

Original Research Article  
**Screening of amaranth (*Amaranthus cruentus* L.)  
mutant lines for salinity tolerance**

**ABSTRACT**

**Aims:** The present study was carried out to access the salt tolerance level of nine amaranth (*Amaranthus cruentus*) mutant lines selected from Benin cultivar 'Locale' at young plants stage in comparison with the cultivar 'Locale' used as control.

**Study design:** The experiment was laid out as a Completely Randomized Design with three replications.

**Place and duration of study:** The experiment was carried out in a screening house at University of Abomey-Calavi, City of Abomey-Calavi, Republic of Benin from May to June, 2020.

**Methodology:** Three-weeks old plants of the nine stable mutant lines and the control cultivar 'Locale' were planted in pots containing a mixture of potting soil and sand. NaCl concentrations: 0; 100; 150 and 200 mM were given by irrigation once in two days Plant growth parameters were evaluated after two weeks.

**Results** Salt effect caused a reduction of young plant growth whatever the growth parameter considered with a significant disparity ( $p=0.001$ ) among the genotypes. Growth of the control cultivar, lines 1, 11 and 15 was most affected under salt stress whereas that of lines 18; 23 and 16 was least affected. A significant difference ( $p=0.01$  or  $p=0.001$ ) was observed among the salt tolerance index of genotypes. The highest salt tolerance index was observed in the line 23 followed by lines 18 and 2; and the lowest in line 15 followed by line 17, line 10 and the control cultivar.

**Conclusion:** Some variability was observed among lines for their salt resistance. Lines 23, 18 followed by line 2 appeared as the most salt resistant types whereas line 15, followed by lines 17, 10 and the control cultivar were the most salt sensitive. Thus, lines 23, 18 and 2 are promising for salt affected areas. Further studies are necessary to determine the physiological and biochemical mechanisms involved in the lines' salt resistance.

**Key words:** *Amaranthus cruentus*, cultivars discrimination, mutation induction, plant growth, NaCl.

## 1. INTRODUCTION

Salt stress is considered one of the main limiting factors affecting plant productivity worldwide and influences almost all aspects of plant biology [1]. Recent statistics show that global salt-affected soils were about 1069.3 Mha in 2016 [2] and predicted that 50% of the world's arable lands will become salt-affected by 2050s [3, 4]. Thus, increasing tolerance to salt stress in crop plants is necessary to increase the yield [5]. The effects of salinity on plant growth and yield are complex, and may result from a combination of toxic, nutritional, and osmotic factors [6]. Plant overall response to increasing NaCl dose appear to be species-specific [7]. In addition, within the same given species, a substantial variation in salt sensitivity may appear in cultivars or varieties as reported in several species of vegetable crops including amaranth [8, 9] chili [10] tomato [11] African eggplant [12] and tossa jute [13]. The genus *Amaranthus* included species cultivated as leaf vegetables, pseudocereals, and ornamental plants. Plants of this genus exhibit a high nutritive value but also a fascinating ability to adapt to diverse harsh environments [14]. As a tropical leafy vegetable, it is acquiring increasing importance as a potential subsidiary food crop for its excellent quality of protein and endogenous micronutrients content [15, 16]. Vegetable crops are predominantly cultivated in the south of Benin, in urban and suburban areas and in the valley of *Oueme* [17] but the area sown in amaranth in this country is not known precisely. In Benin Republic, the area affected by salinity has not been studied but it has been reported that the arable lands of the coastal areas where amaranth are mainly cultivated were affected by soil and irrigation water's salinity [18, 19]. It has been demonstrated that there is variability in relative salt resistance among *Amaranthus cruentus* cultivars at young plant stage and that the most appreciated cultivar either by farmers or consumers, named *Locale* was the most salt sensitive among five available cultivars [9]. Thus, it would be interesting to improve this cultivar with the purpose of enhancing its salt resistance. It has been reported that genetic improvement of *Amaranthus* crops can be achieved by various techniques, such as classical breeding, mutagenesis and biotechnological approaches. The availability of simple and efficient techniques for inducing genetic variation, such as the use of radiation for inducing mutation and selection for desired traits is an essential component of any plant breeding programme [20]. Mutagenesis is a simple and cost-effective technology [21, 22]. Induced mutations have been utilized to create genetic variability for the selection of mutant varieties with improved agronomic traits in several plant species [23, 24, 25, 26, 27]. Induced mutagenesis is well adapted and culturally

accepted local cultivars can produce small genetic changes which will affect critical agronomic traits [28]. Mutation technology could be used as a tool to create genetic variation in different amaranth genotypes with enhanced quality and quantity of grain or with improved drought tolerance [29, 18].

