

## Original Research Article

### **Relationship between photosynthetic capacity and carbohydrate content of *Mangifera indica* cv. Harumanis in response to girdling and paclobutrazol**

#### **ABSTRACT**

Girdling and paclobutrazol have been related with the effects on the photosynthetic capacity and carbohydrate content of plants. The physiological changes caused by these methods distress the growth and development of plants in general. A field experiment was carried out from August 2021 to June 2022 on 5 years of open-field *Mangifera indica* cv. Harumanis trees grown at Malaysian Agriculture Research and Development Institute (MARDI), Serdang, Selangor. The objective of this experiment was to understand the relationship between girdling, paclobutrazol application, combined methods and untreated trees on plant photosynthetic performances and carbohydrate content in the leaves. The treatments were performed on 1<sup>st</sup> December 2021, and the measurements of leaf gas exchanges and carbohydrate content were performed 4<sup>th</sup> weeks later on fully expanded leaves shoot, experiencing similar light exposure. In the study, the combination of girdling and paclobutrazol application resulted in a significant decrease in photosynthetic rate ( $P_n$ ), stomatal conductance ( $g_s$ ) and transpiration rate ( $T_r$ ) but significantly increase in intercellular  $CO_2$  concentration ( $C_i$ ) and carbohydrate content.

#### **KEYWORDS**

Carbohydrate content, Girdling, *Mangifera indica* cv. Harumanis, Paclobutrazol,  
Photosynthetic capacity

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## INTRODUCTION

Harumanis are one of the best mangoes in Malaysia, and they are primarily grown in the Northern Peninsular Malaysian states of Perlis and Kedah (Nasron et al. 2020; Sani et al. 2018). Harumanis mango is only available once a year during the season and is only found north of the homeland due to its sensitivity to climate conditions that necessitates a clear and consistent dry season (Shaidatul Azdawiyah et al.. 2020) that provides prolonged hot weather for 2 to 3 months (Uda et al.. 2020). Harumanis have a distinct flavour unlike any other mango. They have a slightly creamy and milky taste and a very strong and distinct aroma, making it the most popular and receiving a steady increase in demand year after year (Muhamad Hafiz et al. 2019; Nur Afiqah 2015; Rosidah et al. 2010).

Harumanis production is frequently associated with physiological changes in the plant caused by endogenous factors such as hormones, genetic composition, carbohydrates, and stress conditions, as well as the age of the buds (Cho et al. 2017). Furthermore, environmental factors, primarily temperature change (Halder and Bandyopadhyay 2021), as well as management practises such as plant growth regulator use (Malshe et al. 2020), pruning (Hahn et al. 2022), and irrigation (Venkata Subbaiah et al. 2017), are frequently associated with the changes. Endogenous and environmental factors are beyond our control; thus, under management practise, girdling (Prates et al. 2021) and paclobutrazol (Kumar et al. 2020) application are quite common for fruit producers, and have also shown great results in improving

crop growth, yield, and quality @ in increasing the productivity and quality of several agricultural crops (Azizi et al. 2022; Ran et al. 2022; Shivashankara et al. 2019).

Girdling is the removal of a thin strip of bark tissue encircling a trunk or branch, obstructing the downward translocation of photosynthates and metabolites through the phloem (Bottcher et al. 2018; Isogimi et al. 2014). Its most immediate effect is to halt assimilate movement, resulting in an accumulation of carbohydrates produced by photosynthesis along the phloem (Gawankar et al. 2019). The accumulation of carbohydrate reserves in the leaves reduces photosynthetic capacity (Zepeda et al. 2022; Asao and Ryan 2015). Girdling has a significant impact on photosynthesis because it reduces stomatal conductance (William et al. 2000) which affecting the rate of gas exchange and transpiration rate (Lopez et al. 2015) resulting in less leaf transpiration and thus a decrease in photosynthesis rate.

