

Original Research Article

Response of Freshwater macroinvertebrate communities to various anthropogenic stressors in Lolab Streams- a lotic system of the Indian Himalayan Region

Abstract

This study evaluated the response of freshwater macroinvertebrate communities to several human-induced stresses in a characteristic temperate region with amassed population and population-related pollution. With the help of macroinvertebrate species connected with physicochemical parameters and contaminants in dissolved fractions, we aimed to establish an efficient bioassay approach for evaluating the water quality in the lotic ecosystems of Lolab streams. From the mouth of streams, a rapid scanning method was utilised. We discovered considerable differences in physicochemical parameters along a longitudinal gradient, with the greatest values recorded in populated and largely agriculturally developed areas in the catchment. The macroinvertebrates displayed preferences for particular abiotic parameters, highlighting their potential utility in future research as dependable ecological indicators, molded by a synergistic mix of anthropogenic influences and land use intensity.

Keywords: Benthic, environmental stress, indicator, biomonitoring, contamination

Introduction

The increase in human population and the increased need for resources and land have had a significant negative influence on aquatic ecosystems recently. Because increased pollution concentrations put more strain on species and the ecological health of watersheds, river ecosystems are actually at risk (Stara et al., 2020; Blahova et al., 2020; Stara et al., 2019). Quantifying the response of macroinvertebrate communities to stressors is a widely used method for assessing the effect of several pollutants on river habitats (Pagano et al., 2020; Gharaei et al., 2020). Monitoring programmes typically include measurements of the physico-chemistry of water and sediment together with the response of macroinvertebrate populations (Bian et al., 2016). The bioassay investigations provide a supplementary thorough approach to chemical measurements of water alone (Strungaru et al., 2021). The regularly measured stressors in biomonitoring programs are divided into management-relevant stressor groups,

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such as organic pollution, eutrophication, and hydromorphological changes (Stara et al., 2020; Freitas et al., 2020; Banaee et al., 2021).

Consequently, biological, hydromorphological, chemical, and physicochemical investigations are utilized to determine the ecological state of rivers (Merola et al., 2022). Macroinvertebrates, fish, phytobenthic, and saprophytes are among the biological elements that are frequently examined in line with the Water Framework Directive (Marques et al., 2021). In contrast to the limit value technique, this framework offers specific activities to accomplish the common goal of high water quality status (Eder et al., 2021). Therefore, it is critical to assess how benthic macroinvertebrates respond to anthropogenic stresses in a lowland tributary river that flows through cities and agricultural areas and serves as the main receiver of wastewater from nearby wastewater treatment facilities. Major stressors are identified as organic matter load and chemicals, heavy metals, salts, and nitrogen compounds (Lu and Yu, 2018). Studies have shown that groups of benthic macroinvertebrate organisms react quickly to changes in the water's chemical balance (Aliko et al., 2019).

In this work, we looked at the effects of key contaminants on benthic macroinvertebrates in two temperate climate streams, as well as their reaction to water changes. According to the current research, it's possible to see how human-caused main pollutants and mixed land use (i.e. agricultural and urban areas) affect a river's longitudinal gradient from its headwaters to its mouth for the first time.

