with 0.01N NaOH to the yellow endpoint using 100 µl Topfer's
143 reagent as indicator. Furthermore, 100 µl phenolphthalein was added and titration continued till the phenolphthalein
144 endpoint was reached. The amount of 0.01N NaOH required to titrate to the yellow end point was used to calculate the
145 free acidity; the titre value for the use of both Topfer's reagent and phenolphthalein was used to determine the total
146 acidity. The acidity was calculated by the following formula and expressed in mmol/L [29]. The pH of gastric juice was
147 determined using a pH meter.

$$\text{Acidity} = \frac{\text{Volume of NaOH} \times \text{Normality} \times 100 \text{ mmol/L}}{0.1}$$

148 **2.9 Determination of gastric mucus content**

149 The determination of gastric mucus content was carried out according to the method described by Corne *et al.* [30]. The
150 excised stomach tissues were soaked in 0.05M sodium acetate buffered 0.1% alcian blue solution (pH 5.0). Excess
151 uncomplexed dye was removed by washing with 0.025M sucrose and subsequently tissue was soaked in 0.1M MgCl₂.
152 The blue solution obtained was extracted in ether and optical density was measured at 605nm. The mucin content of the
153 sample was determined from the standard curve of alcian blue which was expressed in µg alcian blue/g tissue [30].

154 **2.10 Determination of pepsin activity**

155 Pepsin activity was determined according to the method of Debnath *et al.* [31] and Lowry *et al.* [32]. Briefly, about 1ml of
156 diluted gastric juice was mixed with 0.5ml 2% hemoglobin solution in 0.06 M hydrochloric acid; and incubated for 20 mins
157 at room temperature. The set up was precipitated with 1ml of 0.5M trichloroacetic acid, and centrifuged at 4000rpm for
158 10mins. Then 1.0ml of the supernatant was mixed with 1.0ml of alkaline copper sulfate solution and 0.5ml of dilute Folin-
159 Denis reagent and incubated for 30 min at room temperature. The absorbance of the samples was determined by
160 spectrophotometry at 610 nm. The gastric pepsin activity was expressed in µmol/L.

161 **2.11 Determination of total nitrite contents in gastric mucosal tissues**

162 Total nitrite content was determined in the homogenates as described by Green *et al.* [33]. A 1ml portion of the
163 supernatant was mixed with 1ml of Griess reagent (consisting 0.5ml portion of 0.1% N-(1-naphthyl) ethylenediamine
164 dihydrochloride (NED) and 0.5ml of 1% sulfanilamide in 2.5% phosphoric acid). The mixture was incubated for 10 min at
165 room temperature in the dark, and the absorbance was measured at 540 nm. The concentration of nitrite in sample was
166 determined using a standard calibration curve and expressed in mmol/L.

167 **2.12 Histological studies of stomach tissues**

168 The method described by Okoro [34] was used with minor modifications.

169 2.13 Statistical Analysis

170 The study was carried out using a complete randomized design. Results of data generated were analyzed using analysis
171 of variance with a statistical package for social sciences (version 25). Statistical significance of values was considered at
172 $p < 0.05$ using the turkey and Duncan homogeneity of variance test. Results are presented as mean \pm standard deviation.

173 3. RESULTS AND DISCUSSION

176 3.1 Effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on gastric mucosa 177 protection in rats

178 Figure 1a shows the effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on ulcer index
179 in indomethacin-induced gastric ulcers in male albino rats. Result obtained from the study (Figure 1a) shows that

180 indomethacin administration significantly ($p < 0.05$) increased the ulcer index when compared to the standard drug
181 (omeprazole). The ethyl acetate fractions of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction and their
182 combinations resulted in a varying degree of significant ($p < 0.05$) reduction in ulcer index. The ulcer index of animals that
183 received the ethyl acetate fraction combinations of PA+BP (1:1), PA+BP(1:2), PA+BP (1:3), PA+BP (2:1), and PA+BP
184 (3:1) were significantly ($p < 0.05$) reduced ulcer index when compared to those of groups receiving PA and BP. However,
185 the ethyl acetate fraction combinations of PA+BP (2:1) and PA+BP (3:1) effectively reduced ulcer index but were not
186 normalized to the same level as control or the standard drug omeprazole.