Paclobutrazol functions as a chemical growth and development stimulator by altering associated physiological processes (Abdalla et al. 2021). The use of this plant growth inhibitor on soil inhibits the biosynthesis of gibberellin, which is responsible for regulating growth and development (Desta and Amare 2021). Previous research has shown that PBZ application alters crop plant photosynthetic capacity (Zhao et al. 2022; Nyan et al. 2017; Roseli et al. 2012) and increases carbohydrate production in leaf (Xu et al. 2020; Upreti et al. 2014). Because gibberellins are involved in the biogenesis of chloroplasts, paclobutrazol alters photosynthetic potential and, as a

result, plant gas exchange (Rodrigues et al. 2016). Paclobutrazol also has an effect on the efficiency of carbon use in leaves, which is determined by the balance of photosynthesis and respiration (Christiaens 2014).

Therefore, the objective of this experiment was to investigate the relationship between photosynthetic capacity and carbohydrate content of *Mangifera indica* cv. Harumanis in response to 4 methods; (i) girdling, (ii) paclobutrazol, (iii) combined methods including, and (iv) untreated tree. We hypothesized that combination of girdling and paclobutrazol would decrease photosynthetic capacity inversely proportional to carbohydrate content of *Mangifera indica* cv. Harumanis.

## **MATERIALS AND METHODS**

### **Planting material**

This study was conducted at MARDI, Serdang, Selangor, from August 2021 to June 2022, using five years of open-field *Mangifera indica* cv. Harumanis trees grown in sedentary soil and separated by 4 × 4 m. Thirty-six healthy trees of nearly uniform shape and size are subjected to the same fertilisation programme and other agricultural practises chosen for this study. T1- No induction (Control); T2- girdling at primary branches; T3- soil drenching at 4 mL/L paclobutrazol; and T4- girdling at primary branches + soil drenching at 4 mL/L paclobutrazol were applied to selected trees. Girdling was accomplished by removing a 10 mm wide ring of bark above 30 cm from the base of all primary branches. The commercial product paclobutrazol (25% active ingredient) was used. The treatments were carried out simultaneously

on December 1, 2021, at the same morphological size of the tree that had been subjected to similar light exposure.

### **Photosynthetic**

A portable photosynthesis system was used to measure photosynthetic rate ( $P_n$ ), intercellular  $CO_2$  concentration ( $C_i$ ), stomatal conductance ( $g_s$ ), and transpiration rate ( $T_r$ ) (LI-6400XT; LI-COR, Lincoln, NE, USA). The measurements were taken on the fifth fully expanded leaves from the plant apex on the fourth week after treatment between 8:00 and 10:00 a.m.

### **Determination of Total Carbohydrate Content**

The anthrone method was used to determine the total carbohydrate content (Hedge and Hofreiter 1962). A 100 mg sample of leaves was placed in a boiling tube. It was hydrolyzed for 3 hours in a boiling water bath with 5 mL 2.5 N hydrochloric acid. It was allowed to cool to room temperature after 3 hours. The solution was then neutralised with solid sodium carbonate until the effervescence was stopped. The solution was diluted with distilled water until it reached 100 ml before being centrifuged for 10 minutes. The 0.5 and 1 ml supernatant were then collected, and an aliquot was taken for analysis. Meanwhile, the standard was made by taking 0 ml, 0.2 ml, 0.4 ml, 0.6 ml, 0.8 ml, and 1.0 ml of the working standard and leaving '0' as a blank. By adding distilled water to all of the tubes, including the sample tubes, the volume was increased to 1 ml. The tubes were then filled with 4 ml of anthrone reagent and heated in a boiling water bath for 8 minutes. After that, the solution was

rapidly cooled before being placed in a UV spectrophotometer (Thermo Fisher Scientific Orion AquaMate 7000) and the light absorption at 630 nm was measured. To determine the concentration of glucose in each test sample, a standard graph was created by plotting the concentration of the standard glucose on the X-axis versus absorbance on the Y-axis. Equation 1 was used to calculate the amount of carbohydrate present in the sample tube based on the graph:

$$\text{Amount of carbohydrate present in 100 mg of the sample} = (\text{mg of glucose} \div \text{Volume of test sample}) \times 100$$

### **Statistical Analysis**

The experiment was laid out in randomized complete block design (RCBD). Each treatment was replicated three times and each replicate was represented by three trees. The data obtained was analysed using ANOVA in SAS software (Version 9.4, SAS Institute Inc. Cary, North Carolina, USA) and differences between treatments means were compared using Duncan Multiple Range Test Difference (DMRT) at  $P \leq 0.05\%$ .