2. Material and Methods

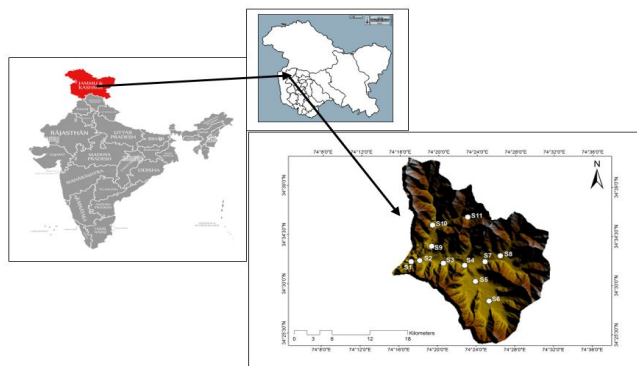
2.1 Study area

Jammu and Kashmir is home to the Himalayan sub-valley Lolab, where the Pir-Panjal mountains are found on all three sides. Some parts of the Lolab Valley have rocky and steep terrain. The Lahwal (Lalkul) river, which flows east to west and drains the Lolab Basin, is the source of water for the Lolab Basin. 34° 43' 30"N to 34°24'0" N latitudes and 74°15'0"E to 74°39'0" E longitudes are its latitude and longitude points. The research region's drainage area is 447 square miles. The Lolab's main axis extends westward for around 30 kilometers. One of the Lolab's lateral tributaries is the Kalaroos stream, which originates below the top of Nalgat (3645 m) and flows into the Lolab below Khumriyal. Morphologically, Lolab valley is flat to moderately sloping, at an elevation of around 1600 meters above mean sea level (AMSL). Temperatures typically range from -5 to 32 degrees Celsius at their coldest and

warmest points. The study area was divided into 11 sub-sites for this investigation. Fig. 1 shows the geographical map of the study area: Table 1 shows latitudes, longitudes, elevation

Table 1: Study sites with geographical coordinates

Site Code	Altitude	Latitude	Longitude
I- (S1)	1594m	34°32'14.53''N	74°17'21.36''E
II- (S2)	1647m	34°3331.4N	74°19'16.21''E
III-(S3)	1555m	34°538701''N	74°33'58.36''E
IV-(S4)	1666m	34°530754''N	74°37'83.57''E
V-(S5)	1675m	34°29'48 69''N	74°23'44.2''E
VI-(S6)	1692m	34°26'50 38''N	74°26'55.38''E
VII-(S7)	1716m	34°32'05.15''N	74°24'47.83''E
VIII-(S8)	1781m	34°31'17.03''N	74°28'11.87''E
IX-(S9)	1604m	34°33'31.4''N	74°19'16.21''E
X-(S10)	1792m	34°33'31.3''N	74°19'16.21''E
XI-(S11)	1799m	34°36'43.83''N	74°24'26.32''E



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Fig.1: Geographical map of Lolab watershed (Study Area)

2.2 Sampling Techniques

The eleven study locations were sampled every month from January 2018 to January 2019, from 9:00 am to 12:30 pm on each sample collection day.

2.2.1. Collection and analysis of Macroinvertebrates

Using D-nets that were 30 cm broad and 30 cm long and had a mesh size of 0.5 mm, macroinvertebrates were captured (Barbour et al., 1999; Ligeiro et al., 2020). The substrate's composition was considered when a semi-quantitative approach for hard-bottomed substrates was created. It is simple to collect samples because the natural flow of organisms in the streambed directs the sampling net upstream. The top layer of cobbles or pebbles was kicked off the bottom substrate for at least a minute in order to scrape the base bed (Malmqvist&Hoffsten, 2000; Ilmonen&Paasivirta, 2005). Using each stage of the approach, one square metre was sampled at 15-meter intervals along the stream's path. Another approach for locating invertebrates in the wild is the rock pick method. Crustaceans were extracted from the slimy river bottoms using an 8-inch square metal box with spring-opened and spring-closed jaws (15,215.2). To cover a 1 m² area, it was applied three to four times at each site. Invertebrates, which had fragile bodies, were preserved in 70% alcohol, while mollusks, which had hard shells, were preserved in 4.5% formalin. Macroinvertebrates were observed and identified using a binocular microscope (x 6 magnification) (Edmondson, 1959; Pennak, 1978; McCafferty & Provonsha, 1983; Ward, 1992; Engblom and Lingdell, 1999). Density (Shannon-Weiner, 1949), variety (Duan et al., 2008), taxonomic richness, dominance (Camargo et al., 1992), evenness (Pielou et al., 1969), and percentage densities of all macroinvertebrate orders are some of the estimated community properties of organisms.