The objective of the present study was to identify salt resistant genotypes among nine mutant lines of *Amaranthus cruentus*

## 2. MATERIAL AND METHODS

### 2.1. Summary of mutation induction and lines stabilization processes

Seeds of cultivar 'Locale' of *Amaranthus cruentus* were irradiated by  $\gamma$  radiation dose 200 Gy at IAEA in Vienna (Austria). The irradiated seeds (M0 generation) were transferred to Benin. M0 seeds were germinated in tanks filled with potting moistened soil for two weeks giving the M1 plants. The well-developed plants were transplanted to field. Each plant was self-pollinated at maturity using « single-seed descent » technique [30] at the experimental site of the International Institute of Tropical Agriculture (IITA / Benin, (latitude: N 6° 25' 260" and longitude: E 2° 19' 682"; altitude: 15 meter above sea level) in the City of Abomey-Calavi (Republic of Benin) from November, 2018 to March, 2020. This city is located in the Gulf of Guinea which is characterized by a subequatorial bimodal climate with two dry seasons and two rainy seasons [31]. The annual rainfall varies between 1200 and 1500 mm/year and the temperature ranges from 24 to 30 °C [31]. At each self-pollinated generation, individual seedling was selected and grown in 3 rows on a 3 m long and 1.5 m wide plot. Five plants were used per line. Plants were spaced 50 cm apart within rows and 50 cm between rows with a total of 18 plants per plot. Among those 18 plants grown per plot, the five well developed and phenotypically similar plants based on plant height, number of branches and leaves production were identified. The selected plants when produced flower panicle the flowers were covered from appearance, against external pollen entry with envelope made up of butter paper. At seed maturity stage, seeds were harvested per plant per line and dried. From these seeds, seedlings were raised from one of the five selected plants per plot per line and transplanted into a new plot to raise the crop of next generation. Similarly crops were raised generation after generation and process was continued in field until generation M6. Nine amaranth mutant lines from generation M6 were raised in pots and analyzed for their relative salt tolerance status in this study with the cultivar 'Locale' used for

mutation induction and called control cultivar (CC). Plants of control cultivar were normal non mutated plants.

## 2.2. Experimental conditions and design

The experiment was carried out in a screening house at the University of Abomey-Calavi (Republic of Benin) in the city of Abomey-Calavi. The experiment was laid out in a Completely Randomized Design (CRD) with three replications (three pots) and three plants per replication (per pot) (9 plants per treatment). Pots (11.3 cm diameter and 14 cm) were filled with 3 kg mixture of potting soil and sandy loam soil (composition in table 1) 50:50. The experiment was conducted as described by [32]. Salt treatments consisted of plant irrigation every two days with solution of four NaCl concentrations: 0, 100, 150 or 200 mM NaCl (CAS n°7647-14-5). The experiment was evaluated after two weeks exposure to salt stress.

**Table 1** : Composition of the sandy loam soil used for plant culture

Year	Depth	Component						
		pH (H <sub>2</sub> O)	C (%)	N (%)	C/N	Organic matter (%)	K exch. (meq/100g)	P avail. (ppm)
2017	0-40 cm	5.74	0.58	0.05	8.14	0.79	0.18	64.25

(Data from the characterization of soils of the National Institute of Agricultural Research of Benin done by the laboratory of the Soil Sciences, Water and Environment in 2017).

## 2.3. Data collected

Plant height, leaf number, root length, shoot and root fresh and dry matters were measured at the end of treatment. For dry matter determination, fresh samples were transferred to an oven at 80 °C for 72 hours. Salt tolerance index (STI) was determined for each growth parameter for the ten genotypes using the modified formulae [33].

