

UV/visible Spectroscopic Studies and Analytical Evaluation of *DiclipteraVerticillate* Leaves Extracts as Eco-friendly Indicator for Acid-Base Titration

Abstract

Synthetic indicator usually employed in acid-base titration to determine the end point has the tendency of polluting the environment since it is non-biodegradable. Hence there is need to develop an eco-friendly acid-base indicator from plant origin to serve as efficient alternative to replace the synthetic indicators for acid-base titration. This research work focused on the development of easily available, cost effective and eco-friendly indicator from *Diclipteraverticillate* leaves extracts for acid-base titration. UV/visible spectroscopic analysis was used to determine the wavelength of maximum absorption (λ_{max}) and the absorbance of ethanol extract (EE), ethanol extract in basic medium (EEB), ethanol extract in acidic medium (EEA), hot water extract (HWE), Hot water extract in basic medium (HWEB) and hot water extract in acidic medium (HWEA) in order to determine the chromophoric changes of the plant extract phytochemicals before and after titration. Titrimetric analysis was carried out in reactions of strong acid/strong base (HCl/NaOH), strong acid/weak base (HCl/NH₄OH), weak acid/strong base (CH₃COOH /NaOH) and weak acid/weak base (CH₃COOH/NH₄OH). The results revealed that both extracts exhibit hypsochromic shift (shift to a shorter wavelength) with corresponding hypochromic effect (decreased in the absorption intensity) in both acidic and basic media. The titrimetric results revealed that the plant extracts gave the end point comparable with that of the synthetic indicators such as methyl orange and phenolphthalein.

Keywords

Uv/visible spectroscopy, Natural indicator, *DiclipteraVerticillate*, Acid-base Titration

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Introduction

Acid-base indicators are chemicals used to determine whether an aqueous solution is acidic, neutral, or alkaline. This is done by observing the change in colour of the solution when indicator is added. It is used to possibly determine the unknown concentration of acid or base through titrimetric analysis. When there is any change in the colour of analyte solution due to indicator, the titration is complete and the final volume of titrant is noted and further calculations are made to determine the concentration of the analyte (Nwosu *et al.*, 2014). Hence, indicators are used to determine the end point of any titrations. Examples of synthetic acid-base indicators include, phenolphthalein, methyl red and methyl orange etc.

The search for natural compounds as acid-base indicators started increasing interest due to the environmental pollution and high cost associated with synthetic indicator (Izonfuo *et al.*, 2006). The advantages of using natural dye from plant extract as acid – base indicator is that it is cheap, easily available, easy to prepare, simple to extract, nontoxic and environmentally friendly since it can easily degrade when released to the environment (Izonfuo *et al.*, 2006). The colour change in the titrimetric process may be attributed to the type of phytochemicals present in the plant extract (Akpakpan *et al.*, 2020). The major phytochemical responsible for the colour change of the natural indicator extracted from the plant source is anthocyanin which is a subgroup of flavonoids. It is a secondary metabolite that contributes to the purple, blue or red colours in fruits, leaves or flowers. Anthocyanin can react or interact with acids or bases, resulting in changes at its molecular structure and hence the exhibition of colour changes (Patil *et al.*, 2009). All the anthocyanin has been considered to be derivatives of 3,5,7-Trihydroxy flavylum chloride. Various anthocyanins and anthocyanidins differ in the nature, number and position of hydroxyl groups, methoxy groups and sugar residue on their structural moiety (Patil *et al.*, 2009).

The use of natural indicator from plant extract have been reported by many authors. Nwosu, (2004) reported the potentials of natural indicator from Hibiscus (red species), Bougainvillea and rose flowers. Industrial and analytical potentials of the plant extracts from the fruit of *Telfair occidentalis* have also been reported by Azundo, *et al.*, (2006). Utilization of Hibiscus flower extract, mango seed extracts, ginger stem extract and kola nut seed extract as natural indicators for acid – base titration has been reported (Uche *et al.*, 2014). Akpakpan *et al.*, (2020), reported

Comment [M2]: The word pollution appeared once in the whole manuscript. No data to support any pollution induced by acid-base indicators.

the acid-base indicator potentials of two varieties of kola nut extracts. Potentialsof *Dissotis rotundifolia*, *Centrosema pubescens* and *Allium cepa* extractsas indicators for acid-base titration was reported by Odiongenyi *et al.*, (2016).

However, there is no report on the utilization of aqueous and ethanolic extracts of *Dicliptera verticillata* leaves as acid base indicator. *Diclipteraverticillata* (Forssk). is a perennial herb, occurring in several parts of the world including Mauritania to Niger, Nigeria,India, Burma, Southern Africa, Zambia, Senegal, Sudan and Lake Chad (Telefo *et al.*, 1998). *Diclipteraverticillata* is a medicinal plant used for the treatment of malaria and diarrhoea in Burkina Faso. In India, the leaves are used in preparation of various soups. It is also used as flavouring and spicingagent (Telefo *et al.*, 1998). The phytochemical constituents of the ethanol and aqueous extracts of this plant leaveshave been reported by Akpakpan *et al.*, (2017). The plant contains several phytochemicals which can enhance colour changes during acid -base titration.