2.2.2 Analysis of environmental variables

At each sampling site, we collected water samples, subsurface water temperatures, air temperatures, dissolved oxygen concentrations, and temperature-corrected electrical conductivity. During each sampling, pH, depth, and flow velocity were all monitored on-site. When it came to taking an accurate reading, a mercury-in-glass thermometer came in handy. Precision Scientific Instruments Corporation India's PCS Tester 35 Eutech Multi-Parameter Tester was utilized to obtain readings for DO, and pH. Water samples were collected in acid-washed plastic bottles and sent to the laboratory on ice to preserve their integrity during transportation to the lab. Within 24 hours of collection, nitrates were examined in the

laboratory, and phosphates were quantified spectrophotometrically after being reduced using appropriate solutions (APHA 2005).

Table 1 Physico Chemical parameters and methods followed

Parameter	Method
Air temperature	Mercury thermometer
Water temperature	Mercury thermometer
(pH)	PCS Tester 35
Dissolved oxygen	Iodometric method
Free CO ²	Titrimetric
Total hardness	EDTA titrimetric
Ammonical nitrogen	Spectrophotometry
Total phosphorus (TP)	Stannous chloride method

2.3. Data analysis

The mean and standard error were computed for each parameter and research site. A one-way ANOVA was used to compare the stations' physical and chemical characteristics. The Shapiro-Wilk and Levene's tests were used to evaluate the assumptions of normality and homogeneity of variance prior to the use of ANOVA. When these presumptions were shown to be false, data were log (x + 1)-transformed, except for pH, and then analyzed using repeated measure ANOVA with sample month as a subfactor. After significant ANOVAs, post hoc Tukey Honest (HSD) testing was employed to find differences across station means (P 0.05).

3. Results and Discussion

3.1 Environmental conditions, chemistry, and pollutants

During the study, air and water temperature were recorded monthly for a year. The minimum temperature was recorded during January 2019 at site II at 1.8°C and, the maximum temperature was recorded in August 2019 at site II at 30.5°C. Typically, summer had the highest temperature, while winter had the lowest (p = 0.0255). Water temperature remained

low throughout the year owing to the krial nature of streams. Like air temperature, water temperature showed the same regime, the lowest temperature in January was recorded was 1.4 °C, and the highest temperature recorded in July was 30.1°C. Seasonally, summer saw the greatest temperatures and winter saw the lowest ($p = 0.04237$).

pH was generally on the alkaline side, and values ranged from 7.6 at site VI in December to 8.67 at site I in March, reflecting limestone dissolution in the catchment. Seasonally spring and autumn showed high pH, while summer recorded the lowest pH. Annual mean pH ranged from a minimum of 7.6 ± 0.4 to a maximum of 8.6 ± 0.3 from upstream to downstream. The importance of pH as a controlling factor in aquatic systems has long been acknowledged, and pH extremes have a detrimental effect on biological components. The two metabolic processes that frequently have the greatest impact on pH in aquatic ecosystems are photosynthesis and respiration.

Conductivity is an excellent indicator of charged ion concentration in water, and landscape-scale circumstances heavily influence it. The catchment's ion geology creates the ions that function as electrical conductors (Ingham et al., 2010; Olson 2012). The majority of the dissolved ions in river water are caused by anthropogenic causes, other natural sources, and rock weathering (Berner and Berner, 1987). Conductivity showed spatial and temporal variations throughout the study period, ranging from a minimum of $66.23 \mu\text{Scm}^{-1}$ at Site V in May 2018 to a maximum of $327.0 \mu\text{Scm}^{-1}$ at Site XI in July 2018. The seasonal average values of conductivity revealed a definite trend recording the highest importance in winter and the lowest values in spring. However, the annual mean values of various study sites revealed that conductivity increased downstream from Site V ($66.0 \pm 20.0 \mu\text{Scm}^{-1}$).