187 Figure 1b shows the percentage ulcer protective effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction
188 administration in indomethacin-induced gastric ulcers in male albino rats. Result obtained from the study (Figure 1b)
189 shows that the ethyl acetate fractions of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction and their
190 combinations administration had a significant ($p < 0.05$) protective effect against induction of gastric ulcer in rats. The
191 percentage ulcero-protection was 2.68 ± 0.91 , 89.53 ± 3.27 , 25.17 ± 4.62 , 12.97 ± 1.57 , 47.52 ± 7.49 , 45.63 ± 7.08 ,
192 35.33 ± 6.07 , 65.56 ± 8.82 and 74.75 ± 5.38 % for the groups UC, OMEP, PA, BP, PA+BP (1:1), PA+BP (1:2), PA+BP
193 (1:3), PA+BP (2:1) and PA+BP (3:1) respectively. The ulcero-protective effect of the fractions binary combinations was
194 significantly ($p < 0.05$) higher when compared to the single plant fractions, but was lower than the effect of the omeprazole
195 standard. The protective effect of the fractions was in the order Omeprazole > PA+BP (3:1) > PA+BP (2:1) > PA+BP (1:1),
196 > PA+BP (1:2) > PA+BP (1:3) > PA > BP.

197 3.2 Effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on free acidity and 198 total acidity in indomethacin-induced gastric ulcer in rats.

199 Figure 1c show the effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on free acidity in
200 indomethacin-induced gastric ulcers in the male albino rat. Results obtained from the study show that induction with
201 indomethacin caused a significant ($p < 0.05$) increase in gastric acidity when compared to normal control animals.
202 However, the administration of the ethyl acetate fractions of PA, BP, and the combinations protected against acid reflux in
203 the gastric mucosa, reducing free acidity. But these fractions did not normalize stomach-free acidity except for the PA+BP
204 (3:1) group. The animals receiving PA+BP (3:1) at the dosage of 400mg/Kgb.wt were able to normalize free acidity
205 comparable to omeprazole standard and normal control.

206 Figure 1d show the effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on total acidity
207 in indomethacin-induced gastric ulcers in the male albino rat. Results obtained from the study show that indomethacin
208 caused a significant ($p < 0.05$) increase in total acidity when compared to normal control animals. The administration of *P.*
209 *americana* seed and *B. pinnatum* leaf fractions resulted in a reduction of total gastric acidity except for the group that
210 received BP (400mg/Kgb.wt). Furthermore, total acidity-reducing effects were more effective in groups PA+BP (2:1) and
211 PA+BP (3:1) respectively. Among these groups, total acidity did not significantly ($p < 0.05$) differ from the normal control
212 and standard group.

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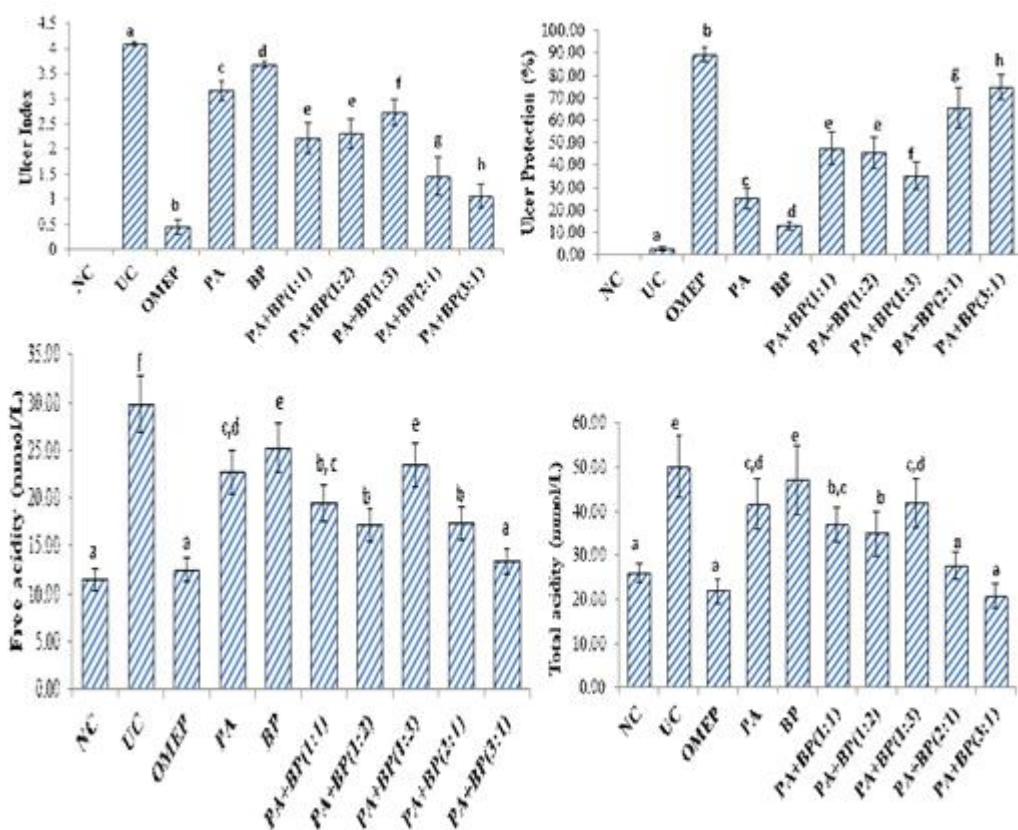