## 2.4. Statistical analysis

The data collected were processed using descriptive statistics utilizing an Excel spreadsheet and presented in the form of tables and graphs. The analysis of the main effects of salt stress was based

on the variance analysis. Means were compared utilizing Students, Newman and Keuls (SNK) test. Statistical analyses were performed using JMP Pro 12 software [34]. For all parameters, each value was presented in the form of mean  $\pm$  standard error with a reading of three independent samples per treatment.

### 3. RESULTS

#### 3.1. Overall reaction of the nine lines and the control cultivar with regard to salt stress

Salt stress reduced plant growth as the NaCl concentration increase (fig. 1) and this growth reduction was more accentuated in the control cultivar (fig. 1-A) than the mutant lines 2 (fig. 1-B) and 23 (fig. 1-C). The two-ways analysis of variance revealed a significant effect of salt stress ( $p=0.01$ ) for all the growth parameters taken into account, a significant difference among genotypes ( $p=0.01$ ) and a significant salt x lines (genotypes) interaction ( $p=0.01$ ) (table 2). With a significant interaction between both factors for all growth parameters considered, the effect of salt stress on growth should be studied by considering each genotype at a time.

**Figure 1:** Plant of control cultivar (A) mutant lines L2 (B) and L23 of *Amaranthus cruentus* under different NaCl concentrations

#### 3.2. Effect of NaCl on plant aerial part growth

##### 3.2.1. NaCl effect on plant height

Salt stress reduced plant height in lines and the control cultivar (fig. 2). NaCl induced a significant ( $p=0.001$ ) reduction of plant height in the control cultivar and mutant lines except lines 18

**Table 2.** Results of two-ways analysis of variance of growth of nine mutant lines of *Amaranthus cruentus* and the control cultivar after two weeks of culture under different NaCl concentrations

Growth parameters	Stress	genotype	Interaction (Stress x genotype)
PH	105.80***	28.38***	2.73***
LN	54.63**	6.59**	2.23**
SFM	138.08***	24.04***	3.24***
SDM	149.42***	42.04***	7.50***
RL	63.06***	4.75***	2.02**
RFM	29.53***	13.32***	1.99**
RDM	59.34***	12.49***	3.18***

\*\* : difference significant at  $p=.01$ ; \*\*\*: difference significant at  $p=.001$

PH: Plant Height

LN: Leaf Number

SFM: Shoot Fresh Mass

SDM: Shoot Dry Mass

RL: Root Length

RFM: Root Fresh Mass

RDM: Root Dry Mass

and 23. The reduction was significant from 100 mM NaCl for lines 10, 15 and 16 whereas it was significant from 150 mM for lines 1, 2, and 17 and the control cultivar. The plant height reduction was significant only at 200 mM NaCl for line 11. Thus, salt effect on plant height inhibition was more accentuated in lines 10, 15 and 16 than the other lines; lines 18 and 23 were the least affected, followed by line 11. In addition, line 2 produced maximum plant height ( cm) followed by line 17 ( cm) and line 23 ( cm) under 200 mM NaCl applied treatment. Moreover, plant height was significantly affected by NaCl for eight of the ten genotypes studied and salt effect was significant from the lowest NaCl concentration used (100 mM) for three of the ten genotypes.

**Figure 2.** Plant height of mutant lines of *Amaranthus cruentus* under different NaCl concentrations (n = 3; vertical bars are standard errors). Values within cultivar with same letters are not significantly different at p=.001.

### **3.2.2. NaCl effect on leaf number**

NaCl induced a significant (p=.001) reduction in leaf number in the control cultivar and mutant lines except lines 2, 16, 17 and 18 (fig. 3). The reduction was significant from 150 mM NaCl for lines 1, 11 and 15 whereas it was significant only at 200 mM NaCl for lines 10, 23 and the control cultivar. Thus, leaf number reduction under salt stress was more increased in lines 11 and 15 than the other lines; lines 2, 16, 17 and 18 were the least affected. . In addition, line 2 produced maximum leaf number ( ) followed by line 17 ( ) under 200 mM NaCl applied treatment Moreover, leaf number was significantly affected by NaCl for six of the ten genotypes studied and none of the ten genotypes was significantly affected at the lowest NaCl concentration used (100 mM).