Dissolved oxygen is the most critical water quality variable in aquatic ecosystems. During the study, the concentration of DO varied from 4.9 mgL^{-1} to 12.6 mgL^{-1} . The highest concentration of 12.6 mgL^{-1} was recorded at site VII in February, while sites V and I recorded the lowest concentration of 4.9 mgL^{-1} in July and August, respectively. The annual mean concentration of DO was found to be highest at site VII with a value $\pm 10.1 \text{ mgL}^{-1}$ while sites I and V recorded the lowest annual mean concentration with a value of 5.59 mgL^{-1} . DO showed a constant increase in values while approaching from human dominating areas site I towards wild areas like site VII. Seasonally, winter had the greatest record, followed by autumn and spring, while summer had the lowest. The annual recorded field values of DO from all sites were found statically significant ($p < 0.0025$).

The concentration of carbon dioxide in aquatic environments is an important feature that reflects both internal carbon dynamics and external biogeochemical processes in terrestrial ecosystems (Richey et al., 1990). It affects the concentration of carbonates, bicarbonates, pH, and overall hardness in water as input parameters for the buffer system. In the present study, the monthly free CO₂ was recorded monthly, and the values ranged from a minimum of 1.4 mgL⁻¹ at site IV and at site III in August 2019 and a maximum of 8.4 mgL⁻¹ at the site I and site II in June and July. The annual higher values of free CO₂ 5.4±2.3 mgL⁻¹ were found at site I, while the lowest level of CO₂ 3.2±1.5 mgL⁻¹ was found at site IV. When comparing the overall mean of free CO₂ values concerning seasons, spring and autumn are depicted as significantly lower than summer and winter. Subjected to statistical analysis, the Tukeys HSD test revealed that the monthly values of free CO₂ were not significantly different (p< 0.005), and the p-value for free CO₂ was found p=0.23.

Hardness is a significant consideration when using water for a variety of applications. Hardness may be caused by the natural accumulation of salts from soil and rocks. During the investigation, total minimum hardness was recorded as 40.6 mg L⁻¹ at Site III in September, while maximum total hardness was found at Site IV in July as 316.9 mg L⁻¹. The annual average maximum concentration of total hardness was 201.64 mg L⁻¹ at site VIII, while the minimum annual average concentration of total hardness was found at 53.34 mg L⁻¹ at site I. Seasonally summer season recorded the highest concentration of total hardness while the spring season depicted the lowest. The concentrations of total hardness at all the sites during the study were found statistically significant (p=0.007).

During the study, ammonical nitrogen concentration was found to vary from a minimum of 83.5 µgL⁻¹ at site II in November to a maximum of 865.3 µgL⁻¹ in July at site V. Further, the average annual concentration of ammonical nitrogen showed the highest concentration of 273µgL⁻¹ at site V while as Site I recorded the lowest concentration of 139.9 µgL⁻¹. The highest value at site V is due to the direct washout of the water from paddy fields into the canal. All the values from study sites were found statistically significant (p=0.04).

Owing to human-made inputs, nitrogen and phosphorus levels are frequently increased (Carpenter et al., 1998). Examples of significant anthropogenic nitrogen and phosphorus additions to streams include agricultural fertilisers, air deposition, nitrogen-fixing plants, and human and animal waste (Boyer et al., 2002). During the study, total phosphorus concentration was found to vary from a maximum of 885.3 µgL⁻¹ at site V in June

to a minimum of 78.6 μgL^{-1} in January at site VII. Further, the average annual concentration of total phosphorus showed the highest concentration of 286.33 μgL^{-1} at site V while Site I recorded the lowest concentration of 112.17 μgL^{-1} at site I. Seasonally summer recorded the highest concentration of nitrate-nitrogen while winter recorded the lowest. All the values from study sites were found statistically significant ($p=0.0010$).

Table 2: Linear (r) Pearson correlation between various Physico-chemical parameters during four seasons of the year 2018-2019 from Lolab Watershed.