Figure 1: Effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction binary combinations administration on ulcer index, ulcer protection, free acidity, and total acidity in indomethacin-induced gastric ulcer in male albino rats. Bars are mean \pm Standard of 5 determinations. Bars bearing different superscript letters across groups are significantly different ($p < 0.05$).

3.3 Effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on Gastric mucus content and pepsin activity in indomethacin-induced gastric ulcer in rats.

Figure 2a show the effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on gastric mucus content in indomethacin-induced gastric ulcer in the male albino rat. The results show that indomethacin administration significantly ($p < 0.05$) depleted gastric mucus content in the UC group when compared to the normal control. Administration of the ethyl acetate fractions in the combinations PA+BP (1:1), PA+BP (1:3), PA+BP (2:1), and PA+BP (3:1) showed a significant ($p < 0.05$) protection of gastric mucus secretion which were comparable to OMEP and normal control.

Figure 2b show the effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on pepsin activity in indomethacin-induced gastric ulcers in male albino rats. The results presented in Figure 2b show that indomethacin administration promoted significant ($p < 0.05$) release of pepsin into the stomach. The ethyl acetate fractions did not normalize pepsin activity to levels found in control, however, showed a significant reduction of pepsin secretion in animals administered the different ethyl acetate fraction and their combinations.

3.4 Effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on nitrite concentration and Gastric pH in indomethacin-induced gastric ulcer in rats.

Figure 2c show the effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on nitrite concentration of stomach homogenates in indomethacin-induced gastric ulcers in male albino rats. Stomach homogenate nitrite concentration was significantly ($p < 0.05$) elevated in the ulcer control, but administration of the ethyl acetate fractions in the groups OMEP, PA+BP(1:1), PA+BP(1:2), PA+BP(1:3), PA+BP(2:1) and PA+BP(3:1) significantly normalized stomach nitrite concentration. However, PA+BP (3:1) administration was the most effective.

Figure 2d show the effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction administration on gastric pH in indomethacin-induced gastric ulcers in the male albino rat. The result obtained from the study indicated an increased gastric pH value in animals receiving indomethacin. However, the increases in gastric pH were not significantly ($p < 0.05$) different from that of the normal control group.