**Figure 3:** Leaf number of mutant lines of *Amaranthus cruentus* under different NaCl concentrations (n = 3; vertical bars are standard errors). Values within cultivar with same letter are not significantly different at p=.001.

### **3.2.3. NaCl effect on shoot fresh mass**

NaCl induced a significant ( $p=0.001$ ) reduction of shoot fresh mass growth in the control cultivar and all mutant lines (fig. 4). The reduction was significant from 100 mM NaCl for lines 2, 10, 15 and 17 whereas it was significant from 200 mM for lines 11, 18 and 23. Shoot fresh mass reduction was significant at 150 mM NaCl for line 1; 16 and the control cultivar. Shoot fresh mass inhibition was more increased in lines 2; 10; 15 and 17 than the other lines; lines 11; 18 and 23 were the least affected. . In addition, line 2 produced maximum shoot fresh mass ( g) followed by line 17 ( g) and line 23 ( g) under 200 mM NaCl applied treatment Moreover, shoot fresh mass was significantly affected by NaCl for all the ten genotypes studied and salt effect was significant from the lowest NaCl concentration used (100 mM) for four of the ten genotypes.

**Figure 4:** Shoot Fresh Mass of mutant lines of *Amaranthus cruentus* cultivars under different NaCl concentrations ( $n = 3$ ; vertical bars are standard errors). Values within cultivar with same letter are not significantly different at  $p=0.001$ .

#### 3.2.4. NaCl effect on shoot dry mass

NaCl induced a significant ( $p=0.001$ ) reduction of shoot dry mass in the control cultivar and all mutant lines (fig. 5). The reduction was significant from 100 mM NaCl for lines 2; 10, 15, 16 and 17 whereas it was significant only at 200 mM NaCl for lines 1; 11; 18 and 23 and the control cultivar. These results showed that salt effect on shoot dry mass inhibition was more accrued in lines 2; 10, 15, 16 and 17 than the other lines and the control cultivar. In addition, line 2 produced maximum shoot dry mass ( g) followed by line 17 ( g) and line 23 ( g) under 200 mM NaCl applied treatment Moreover, shoot fresh mass was significantly affected by NaCl for all the ten genotypes studied and

salt effect was significant from the lowest NaCl concentration used (100 mM) for five of the ten genotypes.

**Figure 5:** Shoot Dry Mass of mutant lines of *Amaranthus cruentus* under different NaCl concentrations (n = 3; vertical bars are standard errors). Values within line with same letter are not significantly different at  $p=0.001$ .

### 3.3. Effect on NaCl on roots growth

NaCl induced a significant reduction ( $p=0.001$ ) of root length in the control cultivar and mutant lines except lines 2; 16 and 23 (table 3). The reduction was significant from 100 mM NaCl for line 18 whereas it was significant only at 200 mM NaCl for lines 10; 15 and 17. Root length reduction was significant from 150 mM NaCl for lines 1, 15, 17, 23 and the control cultivar. Thus, salt effect on root length inhibition was more accrued in lines 18 than the other lines; lines 2; 16 and 23, followed by lines 10; 15 and 17 were the least affected. Moreover, root length was significantly affected by NaCl for seven of the ten genotypes studied and salt effect was significant from the lowest NaCl concentration used (100 mM) for one of the ten genotypes.

For root fresh mass, NaCl induced a significant reduction ( $p=0.001$ ) of root fresh mass in the control cultivar and mutant lines except lines 10; 17; 18 and 23 (table 1). The reduction was significant from 100 mM NaCl for the control cultivar and line 2 whereas it was significant only at 200 mM NaCl for line 1 and 11. Root fresh mass reduction was significant from 150 mM NaCl for lines 15 and 16. The root fresh mass was significant affected only at 200 mM NaCl for line 1 and 11. Thus, salt effect on root fresh mass inhibition was more accrued in the control cultivar and line 2 than the other lines; lines 10; 17; 18 and 23, followed by lines 1 and 11 were the least affected. Moreover, root fresh mass was

significantly affected by NaCl for six of the ten genotypes studied and salt effect was significant from the lowest NaCl concentration used (100 mM) for two of the ten genotypes.