	Air Temp	Water Temp	pH	Dissolved Oxygen	FREE CO2	Total Hardness	Nitrate Nitrogen	Total Phosphorus
Air Temp		0.009802	0.52199	0.14175	0.6841	0.66438	0.26863	0.39438
Water Temp	0.9902		0.47641	0.12484	0.76323	0.57609	0.36661	0.3672
pH	-0.47801	-0.52359		0.13979	0.42234	0.85453	0.95073	0.76621
Dissolved Oxygen	-0.85825	-0.87516	0.86021		0.45448	0.91781	0.58897	0.80196
FREE CO2	-0.3159	-0.23677	0.57766	0.54552		0.22351	0.57485	0.47197
Total Hardness	0.33562	0.42391	0.14547	-0.08219	0.77649		0.95829	0.12752
Nitrate Nitrogen	0.73137	0.63339	0.049266	-0.41103	-0.42515	-0.04172		0.55495
Total Phosphorus	0.60562	0.6328	0.23379	-0.19804	0.52803	0.87248	0.44505	

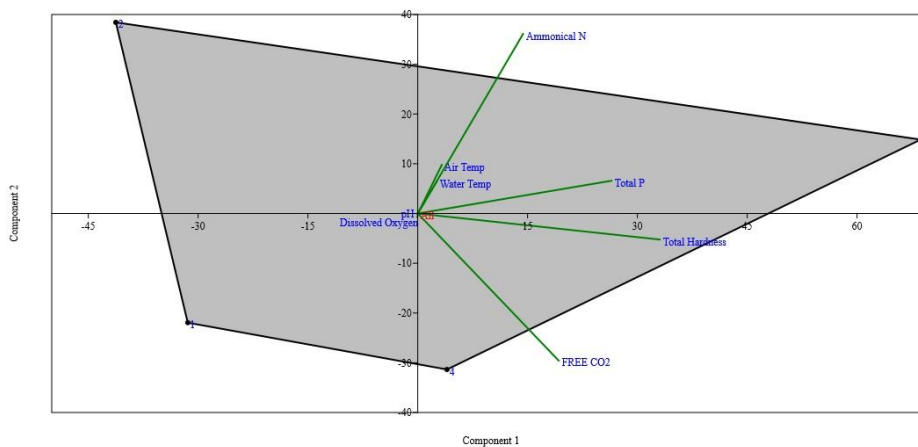


Fig 2: Principal Component Analysis (PCA) ordination plot explaining the variation of physicochemical parameters among sampling sites.

3.2 Species Investigated and diversity Indices

There are numerous ways in which macroinvertebrates can serve as accurate water quality indicators (Pacioglu and Moldovan, 2016). Natural environmental variation and anthropogenic stressors are likely to affect invertebrate communities in diverse river sectors in different ways (Caçador et al., 2012). Two general assumptions underlie the biotic indices: (1) that more stable assemblages have higher diversity values, whereas stressors lead to lower diversity, and (2) that diversity itself is a reasonable indication of environmental integrity because of the diversity it provides (Ravera, 2001; Elumalai et al., 2020). According to their stress tolerance and diversity values, taxa used as ecological indicators are graded when assessing the biological health of water bodies. Plecoptera, Ephemeroptera, Trichoptera, Decapoda, and Chironomidae are the taxonomic groupings utilized as indicators in the Extended Biotic Index, which uses pollution sensitivity as an indicator (Mozanzadeh et al., 2020). In comparison to their adults, mayflies, caddisflies, and dragonflies (including Odonata, Ephemeroptera, and Trichoptera) nymphs are more tolerant of pollution. The mayfly families Baetidae and Canidae, as well as other caddisfly families, have a wide range of pollution-resistant species. As pollution levels grow, tolerant organisms include non-biting midges, aquatic oligochaetes, and the isopod family, which includes the Tubifex sp. the biggest number of taxa (26) was observed at site VII due to the direct discharge of agricultural-rich effusive effluent, with a total of 27 taxa reported from all study sites. Trichoptera accounted for 36% of the total abundance studied, while Diptera accounted for 26%. Ephemeroptera accounts for 16% of all species. Percentage of Plecoptera and Coleoptera As a percentage, Araneae comprise 4%, and the rest 1%. Figure 3 depicts the CCA ordinations that link taxa's frequency to several environmental factors in aquatic environments. As a consequence of the CCAs, it was determined that there were five major

groups, each with distinct preferences for specific abiotic characteristics.