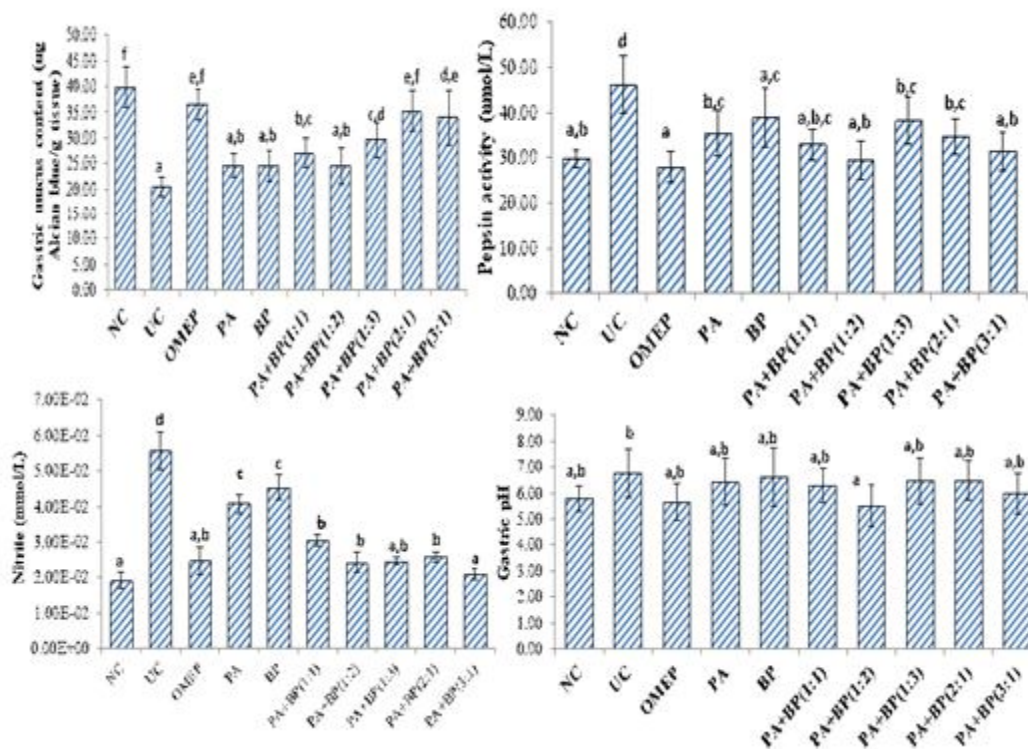


Figure 2 Effect of *P. americana* seed and *B. pinnatum* leaf ethyl acetate fraction binary combinations administration on gastric mucus content, pepsin activity, nitrite concentration, and gastric pH in indomethacin-induced gastric ulcer in male albino rats. Bars are mean \pm Standard of 5 determinations. Bars bearing different superscript letters across groups are significantly different ($p < 0.05$).

3.5 Histological examinations of stomach sections

Histological sections (x100) of the animal stomach tissues including normal control, Ulcer control, Standard group (Omeprazole), *P. americana* (PA) 400mg/Kg bwt, *B. pinnatum* (BP) 400mg/Kg bwt, (PA + BP) (1:1) 400mg/Kg bwt, (PA + BP) (1:2) 400mg/Kg bwt, (PA + BP) (1:3) 400mg/Kg bwt., (PA + BP) (2:1) 400mg/Kg bwt and (PA + BP) (3:1) 400mg/Kg bwt. are shown in plates 1-10.

The first photomicrograph (Plate 1) shows a section of the stomach from group1 (Control) showing organized gastric epithelium or mucosa and gastric pit. Normal gastrointestinal tissue lined by both stratified squamous and columnar epithelium. The mucosa, submucosa, muscularis externa and serosa are normal.