However, this present study is aimed at assessing the potentials of this plants extracts as green indicators for acid-base titration in order to ascertain their suitability or otherwise in replacing the synthetic indicators which are not environmentally friendly.

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2.0 MATERIALS AND METHODS

2.1 Samples Collection and preparation

Fresh samples of *Dicliptera verticillata*leaves were collected from a farm in Eket road, Mkpato Enin local Government, Akwa Ibom State, Nigeria. The samples were identified by a botanist in University of Uyo, Nigeria.

The samples were washed, squeezed and cut into pieces using knife. 20% extracts were prepared by soaking 20 g of the fresh sample separately in 100 ml of ethanol and hot water, after 24 hours, the extracts were filtered using Whatman filter paper and the filtrates obtained were characterizeusing UV/visible spectrometer, and was used as indicator for the titration to evaluate their indicator properties.



Figure 1: *Dicliptera verticillata* plant

2.2 Experimental Procedure

2.2.1 Characterization of *Dicliptera verticillata* extract using UV-VIS Spectroscopic analysis

UV/visible Spectroscopic analysis was conducted on the *Dicliptera verticillata* extract at room temperature. The extracts were diluted to 1: 10 (v/v) using the same solvent. The raw extracts, and the extracts in acidic and basic medium were analyzed using Genesys spectrophotometer in the wavelength ranging from 200-800nm, scan speed 5 nm/s and 1nm resolution in order to generate their characteristic absorption spectra.

2.2.2 Titrimetric analysis

The experimental work was done using the extracts of *Dicliptera verticillata* leaves. Another experiment was also carried out using the standard indicator (phenolphthalein and methyl orange) (Predeep and Dave, 2013). Exactly 25ml of each base (sodium hydroxide or ammonium hydroxide) was measured into a conical flask and 1.0ml of the *Dicliptera verticillata* leaves extract was added using a 1.0ml dropper. A 50ml burette was filled with either hydrochloric acid or ethanoic acid and titrated against each base (sodium hydroxide or ammonium hydroxide) until a sharp colour change was observed signifying the end of the reaction. Another titration was carried out using the standard indicator (phenolphthalein and methyl orange). Each titration sequence was done in triplicate using either the standard indicator or the plant extract and the end points recorded. The mean value was calculated for each set of titrations. The titrations were carried out for the strong acid vs strong base (HCl/NaOH), strong acid vs weak base (HCl/NH₄OH), weak acid vs strong base (CH₃COOH/ NaOH) and weak acid vs weak base (CH₃COOH/ NH₄OH). Titration was done using 0.5M acids and 0.5M bases, the end point values obtained are presented in Table 2.

2.3 Colour changes

The colour of the extracts and their colours in acidic and basic medium are presented in Table 3, and Figure 2 and 3.

4. 0 Results and Discussion

4.1 UV/visible spectroscopic analysis

UV/visible absorption spectra of ethanol and hot water extracts of *Declipitarea verticilata* leaves in different media is presented in Figure 1, and their wavelength of maximum absorption with corresponding absorbance are presented in Table 1.

Table 1: wavelength of maximum absorption and the corresponding absorbance of each extract

Samples	λ_{\max} (nm)	Absorbance
Ethanol extract (EE)	648 - 674	0.500
Ethanol extract in acidic medium (EEA)	650	0.202
Ethanol extract in basic medium (EEB)	648	0.215
Hot water extract (HWE)	558 - 600	0.500
Hot water extract in acidic medium (HWEA)	576	0.027
Hot water extract in basic medium (HWEB)	592	0.147

The variations in the absorbance and wavelength of maximum absorption of the extracts in different media is due to the presence of different forms of phytochemicals especially anthocyanin content in the plant extract. Absorption range of 230-270 nm indicate the transition electrons from lone pair and pi-electrons to antibonding sigma and pi bond in the phytochemicals present in the extracts. This absorption is consistent with the dominant group of hydroxyls, methoxy and benzene conjugated group in anthocyanin structures.

Comparing the wavelength of maximum absorption (λ_{\max}) of ethanol extract and hot water extract, the absorption maxima of ethanol extracts is higher than that of hot water extract, this may be due to the fact that ethanol extract contains both the polar and some non-polar phytochemicals from the plant. The λ_{\max} of ethanol extract ranges from 648 – 674 nm, at the absorbance of 0.5. The λ_{\max} of both ethanol and water extracts in acidic and basic medium were

within the ranges of that of the raw extracts, with corresponding decrease in absorbance. This is due to the dilution that takes place when these extracts were introduced into these media. This hypochromic effect was higher in acidic medium than basic medium.