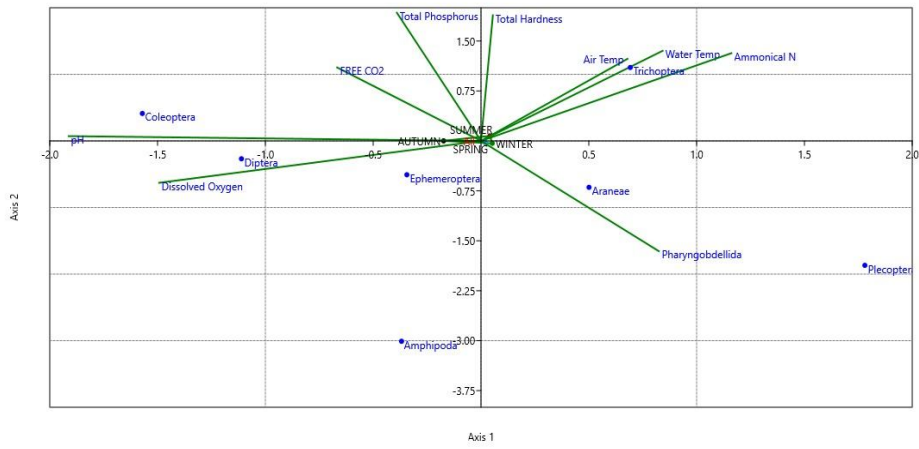


Fig 3: CCA ordinations related the frequency of taxa to environmental parameters in water

CCA was able to distinguish between the damaged and unaffected locations. The CCA ordination also revealed a substantial relationship between the macroinvertebrate fauna and environmental variables found in Lolab Streams. Site II had the highest nitrate concentration, pH, and hardness compared to the other sites, with site VI coming in second. A multiple-scale stressor could be recognised and described using a combination of factors. Numerous individual environmental variables have reasonably strong correlations with the axes for CCA, but these correlations were not statistically significant. These predicted significances, however, might be the outcome of unmeasured environmental variables. At the other stations, the bulk of the tolerant dipteran groups—*Biliocephala sp.*, *Chironomous sp.*, and *Culex sp.*—as well as the *Trichoptera*—*Brachycentrus sp.*, *Cheumatopsyche sp.*, and *Hydroptilidae sp.*—were either rare or absent. Site II was an extreme outlier in our ordination analysis that is a sign of the river's declining biotic and ecological health. In other investigations, Throughout the year of sampling, the species richness, diversity, and evenness indices at the various sampling sites seemed to match the water quality circumstances at each location. Low species diversity at site X indicated environmental stress as a result of gradually growing human influences on the water quality condition at these sites, but high species diversity at stations VIII and IX were associated with less contaminated circumstances. Various diversity indices are shown in table –

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Table 3: Various diversity indices

	SITE I	SITE II	SITE III	SITE IV	SITE V	SITE VI	SITE VII	SITE VIII	SITE IX	SITE X	SITE XI
Taxa_S	17	21	15	21	24	27	26	24	27	11	18
Individuals	256	228	220	136	238	520	670	458	244	92	168
Dominance_D	0.3119	0.3037	0.2743	0.2723	0.2737	0.2673	0.2694	0.2785	0.2821	0.2883	0.3044
Simpson_1-D	0.6881	0.6963	0.7257	0.7277	0.7263	0.7327	0.7306	0.7215	0.7179	0.7117	0.6956
Shannon_H	1.695	1.819	1.913	2.042	2.093	2.168	2.09	1.968	2.035	1.706	1.718
Evenness_e^H/S	0.3204	0.2936	0.4515	0.3671	0.3377	0.3237	0.311	0.2982	0.2833	0.5006	0.3098
Brillouin	1.593	1.685	1.798	1.835	1.936	2.072	2.017	1.879	1.876	1.542	1.577
Menhinick	1.063	1.391	1.011	1.801	1.556	1.184	1.004	1.121	1.728	1.147	1.389
Margalef	2.885	3.684	2.596	4.071	4.203	4.157	3.842	3.754	4.73	2.212	3.318
Equitability_J	0.5982	0.5974	0.7064	0.6708	0.6585	0.6578	0.6416	0.6193	0.6174	0.7114	0.5945
Fisher_alpha	4.095	5.639	3.643	6.943	6.66	6.046	5.38	5.388	7.76	3.259	5.11
Berger-Parker	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Chao-1	23	27	16	28.5	24	27	26.5	27	48	14	39