For root dry mass, NaCl induced a significant reduction ( $p=0.001$ ) in the control cultivar and mutant lines except for lines 16 and 18 (table 1). The reduction was significant from 100 mM NaCl for lines 11; 15 and 17 and the control cultivar, whereas it was significant only at 200 mM NaCl for lines 1 and 23. The root dry mass reduction was significant from 150 mM NaCl for lines 10. Thus, salt effect on root dry mass inhibition was more accrued in the control cultivar and lines 11; 15 and 17 than the other lines; lines 16 and 18, followed by 23 and 1 were the least affected. Moreover, root dry mass was significantly affected by NaCl for eight of the ten genotypes studied and salt effect was significant from the lowest NaCl concentration used (100 mM) for five of the ten genotypes.

In general, the growth reduction due to NaCl salt stress was not significant for four and three of the eight growth parameters studied respectively for lines 18; 23 and 16 whereas this reduction was significant for all the eight or seven growth parameters for lines 1; 11; 15; the control cultivar and line 10. These results showcased that lines 18; 23 and 16 were the least affected by salinity whereas lines 1; 11 and 15 were the most affected followed by line 16 and the control cultivar. Moreover, shoot fresh mass and shoot dry mass were significantly affected by NaCl either in the ten genotypes studied, or from the lowest NaCl concentration used (100 mM) for five or four of the ten genotypes. These two growth parameters appeared as the most suitable character to be considered as selection criteria for more appropriate selection in amaranth, followed by plant height and root dry mass.

Globally, results showed that the standard errors in the absence of NaCl were similar for the ten genotypes whatever the growth parameter considered except for line 18 (SDM) and lines 2; 11; 16; 17 and 23 (RL).

**Table 3:** Effect of different NaCl concentrations (0, 100; 150 and 200 mM) on root length (RL), fresh (RFM) and dry masses (RDM) of control cultivar and nine mutant lines of *Amaranthus cruentus* after two weeks of stress: Values are means  $\pm$ SE (n = 3).