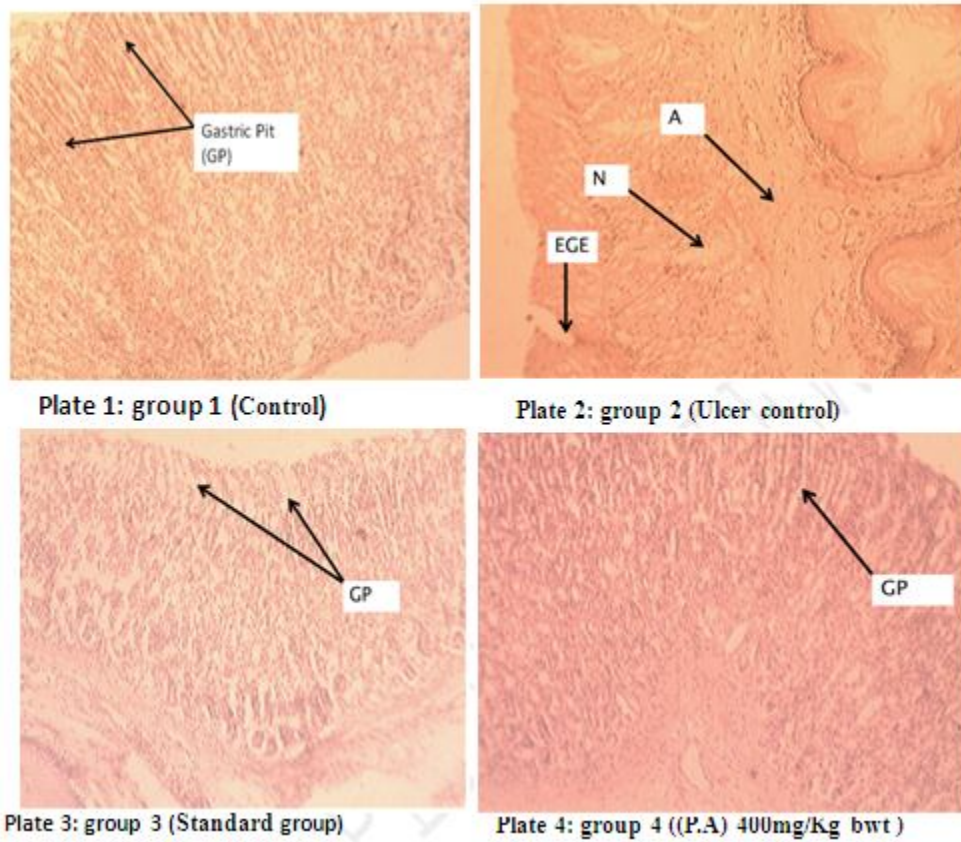
The second photomicrograph (plate 2) represent the ulcer control group. The plate show tha the mucosa, submucosa, muscularis interna and externa are not normal; there is presence of eroded gastric epithelium (EGE), necrosis (N) and atrophy (A). However, in the group that was pre-treated with 20mg/kg body weight of omeprazole and later received Indomethacin 30mg/kg, organized gastric epithelium or mucosa and gastric pit was observed (Plate 3). The normal gastrointestinal tissue lined by both stratified squamous and columnar epithelium. The mucosa, submucosa, muscularis externa and serosa are normal.

Furthermore, the group that was pre-treated with *P. americana* (PA) 400mg/Kg bwt and later received Indomethacin 30mg/kg, (Plate 4), the plate shows organized gastric epithelium or mucosa and gastric pit. Normal gastrointestinal tissues lined by both stratified squamous and columnar epithelium were observed. The mucosa, submucosa, muscularis externa and serosa are normal. Plate 5 shows the group *B. pinnatum* (BP) 400mg/Kg bwt, they was erosion of gastric epithelium (EGE) or mucosa (M); normal submucosa (SM) and muscularis interna (showing a haemorrhagic streak) is evident.

The sixth photomicrograph (plate 6) represent the group that was pre-treated with (PA + BP) (1:1) 400mg/Kg bwt and later received indomethacin 30mg/kg. The plate show organized gastric epithelium or mucosa and gastric pit as well as normal gastrointestinal tissue lined by both stratified squamous and columnar epithelium. The mucosa, submucosa, muscularis externa and serosa are normal and are better than the group pre-treated with PA or BP only and later given Indomethacin 30mg/kg. Plate 7 show the group that was pre-treated with (PA + BP) (1:2) 400mg/Kg bwt and later received Indomethacin 30mg/kg. The plate show organized gastric epithelium or mucosa and gastric pit as well as normal

276 gastrointestinal tissue lined by both stratified squamous and columnar epithelium. There is presence of necrosis (N) and
277 atrophy (A) in the submucosa and muscularis layers. In the group that was pre-treated with (P.A + B.P) (1:3) 400mg/Kg
278 bwt and later received Indomethacin 30mg/kg, (Plate 8), the plate show presence of gastric epithelium or mucosa and
279 gastric pit with some parts eroded.

280 Plate 9 show the group that was pre-treated with (P.A + B.P) (2:1) 400mg/Kg bwt and later received indomethacin
281 30mg/kg, (Plate 9), the Plate shows presence of eroded gastric epithelium with normal submucosa and submuscularis.
282 Also, plate 10 show the group that was pre-treated with (P.A + B.P) (3:1) 400mg/Kg bwt and later received Indomethacin
283 30mg/kg the plate show normal architecture of gastric epithelium or mucosa, and gastric musculature with better
284 presentations than those in groups 6-9.