### **RESULTS AND DISCUSSION**

The treatment reduced the photosynthetic capacity of the *Mangifera indica* cv. Harumanis' plants while increasing carbohydrate content compared to the control (Figure 1). The combination method of girdling and paclobutrazol application

resulted in the lowest net photosynthesis rate of  $2.71 \mu\text{mol CO}_2 \text{ m}^2/\text{s}^1$ . This combination also reduced stomatal conductance, with the lowest result being  $0.07 \mu\text{molH}_2\text{O}/\text{m}^2/\text{s}$ . Furthermore, the combination method yielded the lowest transpiration rate of  $1.66 \mu\text{molH}_2\text{O}/\text{m}^2/\text{s}$ . However, the intercellular  $\text{CO}_2$  concentration of this combined method was significantly higher than the other treatments, at  $312.48 \mu\text{mol CO}_2 \text{ mol}$ . The combination method of girdling and paclobutrazol application increased total carbohydrate content. The highest carbohydrate content was found to be  $2.62267 \text{ mg/g}$ .

This finding is partially agrees with previous reports on mango (Sarker and Rahim 2018). It is well known that paclobutrazol interferes with the hormone abscisic acid's production (Opio et al., 2020). A stress hormone that causes stomatal closure is one of abscisic acid's most crucial roles (Bharath et al. 2021). Based on the degree of stomatal aperture, stomatal conductance calculates the rate of gas exchange through the leaf stomata. A plant may close its leaf stomata under stress in an effort to preserve water and minimise water loss. But in doing so, the plant also limits how much  $\text{CO}_2$  it can absorb. In plants,  $\text{CO}_2$  is an essential component of the process of photosynthesis. Therefore, a decrease in  $\text{CO}_2$  restricts photosynthetic. Furthermore, in addition to this treatment with the girdled branches, which increase in a general decrease in this common adaptation response of plants to the onset of artificial stress by stem girdling conditions, which followed a trend similar to photosynthetic activity (Agurla et al. 2018). Similar to the present study, the removal



of the sink demand of roots typically results in decreased stomatal conductance and transpiration, and accumulation of abscisic acid in leaves (Tang et al. 2016), which affects carbon status and leads to a decrease in photosynthetic results (Urban and Alphonsout 2007).

The findings of this study, which are consistent with earlier reports, strongly suggested that girdling and paclobutrazol can increase the production of carbohydrates in the leaf (Sousa et al. 2022; Wu et al. 2018; Hua et al. 2014). The early termination of growth caused by the action of paclobutrazol as a gibberellin biosynthesis inhibitor is known to reduce vegetative growth rate, which leads to the buildup of carbohydrates (Ashraf and Ashraf 2020; Wieland and Wample 1985). Carbohydrates serve as a main source of energy for growth and differentiation of plant organs during flowering and fruit development (Haziş et al. 2018). Girdling alters the distribution of carbohydrates in the body by preventing assimilates from passing through the phloem. The buildup and depletion of carbohydrates above and below the girdling zone, respectively, cause a number of changes in growth and development (Chai et al. 2021). In addition, when the vegetative sink decreased, the transfer of carbohydrates to the shoot increased (Quentin et al. 2013).

## **CONCLUSION**

In conclusion, girdling and paclobutrazol together dramatically lower stomatal conductance and transpiration, which affect net photosynthesis rate, while

simultaneously raising the carbohydrate content in the leaves of *Mangifera indica* cv. Harumanis.

#### **CONFLICT OF INTEREST STATEMENT**

All authors contributed to the (a) conception and design, (b) writing of the article or its critical revision for essential intellectual content, and (c) approval of the final version. This paper has not been sent to another journal or other publishing outlet, and neither is it being reviewed there.

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## REFERENCES

Abdalla N, Taha, N, Bayoumi Y, Ramady HE, Syalaby TA. Paclobutrazol applications in agriculture, plant tissue cultures and its potential as stress ameliorant: A mini review. *Environment, Biodiversity and Soil Security*. 2021; 5(1):245 - 257.