Conclusion [and recommendations](#)

Rivers in the North Western Himalayas are highly appreciated for their rich biodiversity, however during the past two decades, urbanization in the North India i.e., the Indian Himalayan Region has led to the disruption of freshwater ecosystems. The Lolab streams are not exempt from the influence of a growing human population on the water and biotic quality of the river, primarily as a result of waste disposal. This study provides information on the current state of the Lolab Streams' water quality and acts as a baseline survey of the river's macroinvertebrate species. This study's findings can serve as the basis for a long-term evaluation of the stream and the use of bioindicators in the management of the river system.

[Any Acknowledgement](#)

References

- Aliko, V., Mehmeti, E., Qirjo, M., & Faggio, C. (2019). "Drink and sleep like a fish": Goldfish as a behavior model to study pharmaceutical effects in freshwater ecosystems. *Journal of Biological Research-Bollettino Della Società Italiana Di Biologia Sperimentale*, 92(1).
- Banaee, M., Gholamhosseini, A., Sureda, A., Soltanian, S., Fereidouni, M. S., & Ibrahim, A. T. A. (2021). Effects of microplastic exposure on the blood biochemical parameters in the pond turtle (*Emys orbicularis*). *Environmental Science and Pollution Research*, 28(8), 9221–9234.
- Barbour, M. T. (1999). *Rapid bioassessment protocols for use in wadeable streams and rivers: Periphyton, benthic macroinvertebrates and fish*. US Environmental Protection Agency, Office of Water.
- Bian, B., Zhou, Y., & Fang, B. B. (2016). Distribution of heavy metals and benthic macroinvertebrates: Impacts from typical inflow river sediments in the Taihu Basin, China. *Ecological Indicators*, 69, 348–359.

- Blahova, J., Cocilovo, C., Plhalova, L., Svobodova, Z., & Faggio, C. (2020). Embryotoxicity of atrazine and its degradation products to early life stages of zebrafish (*Danio rerio*). *Environmental Toxicology and Pharmacology*, *77*, 103370.
- Caçador, I., Costa, J., Duarte, B., Silva, G., Medeiros, J., Azeda, C., Castro, N., Freitas, J., Pedro, S., & Almeida, P. R. (2012). Macroinvertebrates and fishes as biomonitors of heavy metal concentration in the Seixal Bay (Tagus estuary): Which species perform better? *Ecological Indicators*, *19*, 184–190.
- Eder, M. L., Oliva-Teles, L., Pinto, R., Carvalho, A. P., Almeida, C. M. R., Hornek-Gausterer, R., & Guimaraes, L. (2021). Microplastics as a vehicle of exposure to chemical contamination in freshwater systems: Current research status and way forward. *Journal of Hazardous Materials*, *417*, 125980.
- Edmondson, C. H. (1959). *Hawaiian Grapsidae*. The Museum.
- Elumalai, P., Kurian, A., Lakshmi, S., Faggio, C., Esteban, M. A., & Ringø, E. (2020). Herbal immunomodulators in aquaculture. *Reviews in Fisheries Science & Aquaculture*, *29*(1), 33–57.
- Freitas, R., Silvestro, S., Coppola, F., Costa, S., Meucci, V., Battaglia, F., Intorre, L., Soares, A. M., Pretti, C., & Faggio, C. (2020). Toxic impacts induced by Sodium lauryl sulfate in *Mytilus galloprovincialis*. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, *242*, 110656.
- Gharaei, A., Karimi, M., Mirdar, J., Miri, M., & Faggio, C. (2020). Population growth of *Brachionus calyciflorus* affected by deltamethrin and imidacloprid insecticides. *Iranian Journal of Fisheries Sciences*, *19*(2), 588–601.
- Hoffsten, P.-O., & Malmqvist, B. (2000). The macroinvertebrate fauna and hydrogeology of springs in central Sweden. *Hydrobiologia*, *436*(1), 91–104.