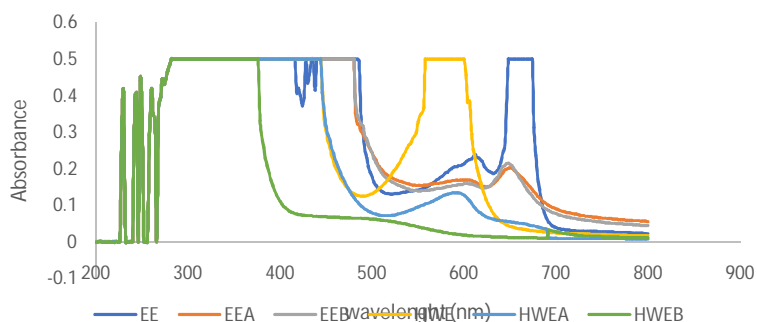


Figure 2: UV/visible absorption spectra of ethanol and hot water extracts of *Decliptarea verticilata* leaves in different media

Decliptarea verticilata leaves extract showed multiple absorption peak in visible region, 500 nm – 700 nm and similar absorption peaks in UV region, 225-270 nm. In the visible region, absorption band corresponding to the complementary colours were observed (Janssens, 2003). Whereas multiple peaks in UV region was attributed to the multiple transitions from the ground state to the excited state of the phytochemical functional groups present in the extracts (Vankar, and Bajpai, 2010).

4.2 Titrimetric analysis

The results of the titrimetric analysis using *Dicliptera verticillata* leaves extracts and commercial indicators are presented in Table 2.

Table 2: Titrimetric analysis using ethanol and hot water extracts of *Dicliptera verticillata* leaves and commercial indicators

Acid/base	End point (ethanol extract)	End point (hot water extract)	End point (Methyl orange)	End point (Phenolphthalein)
HCl/NH ₄ OH	No visible colour	No visible colour change	37.02 ± 0.01	34.06 ± 0.00

	change			
HCl/NaOH	28.00 ± 0.01	28.08 ± 0.01	28.00 ± 0.03	28.03 ± 0.03
CH ₃ COOH/NH ₄ OH	No colour change	33.02 ± 0.01	No visible colour change	No visible colour change
CH ₃ COOH/NaOH	30.00 ± 0.01	26.04 ± 0.01	32.40 ± 0.02	30.32 ± 0.01

In strong acid/ strong base titration, the average titre value of *Dicliptera verticillata* leaves extracts was 28.00 cm³ and 28.08 cm³ for ethanol and hot water extracts respectively, whereas methyl orange gives a titre value of 28.00 cm³ and phenolphthalein 28.03 cm³. These results indicate that *Dicliptera verticillata* leaves extract has end point comparable with standard indicators, hence it can serve as a suitable replacement for commercial indicator.

However hot water extract gave a remarkable colour changes for all titrations while ethanol extracts gave colour changes for the titration of strong acid with strong base and weak acid with strong base. Hence ethanol extract is not recommended for the titration of strong acid with weak base, and weak acid and weak base.

Comparatively, *Dicliptera verticillata* leaves extracts gave titre values less than or comparable to that of the commercial indicators for all titrations, hence titration using this plant extracts as indicator will consume small volume of acids to reach the end point.

The titrimetric equations of reactions which occur during the titration processes, in the presence of these indicators are presented in equation 1-4.

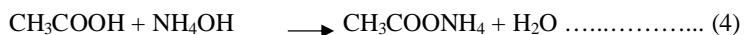
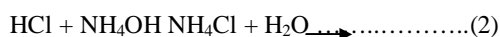
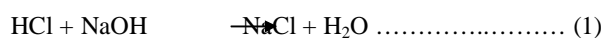


Table 3: Colour of the extracts in acidic and basic medium

Extracts	Colour of the extracts	Colour in Acid	Colour in Base
Hot water extract	Red	Light Red	Dirty green
Ethanol extract	Redish brown	Redish brown	green

Comment [M6]: There are no any evidence on the course for color change since no any chemical structures of the extract were established

Plant extract showed different colours in different media, this is because of the changes in the chemical structure and functional groups (Chromophore) of its phytochemicals in different media. (Abugri *et al.*, 2012). The plant extracts gave remarkable colour changes as shown in Figure 2 and 3.