		RL	RFM	RDM
CC	00 mM	9.13 $\pm$ 0.46a	0.14 $\pm$ 0.01a	0.02 $\pm$ 0.00a
	100 mM	7.66 $\pm$ 0.66ab	0.07 $\pm$ 0.01b	0.01 $\pm$ 0.00b
	150 mM	4.73 $\pm$ 0.34b	0.05 $\pm$ 0.01b	0.01 $\pm$ 0.00b
	200 mM	5.06 $\pm$ 1.23b	0.03 $\pm$ 0.00b	0.01 $\pm$ 0.00b
L1	00 mM	8.00 $\pm$ 0.57a	0.31 $\pm$ 0.12a	0.02 $\pm$ 0.00a
	100 mM	7.30 $\pm$ 1.04ab	0.27 $\pm$ 0.03a	0.01 $\pm$ 0.00ab
	150 mM	4.75 $\pm$ 0.27bc	0.21 $\pm$ 0.01ab	0.01 $\pm$ 0.00ab
	200 mM	4.00 $\pm$ 0.00c	0.04 $\pm$ 0.00b	0.00 $\pm$ 0.00b
L2	00 mM	9.16 $\pm$ 0.92a	0.60 $\pm$ 0.06a	0.05 $\pm$ 0.00a
	100 mM	8.20 $\pm$ 0.35a	0.37 $\pm$ 0.04b	0.03 $\pm$ 0.00b
	150 mM	6.56 $\pm$ 0.53a	0.29 $\pm$ 0.05b	0.02 $\pm$ 0.00b
	200 mM	9.33 $\pm$ 0.66a	0.25 $\pm$ 0.01b	0.02 $\pm$ 0.00b
L10	00 mM	8.75 $\pm$ 0.72a	0.40 $\pm$ 0.13a	0.04 $\pm$ 0.01a
	100 mM	6.75 $\pm$ 0.72ab	0.26 $\pm$ 0.05a	0.02 $\pm$ 0.00ab
	150 mM	6.60 $\pm$ 0.46ab	0.08 $\pm$ 0.01a	0.01 $\pm$ 0.00b
	200 mM	5.75 $\pm$ 0.05b	0.16 $\pm$ 0.02a	0.00 $\pm$ 0.00b
L11	00 mM	10.50 $\pm$ 1.44a	0.36 $\pm$ 0.04a	0.05 $\pm$ 0.00a
	100 mM	7.00 $\pm$ 0.00ab	0.35 $\pm$ 0.00a	0.03 $\pm$ 0.00b
	150 mM	4.90 $\pm$ 0.80b	0.24 $\pm$ 0.03a	0.02 $\pm$ 0.00bc
	200 mM	3.25 $\pm$ 1.38b	0.07 $\pm$ 0.01b	0.01 $\pm$ 0.00c
L15	00 mM	9.00 $\pm$ 0.00a	0.55 $\pm$ 0.06a	0.04 $\pm$ 0.00a
	100 mM	7.90 $\pm$ 0.00a	0.33 $\pm$ 0.00ab	0.01 $\pm$ 0.00b
	150 mM	6.33 $\pm$ 1.20ab	0.30 $\pm$ 0.03b	0.01 $\pm$ 0.00b
	200 mM	4.10 $\pm$ 0.05b	0.18 $\pm$ 0.07b	0.01 $\pm$ 0.00b
L16	00 mM	8.55 $\pm$ 0.99a	0.33 $\pm$ 0.02a	0.02 $\pm$ 0.00a
	100 mM	7.33 $\pm$ 1.01a	0.22 $\pm$ 0.03ab	0.02 $\pm$ 0.00a
	150 mM	5.70 $\pm$ 1.15a	0.18 $\pm$ 0.2b	0.01 $\pm$ 0.00a
	200 mM	4.53 $\pm$ 0.03a	0.15 $\pm$ 0.03b	0.01 $\pm$ 0.00a
L17	00 mM	9.33 $\pm$ 1.76a	0.63 $\pm$ 0.13a	0.06 $\pm$ 0.00a
	100 mM	6.53 $\pm$ 0.54ab	0.35 $\pm$ 0.04a	0.03 $\pm$ 0.00b
	150 mM	5.00 $\pm$ 0.00ab	0.27 $\pm$ 0.08a	0.02 $\pm$ 0.00b
	200 mM	4.40 $\pm$ 0.60b	0.34 $\pm$ 0.01a	0.02 $\pm$ 0.00b
L18	00 mM	10.00 $\pm$ 0.57a	0.25 $\pm$ 0.05a	0.02 $\pm$ 0.00a
	100 mM	7.00 $\pm$ 0.57b	0.22 $\pm$ 0.00a	0.03 $\pm$ 0.00a
	150 mM	6.66 $\pm$ 0.33b	0.26 $\pm$ 0.02a	0.02 $\pm$ 0.00a
	200 mM	3.50 $\pm$ 0.50c	0.21 $\pm$ 0.02a	0.01 $\pm$ 0.00a
L23	00 mM	6.66 $\pm$ 0.88a	0.46 $\pm$ 0.08a	0.04 $\pm$ 0.00a
	100 mM	5.46 $\pm$ 0.48a	0.43 $\pm$ 0.09a	0.03 $\pm$ 0.00ab
	150 mM	4.06 $\pm$ 0.53a	0.38 $\pm$ 0.06a	0.02 $\pm$ 0.00ab
	200 mM	4.66 $\pm$ 0.33a	0.21 $\pm$ 0.04a	0.01 $\pm$ 0.00b

Means with different letters within column for each line are significantly different ( $p=0.01$  or  $p=0.001$ ).