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286 **Photomicrograph of section of the gastric tissues**

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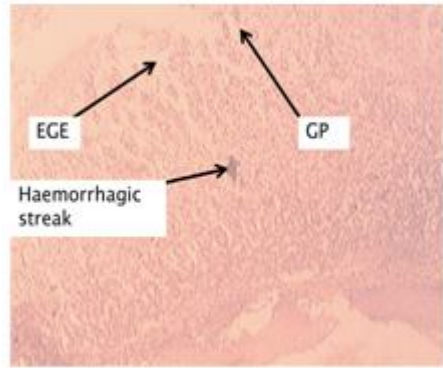


Plate 5: group 5 (BP) 400mg/Kg bwt)

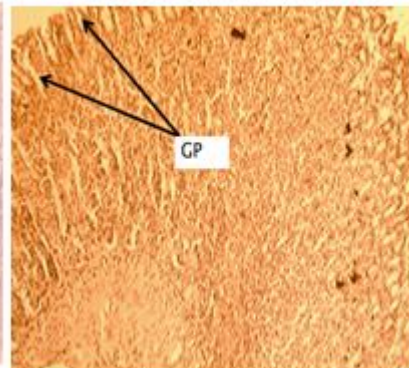


Plate 6: group 6 (PA + BP) (1:1) 400mg/Kg bwt)

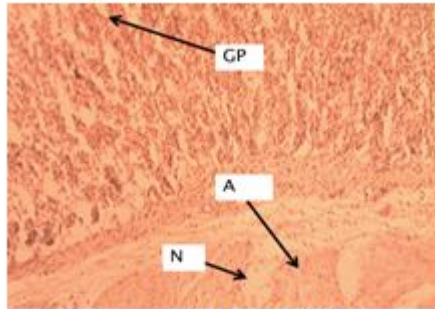


Plate 7: group 7 (PA + BP) (1:2) 400mg/Kg

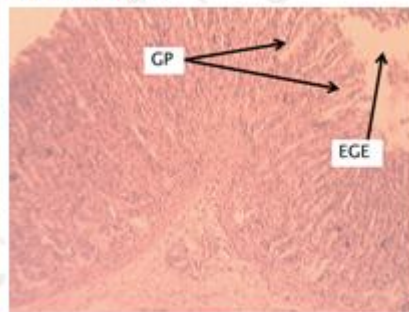


Plate 8 group 8 (PA + BP) (1:3) 400mg/Kg bwt)

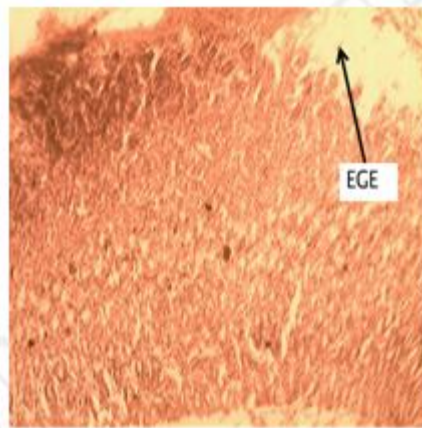


Plate 9: group 9 (P . A + B.P) (2:1) 400mg/Kg bwt).

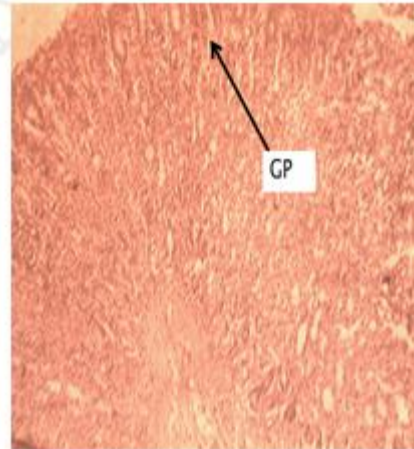


Plate 10: group 10 (P.A + B.P) (3:1) 400mg/Kg bwt)

Photomicrograph of section of the gastric tissues

3.6 Discussion

The present study assessed the ulcero-protective potential of ethyl acetate fractions of *P. americana* seed and *B. pinnatum* leaf binary combinations in indomethacin-induced gastric ulcers. Investigation of the protective effect of *P. americana* seed, *B. pinnatum* leaf, and their binary combinations against gastric ulcers showed that indomethacin administration resulted in significant changes in gastric ulcer indicators. The ethyl acetate fractions of *P. americana* seed and *B. pinnatum* leaf and their combinations provided varying degree of ulceroprotection. The ethyl acetate fraction combinations of PA+BP (1:1), PA+BP (1:2), PA+BP (1:3), PA+BP (2:1), and PA+BP (3:1) significantly reduced ulcer index when compared to groups that received PA and BP only. However, the ethyl acetate fraction combinations of PA+BP (2:1) and PA+BP (3:1) effectively reduced ulcer index but were not normalized to level found in control or the standard drug treated animals.