Asao S, Ryan MG. Carbohydrate regulation of photosynthesis and respiration from branch girdling in four species of wet tropical rain forest trees. *Tree Physiology*. 2015; 35:608–620.

Agurla S, Gahir S, Munemasa S, Murata Y, Raghavendra AS. Mechanism of stomatal closure in plants exposed to drought and cold stress. *Advances in Experimental and Biology*. 2018;1081:215-232.

Ashraf N, Ashraf, M. Response of Growth Inhibitor Paclobutrazol in Fruit Crops. (Prunus, IntechOpen: Ed. Kuden A, Ali A) 2020:92883.

Azizi S, Rana VS, Sharma S, Chauhan J, Kumar V. Effect of girdling on yield and quality of kiwifruit (*Actinidia deliciosa Chev.*) raised through different propagation methods. *Indian Journal of Ecology*. 2022;49(2):496-501.

Bharath P, Gahir S, Raghavendran AS. Abscisic acid induced stomatal closure: An important component of plant defense against abiotic and biotic stress. *Frontier of Plant Science*. 2021;12(1):615114.

Bottcher C, Boss PK, Harvey KE, Burbidge CA, Davies C. Peduncle-girdling of Shiraz (*Vitis vinifera* L.) bunches and sugar concentration at the time of girdling

affect wine volatile compounds. *Australian Journal of Grape and Wine Research*. 2018;24:206–218.

Chai L, Li Q, Wang H, Wang C, Xu J, Yu H, Jiang W. Girdling alters carbohydrate allocation to increase fruit size and advance harvest in tomato production. *Scientia Horticulturae*. 2021;276:109675.

Christiaens A. Factors Affecting Flower Development and Quality in *Rhododendron simsii*. [PhD Thesis]. Ghent University, Ghent, Belgium; 2014.

Cho LH, Yoon J, An G. The control of flowering time by environmental factors. *The Plant Journal*. 2017;90(4):708-719.

Desta B, Amare G. Paclobutrazol as a plant growth regulator. *Chemical and Biological Technologies in Agriculture*. 2021; 8(1).

Gawankar MS, Haldankar PM, Salvi BR, Parulekar YR, Dalvi NV, Kulkarni MM, Saitwal YS, Nalage NA. Effect of Girdling on Induction of Flowering and Quality of Fruits in Horticultural Crops: A review. *Advanced Agricultural Research and Technology Journal*. 2019;3(2):201-215.

Hahn F, Valle S, Navarro-Gomez C. Pruning and water saving management effects on mango high-density and mature orchards. *Agronomy*. 2022;12:2623.

<https://doi.org/10.3390/agronomy12112623>

Halder B, Bandyopadhyay J. Evaluating the impact of climate change on urban environment using geospatial technologies in the planning area of Bilaspur, India. *Environmental Challenges*. 2021;5:100286.

azis NH, Aznan AA, Jaafar MN, Azizan FA, Ruslan R, Rukunudin IH. 2018. Assessment of carbohydrate contents in Perlis Harumanis mango leaves during vegetative and productive growth. *Proceeding of the International Conference on Advanced Manufacturing and Industry Applications; 2018 August 15-17; Malaysia. Sarawak; p. 012025.*

Hedge JE, Hofreiter BT. Determination of total carbohydrates by anthrone method, In: Whistler RL, Be Miller JN, editor. *Carbohydrate chemistry* 17. New York: Academic Press; 1962.

Hua S, Zhang Y, Yu H, Lin B, Di H, Zhang D, Ren Y, Fhang, Z. Paclobutrazol application effects on plant height, seed yield and carbohydrate metabolism in canola. *International Journal of Agriculture and Biology*. 214;16(3):471-479.

Isogimi T, Matsushita M, Nakagawa, M. Species-specific sprouting pattern in two dioecious *Lindera* shrubs: The role of physiological integration. *Flora*. 2014;209:718–724.

Kumar A, Ram S, Bis L, Singh C. Paclobutrazol boost up for fruit production: A review. *International Journal of Energy and Environmental Science*. 2020;1(1):019-031.