### 3.4. Salt tolerance index of lines

There is a significant difference ( $p=.01$  or  $p=.001$ ) among lines for their salt tolerance index for the seven growth parameters evaluated (table 3). For plant height, line 23 (0.87) and line 18 (0.82) showed the highest values whereas line 15 (0.58) followed by line 1 (0.65) and the control cultivar (0.63) presented the weakest values. The other lines showed intermediary values. For leaf number, lines 18 (1.03) and 23 (0.80) showed the highest value whereas line 15 (0.59) presented the weakest value. The other lines showed intermediary values. For shoot fresh mass, lines 18 (0.75) and 23 (0.67) showed the highest value whereas line 15 (0.30) presented the weakest value. The other lines showed intermediary values. For shoot dry mass, line 18 (1.02) followed by line 23 (0.82) showed the highest values whereas line 15 (0.28) followed by line 10 (0.41) presented the weakest values. The other lines showed intermediary values. For root length, line 2 (1.07) followed by line 23 (0.93) showed the highest values whereas lines 11 (0.49) followed by 17 (0.55) and 18 (0.57) presented the weakest values. The other lines showed intermediary values. For root fresh mass, lines 18 (0.76) and 23 (0.69) showed the highest values whereas lines 17 (0.31), 10 (0.33), the control cultivar (0.34), lines 16 (0.36) and 1 (0.36) presented the weakest ones. The other lines showed intermediary values. For root dry mass, lines 18 (1.10) followed by L23 (0.86) and L2 (0.73) showed the highest values whereas lines 11 (0.24), L15 (0.30) and L10 (0.33) presented the weakest values. The other lines showed intermediary values. Thus, based on the salt tolerance index, lines 23 and 18 showed the highest salt tolerance index for 7 and 6 growth parameters respectively. These two lines appeared as the most salt resistant, followed by line 2 which presented the highest STI for two parameters. In contrast, line 15 presented the weakest salt tolerance index for five growth parameters and appeared then as the most salt sensitive followed by the control cultivar and line 17 which presented the weakest STI for two parameters.

**Table 4.** Salt tolerance index of nine mutant lines of *Amaranthus cruentus* and the control cultivar after two weeks of stress: *Values are means  $\pm$ SE (n = 3).*

Means with different letters within column are significantly different ( $p=0.001$ )

Lines	Growth parameters						
	HP	LN	SFM	SDM	RL	RFM	RDM
CC	0.63 $\pm$ 0.05bc	0.77 $\pm$ 0.06ab	0.47 $\pm$ 0.07ab	0.61 $\pm$ 0.07bcd	0.63 $\pm$ 0.06bcd	0.34 $\pm$ 0.06c	0.40 $\pm$ 0.08bc
L1	0.65 $\pm$ 0.06bc	0.68 $\pm$ 0.09ab	0.43 $\pm$ 0.12ab	0.49 $\pm$ 0.13abc	0.76 $\pm$ 0.09abcd	0.36 $\pm$ 0.06c	0.44 $\pm$ 0.09bc
L2	0.74 $\pm$ 0.03abc	0.78 $\pm$ 0.05ab	0.47 $\pm$ 0.04ab	0.71 $\pm$ 0.07abc	1.07 $\pm$ 0.08a	0.49 $\pm$ 0.08ab	0.73 $\pm$ 0.11ab
L10	0.67 $\pm$ 0.07abc	0.67 $\pm$ 0.05ab	0.41 $\pm$ 0.05ab	0.41 $\pm$ 0.04cd	0.86 $\pm$ 0.04abc	0.33 $\pm$ 0.05c	0.33 $\pm$ 0.07c
L11	0.78 $\pm$ 0.06abc	0.71 $\pm$ 0.06ab	0.61 $\pm$ 0.10ab	0.75 $\pm$ 0.11abc	0.49 $\pm$ 0.07d	0.40 $\pm$ 0.05bc	0.24 $\pm$ 0.05c
L15	0.58 $\pm$ 0.02c	0.59 $\pm$ 0.05b	0.30 $\pm$ 0.03b	0.28 $\pm$ 0.02d	0.67 $\pm$ 0.07bcd	0.25 $\pm$ 0.07bc	0.30 $\pm$ 0.04c
L16	0.68 $\pm$ 0.04abc	0.78 $\pm$ 0.07ab	0.50 $\pm$ 0.07ab	0.51 $\pm$ 0.05bcd	0.66 $\pm$ 0.05bcd	0.36 $\pm$ 0.05c	0.59 $\pm$ 0.10bc
L17	0.73 $\pm$ 0.02abc	0.77 $\pm$ 0.05ab	0.49 $\pm$ 0.05ab	0.50 $\pm$ 0.02bcd	0.55 $\pm$ 0.05cd	0.31 $\pm$ 0.06c	0.40 $\pm$ 0.03bc
L18	0.82 $\pm$ 0.05a	1.03 $\pm$ 0.08a	0.75 $\pm$ 0.07a	1.02 $\pm$ 0.05a	0.57 $\pm$ 0.06cd	0.76 $\pm$ 0.24a	1.10 $\pm$ 0.29a
L23	0.87 $\pm$ 0.02a	0.80 $\pm$ 0.06a	0.67 $\pm$ 0.07a	0.82 $\pm$ 0.09ab	0.93 $\pm$ 0.09ab	0.69 $\pm$ 0.09a	0.86 $\pm$ 0.14ab
Prob>F	<b>0.0005</b>	<b>0.0046</b>	<b>0.0064</b>	<b>&lt;0.0001</b>	<b>&lt;0.0001</b>	<b>0.0007</b>	<b>&lt;0.0001</b>