301 Analysis of the percentage ulceroprotection shows that the ethyl acetate fractions of *P. americana* seed and *B. pinnatum*
302 leaf binary combination administration had a significant protective effect against the induction of gastric ulcers in rats. The
303 protective effect of these fractions relative to the single plant fractions was in the order PA+BP (3:1) > PA+BP (2:1) >
304 PA+BP (1:1) > PA+BP(1:2) > PA+BP(1:3) > PA > BP. The protective effect of the binary combinations was observed to
305 increase with the increasing ratio of *P. americana* in the combination. The PA+BP (2:1) and PA+BP (3:1) binary
306 combination caused a 2.5-fold and 3-fold increase in percentage protection compared to PA or BP alone. Previous studies
307 have reported the gastroprotective properties of the ethyl acetate fractions of *P. americana* leaf and seed fractions [35-
308 37]. Brena *et al.* [36], reported that avocado seeds may prevent gastric mucosal injury and oxidative stress induced by
309 indomethacin. Also, the works of Sharma *et al.* [38], Amadi *et al.* [39]; De Araújo *et al.* [40] reported gastric protecting
310 property of different solvent extracts of *B. pinnatum*. In our study, the ulceroprotective effect of the binary combinations
311 was significantly higher when compared to the single plant fractions. The higher protective activity of the binary
312 combinations containing higher ratios of *P. americana* seed fraction may be attributable to a the reported rich polyphenolic
313 blue-print of this fraction [35].

314 Also, analysis of other gastroprotection markers showed that administration of indomethacin caused a significant rise in
315 gastric free and total acidity. However, the administration of *P. americana* seed, *B. pinnatum* leaf, and binary
316 combinations significantly protected against gastric acid reflux. The potency of the protective effect was found to be
317 similar to the trends seen for ulceration. Results demonstrated a dose-dependent effect on gastric acidity which
318 progressively increased with the ratio of *P. americana* seed ethyl acetate fraction in the binary combinations of the two
319 plants. Administration of the combinations PA+BP (1:1), PA+BP (1:2), PA+BP (2:1), and PA+BP (3:1) to indomethacin-
320 exposed animals effectively resulted in a reduction of gastric acidity, but only PA+BP (2:1) and PA+BP (3:1) produced a
321 normalization of gastric acidity similar to those seen for animals receiving Omeprazole and normal control animals. The
322 result obtained for gastric pH indicated a slight elevation of gastric pH values in animals receiving indomethacin. However,
323 the changes were not significantly different from the normal control group. Administration of *P. americana* seed and *B.*
324 *pinnatum* leaf ethyl acetate fraction and the different combinations in this study maintained gastric acidity preventing
325 gastric acid concentration increase. In addition, indomethacin administration significantly depleted gastric mucus content
326 in the UC group when compared to the normal control. Administration of the ethyl acetate fractions in the combinations
327 PA+BP (1:1), PA+BP (1:3), PA+BP (2:1), and PA+BP (3:1) showed significant protection on gastric mucus secretion
328 which was comparable to OMEP and normal control. Also, stomach homogenates nitrite concentration was significantly
329 elevated in the ulcer control, but administration of the ethyl acetate fractions in the groups receiving binary combinations,
330 significantly normalized stomach nitrite concentration, with the group PA+BP (3:1) being more effective.

331 Furthermore, indomethacin induction promoted the significant release of pepsin into the stomach. The ethyl acetate
332 fractions did not normalize pepsin activity to the concentration found in control; they showed a significant effect on pepsin
333 secretion in animals administered the different ethyl acetate fractions and their combinations. These observations are
334 confirmed by microscopic evidence obtained in our histopathological analysis. In histological observation, the stomach of
335 control animals showed no damage (Plate 1). However, in the ulcer control group (Plate 2), exposure to indomethacin
336 presented damage to gastric tissue at a microscopic level. Histopathological injury caused by indomethacin administration
337 is characterized by erosion of gastric epithelium, necrosis, and cell atrophy (Plate 2). Also, the mucosa, submucosa,
338 muscularis interna and externa were not normal.