Lopez R, Brossa R, Gil L, Pita P. Stem girdling evidences a trade-off between cambial activity and sprouting and dramatically reduces plant transpiration due to feedback inhibition of photosynthesis and hormone signaling. *Frontiers in Plant Science*. 2015;6:285.

Malshe KV, Haldankar PM, Patil SS. Effect of foliar application of plant growth regulators on seasonal variation in physiological behavior in mango cv. Alphonso. *International Journal of Current Microbiology and Applied Sciences*. 2020;9(6):1629-1642. <https://doi.org/10.20546/ijcmas.2020.906.201>

Muhamad Hafiz MS, Hartinee A, Dalila N, Zul Helmey MS, Razali M, Siti Aisyah A, Wan Mohd Reza WI, Shaidatul Azdawiyah AT. Effect of multilocation production on the reproductive growth, yield, and quality of Harumanis mango. *International Journal of Current Advanced Research*. 2019;8(04):18175-18180.

Nasron N, Ghazali NS, Shahidin NM, Mohamad AA, Pugi SA, Razi NM., 2020. Soil suitability assessment for Harumanis mango cultivation in UiTM Arau, Perlis. *Proceeding of the Advanced Geospatial and Surveying Conference; 2020 October 7;Malaysia. Perlis; p. 012007.*

Nur Afiqah A. Growth, physiological and biochemical responses as affected by paclobutrazol for flowering induction on water induced stress mango plants (*Mangifera indica* L. cv. Harumanis). [Masters thesis]. Universiti Putra Malaysia; 2015.

Nyan TM, Yahya A, Izham A, Ranj SN. Gas exchange, growth and flowering of *lagerstroemia indica* treated with different concentration and application techniques of paclobutrazol. *Asian Journal of Plant Sciences*. 2017;16(1):37-44.

Opio P, Tomiyama H, Saiyo T, Ohkawa K, Ohara H, Kondo S. Paclobutrazol elevates auxin and abscisic acid, reduces gibberellins and zeatin and modulates their transporter genes in Marubakaido apple (*Malus prunifolia Borkh.* var. ringo Asami) rootstocks. *Plant Physiology and Biochemistry*. 2020;155(1):502-511.

Prates AR, Zuge PGU, Leonel S, Souza JMA, Avila JD. Flowering induction in mango tree: updates, perspectives and options for organic agriculture. *Pesquisa Agropecuaria Tropical*. 2021;51. <https://doi.org/10.1590/1983-40632021v5168175>

Quentin A, Close D, Hennen L, Pinkard, E. Down-regulation of photosynthesis following girdling, but contrasting effects on fruit set and retention, in two sweet cherry cultivars. *Plant Physiology and Biochemistry*. 2013;73:359-367.

Ran, J, Guo W, Hu C, Wang X, Li P. Adverse effects of long-term continuous girdling of jujube tree on the quality of jujube fruit and tree health. *Agriculture*. 2022;12(7):922.

Rodrigues LCA, Castro EM, Pereira FJ, Malileque IF, Barbosa JPRAD, Rosado SCS. Effects of paclobutrazol on leaf anatomy and gas exchange of *Toona ciliata* clones. *Australian Forestry*. 2016;79(4):241-247.

Roseli ANM, Ying TF, Ramlan MF. Morphological and physiological response of *Syzygium myrtifolium* (Roxb.) walp. to paclobutrazol. *Sains Malaysiana*. 2012;41(10):1187–1192.

Rosidah, M, Faridah H, Jamaliah MY, Norzaidi MD. Examining market accessibility of Malaysia's Harumanis mango in Japan: Challenges and potentials. *Business Strategy Series*. 2010;11(1):3-12.

Sani M.A, Abbas H, JaafarMN, Bahagia MAG., 2018. Morphological Characterisation of Harumanis mango (*Mangifera indica* Linn.) in Malaysia. *International Journal of Environmental and Agriculture Research*. 2018;4(1):45.

Sarker BC, Rahim MA. Influence of paclobutrazol on growth, yield and quality of mango. *Bangladesh Journal of Agricultural Research*. 2018;43(1).