#### 4. DISCUSSION

Salt stress caused an inhibition on plant root and shoot growth. Such observation is commonly reported in several amaranth genotypes including *Amaranthus cruentus* genotypes [35, 36, 37, 38, 39, 40, 41]. Our results also revealed that the growth parameters used were differently affected by salinity according to the mutant line indicating that the effect of salinity on mutant lines depend on the growth parameters taken into account as previously reported in several vegetable species including amaranth [9]. This statement suggests that difference among lines was genetically controlled. Globally, the standard errors in the absence of NaCl were similar for the ten genotypes whatever the growth parameter considered. This result indicates that the selected lines have good stability validating thus the selection method used.

Results showed that lines 18; 23 and 16 were the least affected by salinity whereas lines 1; 11 and 15 were the most affected followed by line 16 and the control cultivar. Moreover, lines 23 and 18 showed the highest salt tolerance index followed by line 2 whereas line 15 presented the weakest salt tolerance index followed by the control cultivar and line 17. These results revealed certain variability in the response to salt stress of the nine mutant lines. Moreover, some of the mutant lines were less

affected than the control cultivar with the highest salt tolerance index indicating that the induced mutation process used was efficient to produce useful variability related to response to salt stress within lines from a salt sensitive cultivar. Induced mutations have been utilized for creation of genetic variability for the selection of mutant varieties with improved agronomic traits [23, 24, 25, 26, 27]. Therefore it is logical to consider that the mutation technology used in this study created a genetic variation in *Amaranthus cruentus* usefull to generate lines with improved salinity tolerance as reported in *A. tricolor* for drought tolerance [42]. Combining plant growth and salt tolerance index, lines 23 and 18 followed by line 2 appeared as the most salt resistant whereas line 15, followed by line 17, line 10 and the control cultivar were the most salt sensitive. According to Gandonou et al. [43], the effect of salt stress on plants depends on three interacting components: i) dehydration of the cells in response to the low external water potential, ii) nutritional imbalance caused by the interference of saline ions with essential nutrients and iii) toxicity due to the high accumulation of  $\text{Na}^+$  and  $\text{Cl}^-$  in the cytoplasm. It would be important to evaluate which of these three components played a main role in growth reduction under salinity stress of the tested mutant lines.

## 5. CONCLUSION

This study showcased that there is variability in the response of the nine amaranth mutant lines to NaCl salt stress at young plant stage. We noticed lines with high salt tolerance from other with low salt tolerance. Among the nine mutant lines evaluated, lines 23 and 18 followed by line 2 appeared as the most salt tolerant whereas lines 15 followed by line 17, line 10 and control cultivar appeared as the most saline sensitive. Thus, lines 23, 18 and 2 are promising for salt affected areas and can be used as donors in amaranth breeding program for salt tolerance. Among the seven growth parameters studied, shoot fresh mass and shoot dry mass were the best characters to be considered as selection criteria for more appropriate and easy selection for salt tolerance in amaranth, followed by plant height.

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