339 However, administration of the *P. americana* and *B. pinnatum* ethyl acetate fractions combinations was able to protect
340 against damage caused by indomethacin to a varying degree. The ethyl acetate fraction combination PA + BP (3:1)
341 400mg/Kg bwt, was able to effectively protect the histological structure of the gastric mucosa, preventing distortion and
342 mucosal damage. This was similar to that observed for normal animals and those treated with omeprazole standard. The
343 findings of the histopathological analysis corroborate our biochemical findings on the gastro protective effect and the
344 reduction in gastric acid secretion by the combinations of ethyl acetate fraction of *P. americana* and *B. pinnatum* at the
345 different binary ratios.

346 Non-steroidal anti-inflammatory drugs such as indomethacin produce anti-inflammatory effects by inhibiting
347 cyclooxygenase enzymes, this in turn, suppresses the formation of prostaglandins and thromboxane from arachidonic
348 acid [41]. This suppressive effect leads to gastric lesions due to the reduction of cytoprotection of prostaglandins in the
349 gastric mucosa [42] or via the production of oxygen radicals [43]. The gastroprotective mechanism of these fractions may
350 be attributed to several possible mechanisms which though speculative may bring insight into their effect. The
351 homeostasis of mucosal integrity is achieved by a number of mucosal protective factors among which are secretion and
352 action of mucus and bicarbonate [44, 45]. Prostaglandins (E2 and I2) are known to stimulate secretion of mucus and
353 bicarbonate, maintain mucosal integrity as well promote mucosal regeneration [44, 46]; as well promote mucosal
354 regeneration. From the result of our study, it can be seen that the fractions and their combinations inhibited indomethacin-
355 induced ulceration of the mucosa. The ethyl acetate fractions of the extracts may be acting by preventing the inhibitory
356 effect of indomethacin on prostaglandin synthesis.

357 Furthermore, the gastro-protective effect and the reduction in the gastric acid secretion of the ethyl acetate fraction may
358 also be attributed to the active component of the ethyl acetate fraction polyphenols, alkaloids, and flavonoids which have
359 been proven to be useful for digestive disorders and disturbances of the gastrointestinal tract [47, 48]. NSAIDs such as
360 indomethacin produce anti-inflammatory effects by suppressing of formation of prostaglandins and several other
361 mechanisms which result in the production of oxygen radicals. Therefore, oxidative stress has been strongly suggested to
362 be a dominant presence in the progression of indomethacin-induced gastric ulcers [49, 50].
363

364 4. CONCLUSION

365 The outcome of this study revealed that oral pre-administration of these ethyl acetate fractions before the exposure of the
366 gastric mucosa to indomethacin resulted in significant protection from ulceration. These findings suggest that *P.*
367 *americana* and *B. pinnatum* ethyl acetate fraction combination favorably protect against indomethacin-induced gastric
368 mucosal oxidative damage, reducing ulcer index, gastric acid output, gastric mucus content and pepsin activity. The ethyl
369 acetate fractions exhibited a dose-dependent inhibition of ulcer formation in indomethacin gastric ulcer model.
370

371 ETHICAL APPROVAL

372 The animal rights and ethical committee of the Federal University of Technology Owerri approved the study before the
373 commencement of the study. All treatment of the animals was in accordance with the Principles of Laboratory Animal
374 Care (NIH Publication, 1985 to 1993; revised, 1985).
375

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379

380 COMPETING INTERESTS

381 The authors declare that they have no competing interests.
382

383 AUTHORS' CONTRIBUTIONS

384 There was a total collaboration between the authors in the execution of this study. CUI, ESA, VAO, KMEI, and CSA
385 designed the study and wrote the protocol, while CUI, ESA, COU, and NGE performed the statistical analysis and
386 interpretation of study data. Authors ESA and NGE did the literature searches, while ESA wrote the first draft of the
387 manuscript and incorporated all corrections from co-authors. CUI, VAO KMEI, and CSA critically revised the manuscript
388 for intellectual content. All authors read and approved the final manuscript.
389

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510

511 LIST OF ABBREVIATIONS

512 **GU:** Gastric ulcer; **PA:** *Persea americana*; **BP:** *Bryophyllum pinnatum*; **NC:** Normal control; **UC:** Ulcer control; **OMEPR:**
513 Omeprazole; **NSAIDS:** Non-steroidal anti-inflammatory drugs.

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