Shaidatul Azdawiyah AT, Muhamad HafizMH, Mohd Aziz R, Zul Helmey MS, Muhammad Zamir AR, Wan Mahfuzah WI, Mohammad Hariz AR, Mohd Ghazali R, Syarol Nizam AB, Mohd Alif OM. Effects of environmental temperature and precipitation pattern on growth stages of *Mangifera indica* cv. Harumanis mango. *Journal of Agricultural Science*. 2020;12(12).

Shivashankara KS, Geetha GA, Roy TK. Influence of girdling on flower sex ratio, biochemical constituents, and fruit set intensity in mango (*Mangifera indica* L.). *Biologia Plantarum*. 2019;63:432-439.



Sousa K, Lopes PRC, Cavalcante I, Filho JC, Silva LDS, Pereira ECV, Silva JTL., 2022. Impact of paclobutrazol on gibberellin-like substances and soluble carbohydrates in pear trees grown in tropical semiarid. *Revista de la Facultad de Ciencias Agrarias*. 2022;54(1):46-56.

Tang GL, Li XY, Zeng FJ., 2016. Different causes of photosynthetic decline and water status in different stages of girdling in *Alhagi sparsifolia* Shap. (Fabaceae). *Brazilian Journal of Botany*. 2016;39(2):19-529.

Uda MNA, Subash C, Gopinath B, Hashim U, Asyraf H, Afnan MN, Aminudin A, Bakar MAA, Sulaiman MK, Parmin NA., 2020. Harumanis mango: Perspectives in disease management and advancement using interdigitated electrodes (IDE) nano-biosensor. *Proceeding of the 2nd Joint Conference on Green Engineering Technology and Applied Computing*; 2020 February 4-5; Thailand. Bangkok. P. 864.

Upreti KK, Prasad SRS, Reddy YTN, Rajeshwara AN. Paclobutrazol induced changes in carbohydrates and some associated enzymes during floral initiation in mango (*Mangifera indica* L.) cv. Totapuri. *Indian Journal of Plant Physiology*. 2014;19(4):317-323.

Urban L, Alphonsout L. Girdling decreases photosynthetic electron fluxes and induces sustained photoprotection in mango leaves. *Tree Physiology*. 2007;7:354-352.

Xu LJ, Liu HX, Wu J, Xu CY. Paclobutrazol improves leaf carbon-use efficiency by increasing mesophyll conductance rate, while abscisic acid antagonizes this increased rate. *Photosynthetica*. 2020;58(3):762-768.

Venkata Subbaiah K, Reddy NN, Reddy MLN. Effect of paclobutrazol and other chemicals on yield and quality of mango cv. Banganpalli. *International Journal of Science, Environment and Technology*. 2017;6(3):1809-1819.

Wieland WF, Wample RL. Effects of paclobutrazol on growth, photosynthesis and carbohydrate content of 'delicious' apples. *Scientia Horticulturae*. 1985;26:139-147.

Wu Y, Sun M, Zhang J, Zhang L, Ren Z, Min R, Wang X, Xia Y. Differential effects of paclobutrazol on the bulblet growth of oriental lily cultured in vitro: Growth behavior, carbohydrate metabolism, and antioxidant capacity. *Journal of Plant Growth Regulation*. 2019;38(2):359-72.

Zepeda AC, Heuvelink E, Marcelis LFM. Non-structural carbohydrate dynamics and growth in tomato plants grown at fluctuating light and temperature. *Frontiers in Plant Science*. 2022;3:968881.

Zhao I, Zhao J., Lai H, Chen BC, B, Mengjie ZM, Zhao YL, Liu Y, Xiangdong L, Li X, Yang DYD. Effects of paclobutrazol application on plant architecture, lodging resistance, photosynthetic characteristics, and peanut yield at different single-seed precise sowing densities. *The Crop Journal*. 2022. <https://doi.org/10.1016/j.cj.2022.05.012>