Figure 2: Colour of the hot water extract in different medium

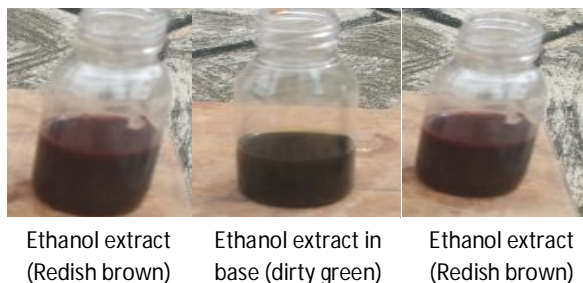


Figure 3: Colour of ethanol extract in different media

Conclusion

This study was carried out to evaluate the potentials of ethanol and hot water extracts of *Dicliptera verticillata* leaves as eco-friendly acid-based indicator to serve as an effective replacement for synthetic indicators. The UV/visible spectroscopic studies revealed that the absorbance and λ_{max} of the extract changed in different media due to the changes in the structural features of phytochemicals/chromophore present in the leaves extract. The results obtained for the titration using *Dicliptera verticillata* leaves extracts as indicator were comparable with that of synthetic commercial indicators. Hot water extract gives sharp colour changes than ethanol extract. Hence it is highly recommended that these extracts of *Dicliptera verticillata* leaves should be utilized as green indicator for acid-base titration especially at the

primary and secondary school levels of education since they are easily available, cheap, biodegradable and eco-friendly.

Comment [M7]: Geographical details of the plant are missing that makes not much convincing as an alternative

References

Abugri, D. Apea O. and Pritchett, G., Investigation of a Simple and Cheap Source of a Natural Indicator for Acid-Base Titration: Effects of System Conditions on Natural Indicators," *Green and Sustainable Chemistry*, 2, 117-122, 2012.

Akpakpan, A. E., Uwanta, E. J., Akpan, P. E. and Owen A. E. (2020). Phytochemical screening and analytical assessment of acid-base indicator properties of red and white kola nuts extracts. *Asian Journal of Advances in Research* 3(1): 1-7

Akpakpan, A. E., Ukpong, E. J., Odiongenyi A. O. and I. E. Willie (2017) Phytochemical Screening of the Ethanol and Aqueous Extracts of *Dicliptera verticillata* Leaves. *Archives of Current Research International* 11(2): 1-5, 2017

Azundo, M. U., Chukwunke, C. and Agbo, G. (2006). The dyeing of textile fabrics with natural dyes from some local tress. *J. Chem. Soc. Nig.* 23: 47-52.

Castaneda-ovando, A., Pacheco-hernández, M.D.L., Paezhernandez, M.E., Rodríguez, J.A. and Galán-vidal, C.A. (2009). Chemical studies of anthocyanins: A review. *Food Chem.* 113, 859-871.

Izonfuo, W. A. L., Fekaruhobo, G. K., Obomanu. F. G. and Daworiye. L. J. (2006): Acid-base indicator properties of dye from local plants: dyes from *Basella alba* (Indian spinach) and *Hibiscus. Sabdariffa* (zobo). *Journal of Applied Environmental Management* 10(1); 5-8

Janssens, K. (2003). Handbook of Spectroscopy.

Nwosu, F. O., Adekola, F.A., and Ihedioha, K. C. (2004). Simple Titrimetric Colour Indicators from some Natural Flower Petals. *Centrepoint (science edition)*. 12 (2):74-89.

Odiongenyi, A. O. Akpakpan, A.E. Enengedi I. S. and Ekpo E. I. (2016). Evaluation of Potentials of *Dissotis Rotundifolia*, *Centrosema Pubescens* and *Allium Cepa* extracts as indicator for Acid-Base titration. *International Journal of Advanced and Innovative Research* (5)3: 2278-7844.

Patil, S. B., Kondawar, M. S., Ghodke, D. S., Naikwade N. S and Magdum, C. S (2009). Use of flower extracts as an indicator in Acid-Base titrations. *Research J. Pharm. and Tech.* 2(2): 421 – 42

Pradeep, D. J. and K. Dave (2013). A Novel, inexpensive and less hazardous acid-base indicator. *Journal of Laboratory Chemical Education*, 1(2): 34-38

Telefo, P.B., Moindipa, P.E., Tihana, A. N, Tchouanguep, D.C., Mbiapo, F.T. (1998). Effects of an aqueous extract of Aloe buellneri, Justina insular, Hibiscus macranthus, Dicliptera verticillata some physiological and biochemical parameters of reproduction in immature female rats. *Journal of Ethanopharmacology*, 63:225-230.

Uche, I. Onwuachu, Uju A. Aboh and Godson Iwuoha (2014). Extraction, Characterization and Workability of Some Local Plant Dyes as Acid-Base Indicators. *International Journal of Research in Applied, Natural and social Sciences*. 2(7):1-6

Vankar P. S. and Bajpai, D.(2010) "Rose anthocyanins as acid base indicators," *EJEAFChe*, 9, 5: 875–884,