- Hoseinifar, S. H., Shakouri, M., Yousefi, S., Van Doan, H., Shafiei, S., Yousefi, M., Mazandarani, M., Mozanzadeh, M. T., Tulino, M. G., & Faggio, C. (2020). Humoral and skin mucosal immune parameters, intestinal immune related genes expression and antioxidant defense in rainbow trout (*Oncorhynchus mykiss*) fed olive (*Olea europea* L.) waste. *Fish & Shellfish Immunology*, *100*, 171–178.
- Ilmonen, J., & Paasivirta, L. (2005). Benthic macrocrustacean and insect assemblages in relation to spring habitat characteristics: Patterns in abundance and diversity. *Hydrobiologia*, *533*(1), 99–113.
- Ligeiro, R., Hughes, R. M., Kaufmann, P. R., Heino, J., Melo, A. S., & Callisto, M. (2020). Choice of field and laboratory methods affects the detection of anthropogenic disturbances using stream macroinvertebrate assemblages. *Ecological Indicators*, *115*, 106382.
- Lindell, K., Engblom, J., Engström, S., Jonströmer, M., & Carlsson, A. (1998). Influence of a charged phospholipid on the release pattern of timolol maleate from cubic liquid crystalline phases. In *The Colloid Science of Lipids* (pp. 111–118). Springer.
- Marques, M. B. L., Brunetti, I. A., Faleiros, C. A., da Cruz, C., Iqbal, H., Bilal, M., & Américo-Pinheiro, J. H. P. (2021). Ecotoxicological assessment and environmental risk of the insecticide chlorpyrifos for aquatic neotropical indicators. *Water, Air, & Soil Pollution*, *232*(10), 1–14.
- Pacioglu, O., & Moldovan, O. T. (2016). Response of invertebrates from the hyporheic zone of chalk rivers to eutrophication and land use. *Environmental Science and Pollution Research*, *23*(5), 4729–4740.
- Pagano, M., Stara, A., Aliko, V., & Faggio, C. (2020). Impact of neonicotinoids to aquatic invertebrates—In vitro studies on *Mytilus galloprovincialis*: A review. *Journal of Marine Science and Engineering*, *8*(10), 801.
- Pennak, R. W. (1978). *The dilemma of stream classification*.

- Provonsha, A., & McCafferty, W. (2006). A second species of the North American mayfly genus *Amercaenis* Provonsha and McCafferty (Ephemeroptera: Caenidae). *Journal of Insect Science*, 6(1).
- Ravera, O. (2001). Monitoring of the aquatic environment by species accumulator of pollutants: A review. *Journal of Limnology*, 60(1s), 63–78.
- Stara, A., Kubec, J., Zuskova, E., Buric, M., Faggio, C., Kouba, A., & Velisek, J. (2019). Effects of S-metolachlor and its degradation product metolachlor OA on marbled crayfish (*Procambarus virginalis*). *Chemosphere*, 224, 616–625.
- Stara, A., Pagano, M., Capillo, G., Fabrello, J., Sandova, M., Vazzana, I., Zuskova, E., Velisek, J., Matozzo, V., & Faggio, C. (2020). Assessing the effects of neonicotinoid insecticide on the bivalve mollusc *Mytilus galloprovincialis*. *Science of the Total Environment*, 700, 134914.
- Tian, H., Lu, C., Pan, S., Yang, J., Miao, R., Ren, W., Yu, Q., Fu, B., Jin, F.-F., & Lu, Y. (2018). Optimizing resource use efficiencies in the food–energy–water nexus for sustainable agriculture: From conceptual model to decision support system. *Current Opinion in Environmental Sustainability*, 33, 104–113.