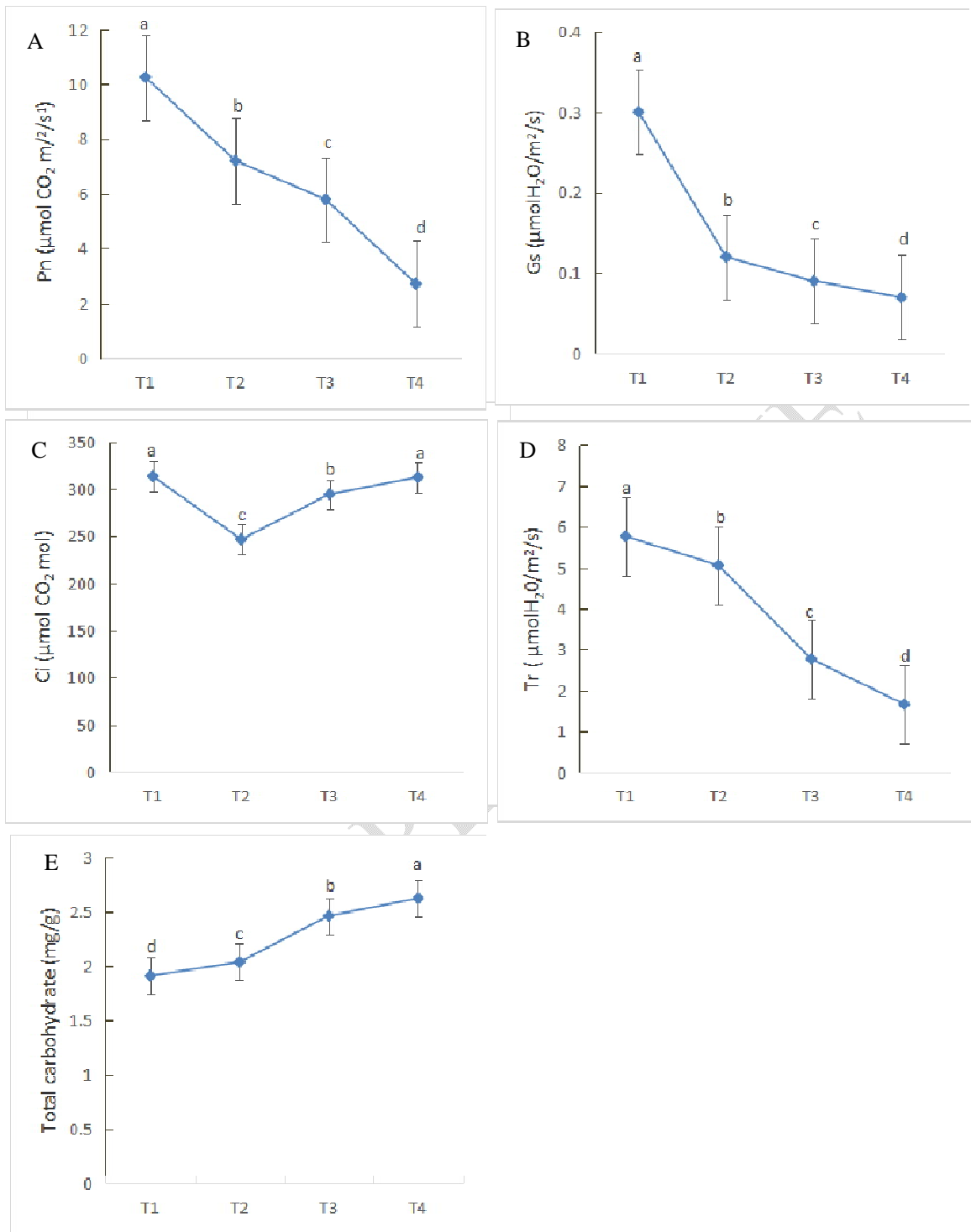


Figure 1: Net photosynthesis rate (A), stomatal conductance, intercellular CO<sub>2</sub> levels (C), transpiration rate (D) and total carbohydrate (E) at T1- No induction (Control); T2- girdling at primary branches; T3- soil drenching at 4 mL/L PBZ and T4- girdling at primary branches + soil drenching at 4 mL/L PBZ. Bars are mean for standard error. Different letters above bars indicate significant differences using Duncan Multiple Range Test Difference (DMRT) at P ≤ 0.05